

**UNIVERSIDADE FEDERAL DE CIÊNCIAS DA SAÚDE DE
PORTO ALEGRE – UFCSPA
PROGRAMA DE PÓS-GRADUAÇÃO EM PATOLOGIA**

Munique Mendonça Azevedo

**Avaliação de Marcadores de
Prognóstico no Melanoma**

UFCSPA

**Universidade Federal de Ciências da Saúde
de Porto Alegre**

**Porto Alegre
2017**

Munique Mendonça Azevedo

Avaliação de Marcadores de Prognóstico no Melanoma

Tese submetida ao Programa de Pós-Graduação em Patologia da Fundação Universidade Federal de Ciências da Saúde de Porto Alegre como requisito para a obtenção do grau de Doutor

Orientador: Dra. Claudia Giuliano Bica

**Porto Alegre
2017**

*Aos meus queridos pais,
Viviane e Wanderley, minha
inspiração.*

*Aos meus anjos de quatro
patas, meu refúgio.*

*Ao meu marido, Eduardo,
com muito amor*

AGRADECIMENTOS

Passados anos de tanta dedicação e empenho, neste momento, gostaria de agradecer:

À minha família: mãe, Viviane Pereira; pai, Wanderley Mendonça; avó, Ilma Pereira; tio, Eraldo Júnior. Motivo de todo meu esforço para buscar sempre o crescimento pessoal e profissional;

Ao meu marido, Eduardo Azevedo, pela paciência, ajuda e, sobretudo, pela motivação, acreditando em mim quando muitas vezes tive dúvidas;

Aos meus colegas de laboratório e alunos de iniciação científica, pelos momentos de descontração e pelo auxílio na execução deste estudo. Sem vocês, este caminho teria sido muito mais difícil;

Ao Dr. Felice Ricardi, primeiramente por ter me apresentado à minha orientadora e, pela oportunidade de acompanhá-lo no Núcleo de Melanoma. A experiência foi fundamental na minha formação profissional;

À Prof. Dra. Ivana Cruz, pelas horas de dedicação na análise dos dados, além dos ensinamentos;

Aos pacientes, que gentilmente aceitaram participar do estudo e sempre se mostraram muito interessados em cada etapa do processo;

À minha orientadora Prof. Dra. Claudia Bica, que mais do que orientar esta tese foi amiga e parceira. Muito obrigada!

"Nothing in life is to be feared, it is only to be understood. Now is the time to understand more, so that we may fear less".

— Marie Curie

SUMÁRIO

Lista de abreviaturas	VII
Lista de figuras	IX
Lista de tabelas	X
Resumo	XI
1. Introdução	13
1.1 Melanoma cutâneo	13
1.2 Epidemiologia	18
1.3 Metástases	20
1.4 Biomarcadores no prognóstico do melanoma	23
1.5 <i>Cell-free</i> DNA e inflamação	26
1.5.1 Liberação de DNA na corrente sanguínea	26
1.5.2 O papel da inflamação no câncer	28
1.6 Polimorfismo da enzima MnSOD	31
1.6.1 Espécies reativas de oxigênio e o estresse oxidativo	31
1.6.2 Estresse oxidativo e o melanoma	34
1.6.3 Detoxificação enzimática de espécies reativas de oxigênio ...	35
1.7 Justificativa	39
1.8 Referências bibliográficas	41
2. Objetivos	65
2.1 Objetivo geral	65
2.2 Objetivos específicos	65
3. Artigos científicos	67

3.1 Artigo científico I	67
3.2 Artigo científico II	95
3.3 Artigo científico III	119
4. Considerações finais	139
5. Anexos	143
5.1 Parecer do Comitê de Ética em Pesquisa da UFCSPA	144
5.2 Parecer do Comitê de Ética em Pesquisa da ISCMPA versão 3	147
5.3 Parecer do Comitê de Ética em Pesquisa da ISCMPA versão 4	151
5.4 Comprovante de submissão de artigo à revista Pathology & Oncology Research	154
5.5 Normas da revista Pathology & Oncology Research	155
5.6 Comprovante de submissão de artigo à revista Biomarkers	169
5.7 Normas da revista Biomarkers	170
5.8 Comprovante de submissão de artigo à revista The American Journal of Dermatopathology	177
5.9 Normas da revista The American Journal of Dermatopathology	178

LISTA DE ABREVIATURAS

AJCC: *American Joint Committee on Cancer*

AL: acral lentiginoso

Ala: alanina

A-MnSOD: alelo variante Ala do polimorfismo da MnSOD

CAT: catalases

CEP: comitê de ética em pesquisa

CuZnSOD: superóxido dismutase dependente de cobre e zinco

cfDNA: *cell-free* DNA (DNA livre circulante)

DS: disseminativo superficial

EMT: transição epitélio-mesenquimal

EROs: espécies reativas de oxigênio

FDA: *Food and Drug Administration*

GSH: glutationa

GPX: glutationa peroxidase

IFN: interferon

IL: interleucina

INCA: Instituto Nacional do Câncer

ISCMPA: Irmandade Santa Casa de Misericórdia de Porto Alegre

LDH: lactato desidrogenase

LM: lentigo maligno

MM: melanoma maligno

MnSOD: superóxido dismutase dependente de manganês

MTS: *mitochondrial targeting sequence* (sequência de direcionamento mitocondrial)

N: nodular

PRX: peroxirredoxinas

RGP: *radial growth phase* (fase de crescimento radial)

SNP: *single nucleotide polymorphism* (polimorfismo de nucleotídeo único)

SOD: superóxido dismutase

SOD2: superóxido dismutase 2

Th1: células T *helper* 1

Th2: Th1: células T *helper* 2

Tis: tumor *in situ*

TNF: fator de necrose tumoral

TNM: tumor, linfonodos regionais, metástases à distância

UFCSPA: Universidade Federal de Ciências da Saúde de Porto Alegre

UV: ultravioleta

Val: valina

V-MnSOD: alelo variante Val do polimorfismo da MnSOD

VGP: *vertical growth phase* (fase de crescimento vertical)

LISTA DE FIGURAS

Figura 1: Ilustração demonstrando o critério ABCDE na detecção do melanoma14

Figura 2: Subtipos histológicos do melanoma cutâneo. DS, disseminativo superficial; N, nodular; LM, lentigo maligno; AL, acral lentiginoso15

Figura 3: Produção de EROs na mitocôndria. NADH coleta elétrons de moléculas de oxigênio (O_2). Os elétrons fluem através da cadeia mitocondrial de transportes de elétrons para síntese de ATP. Uma pequena quantidade de O_2 sofre uma redução incompleta dando origem ao ânion superóxido ($O_2^{\cdot-}$), a primeira espécie reativa de oxigênio dos organismos aeróbicos. Posteriormente, a enzima superóxido dismutase dependente de manganês (MnSOD) dismuta $O_2^{\cdot-}$ em peróxido de hidrogênio (H_2O_2) e O_2 ; H_2O_2 é ainda neutralizado pela glutathione peroxidase (GPX) em H_2O com glutathione (GSH) como substrato. Igualmente, H_2O_2 pode ser convertido no radical hidroxila ($\cdot OH$) através da reação de Fenton com ferro (Fe^{2+})32

Figura 4: Conteúdo de MnSOD na mitocôndria de acordo com os diferentes precursores originados do polimorfismo da enzima: alelo variante Ala e alelo variante Val. Enquanto que o precursor da MnSOD Ala-variante (MTS-Ala) é corretamente transportado através das duas membranas mitocôndrias, o precursor Val-variante (MTS-Val) é parcialmente contido na membrana interna da mitocôndria. O aumento do precursor da MnSOD contendo o alelo Ala é traduzido em uma detoxificação mais eficiente do superóxido, enquanto que o precursor Val permite o acúmulo do ERRO na mitocôndria38

LISTA DE TABELAS

Tabela 1: Classificação TNM para melanoma16

Tabela 2: Estadiamento do melanoma com base na classificação TNM17

RESUMO DA TESE

Introdução:

O melanoma é altamente metastático, representando a causa mais comum de morte por câncer de pele em escala global. Sua classificação e prognóstico são definidos pelo sistema de estadiamento da AJCC, o qual não é suficiente para acessar o risco de mortalidade para cada paciente individualmente devido ao caráter heterogêneo da doença, o que garante a busca por novos biomarcadores.

Objetivos:

Avaliar o estado-da-arte no que diz respeito à biomarcadores de prognóstico no melanoma, bem como analisar a associação entre a neoplasia e os seguintes marcadores: DNA livre circulante (*cfDNA*), citocinas inflamatórias e polimorfismo Ala16Val do gene SOD2 que codifica a enzima antioxidante superóxido dismutase dependente de manganês (MnSOD).

Material e Métodos:

Primeiramente, foi realizada uma revisão sistemática da literatura no que se refere a marcadores genéticos e epigenéticos com valor prognóstico no melanoma. Também, foram realizados dois estudos caso-controle onde foram incluídos pacientes com melanoma do Hospital Santa Rita e indivíduos saudáveis. No primeiro, foi realizada a quantificação dos níveis de *cfDNA* no plasma de casos e controles por meio do Kit Quant-iT™ PicoGreen® dsDNA (Invitrogen) e posterior leitura da fluorescência em fluorômetro. Para avaliar o *status* inflamatório dos indivíduos, a concentração de citocinas inflamatórias (IL-1, IL-6, TNF α , INF γ and IL-10) foi medida. No segundo estudo, foi avaliada

a associação entre o melanoma e o polimorfismo Ala16Val do gene SOD2 através da genotipagem de pacientes com melanoma e indivíduos saudáveis. Para tanto, um fragmento do gene SOD2 contendo a região do polimorfismo foi amplificado com *primers* específicos seguido de clivagem com a enzima de restrição *Hae III*.

Resultados:

No artigo científico I, o texto completo de 25 artigos foi avaliado. A análise do perfil de microRNA e a expressão de mRNA associada ao status mutacional dos genes BRAF e NRAS foram as abordagens mais frequentes. No artigo II, níveis elevados de *cfDNA* e citocinas pró-inflamatórias foram observados nos pacientes com melanoma quando comparados aos controles. Por outro lado, os casos apresentaram níveis reduzidos da citocina anti-inflamatória interleucina 10 em relação aos indivíduos saudáveis. Após a excisão do tumor primário, houve redução significativa da concentração de *cfDNA* no sangue dos pacientes. Quanto ao artigo III, foi reportada a associação entre o polimorfismo da MnSOD e o risco de melanoma, a qual é influenciada por sexo e idade.

Conclusão:

Apesar de muito conhecimento ter sido acumulado no que se refere aos marcadores no melanoma, nenhum se mostrou eficiente para detectar o potencial de progressão da doença ainda em seus estágios iniciais. A implementação de biomarcadores sanguíneos pouco invasivos durante o acompanhamento clínico para identificar pacientes em alto risco de metástases é fundamental para melhorar o desfecho dos pacientes com melanoma.

1. INTRODUÇÃO

1.1 Melanoma Cutâneo

O melanoma é o tipo de câncer de pele com origem nos melanócitos e é bem definido histopatologicamente (Chin *et al.*, 1998). Esta doença pode se desenvolver tanto a partir de nevos pré-existentes (20 - 40% dos casos) quanto pode iniciar *de novo*, ou seja, não associado a nevos prévios (60 – 80% dos casos) (Eggermont *et al.*, 2014).

A etiologia do melanoma maligno (MM) é multifatorial. Entre os fatores etiológicos e predisponentes associados a esta neoplasia estão a exposição ao sol e a susceptibilidade genética, os quais podem explicar o aumento da incidência deste câncer na população mundial (Lens e Dawes, 2004).

A identificação visual do MM é acessível, uma vez que se desenvolve na superfície da pele. Entretanto, o desafio está na variabilidade das formas que as lesões podem apresentar durante as primeiras fases da doença, o que retarda o início do tratamento (Friedman *et al.*, 1985). Nos estágios iniciais, os quais apresentam maiores chances de cura, as lesões malignas podem mimetizar histologicamente as benignas. Nos anos 80, foi introduzido o critério ABCDE na prática clínica a fim de reconhecer as lesões de melanoma. Esse critério é baseado em características morfológicas do tumor, onde: **A** = assimetria; **B** = bordas irregulares; **C** = cor; **D** = diâmetro (> 6 mm); **E** = evolução (Herman, 2012) (figura 1). No entanto, o diagnóstico baseado neste

método tem sensibilidade de apenas 65 – 80%, além de exigir que muitas excisões desnecessárias sejam conduzidas uma vez que lesões benignas podem mimetizar melanomas na avaliação clínica (Grin *et al.*, 1990). Desta forma, a dermatoscopia – uma técnica diagnóstica não invasiva para observação de lesões pigmentadas *in vivo* – é capaz de melhorar a acurácia diagnóstica através da caracterização dos aspectos morfológicos dos tumores cutâneos (Rigel, 1997). Isso porque este método permite a visualização de estruturas pigmentares presentes na epiderme, junção dermoepidérmica, derme superior e vasos do plexo superior através de um sistema de lentes e filtros de aumento associados a uma fonte de luz (Pehamberger *et al.*, 1987).

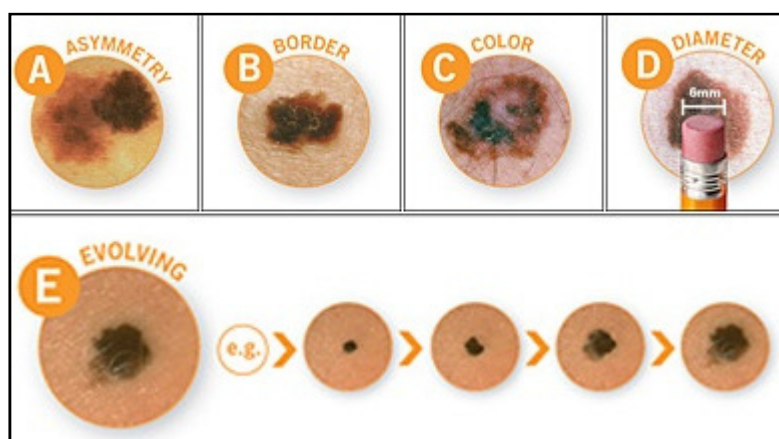


Figura 1. Ilustração demonstrando o critério ABCDE na detecção do melanoma. Adaptado de (Tsao *et al.*, 2015).

O melanoma cutâneo é classificado em quatro subtipos histológicos: disseminativo superficial (DS), lentigo maligno (LM), nodular (N) e acral lentiginoso (AL) (Clark *et al.*, 1969; Phan *et al.*, 2007) (figura 2). De acordo com a literatura, o subtipo disseminativo superficial é o mais freqüente (até 70 % dos casos de melanoma) e caracteriza-se por uma fase de crescimento radial

predominante (Clark *et al.*, 1986). O melanoma lentigo maligno surge com maior frequência na face e cabeça, ou seja, áreas mais expostas ao sol (Langley e Sober, 1998). O segundo subtipo mais freqüente é o nodular. Clinicamente apresenta-se como pápula ou nódulo, os quais podem apresentar coloração escura ou amelanótica (Mcgovern *et al.*, 1973). Por último, o melanoma acral lentiginoso é aquele que acomete as palmas das mãos, as plantas dos pés e as unhas.

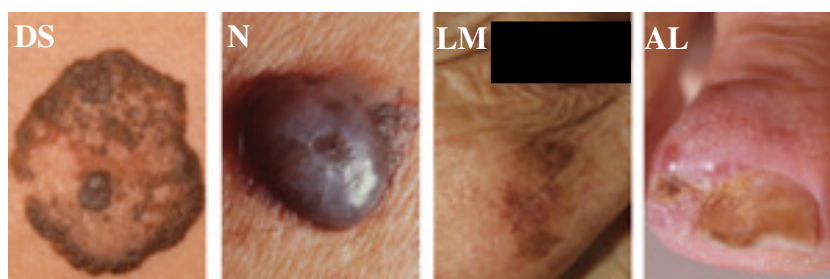


Figura 2. Subtipos histológicos do melanoma cutâneo. DS, disseminativo superficial; N, nodular; LM, lentigo maligno; AL, acral lentiginoso (imagem retirada e adaptada de http://www.jurmo.ch/work_heterogeneity.php, acessado em 26 de Novembro de 2015).

Os melanomas cutâneos passam por duas fases: fase de crescimento radial (RGP) e fase de crescimento vertical (VGP). Na primeira, o tumor pode invadir a epiderme e a derme papilar, mas a capacidade metastática está ausente. Durante a fase subsequente, ocorre a mudança para um estágio mais agressivo, onde metástases podem ocorrer. A transição entre a RGP e a VGP é marcada por alterações nas expressões gênicas, as quais são de grande interesse para estudos sobre a progressão da malignidade (Gallagher e Bergin, 2005).

Os principais fatores prognósticos do MM, segundo a classificação da *American Joint Committee on Cancer* (AJCC) 2009 (Balch *et al.*, 2009), são baseados na classificação padrão TNM (tumor, linfonodos regionais, metástases à distância). A categoria T descreve o estado do tumor primário através de sua espessura de Breslow (mm), presença de ulceração e taxa mitótica (mitoses / mm²). N significa o grau do envolvimento do linfonodo sentinela e M define a presença e localização de metástases (Balch *et al.*, 2009) (tabela 1). A partir do sistema de classificação, foram designados quatro estágios clínicos e patológicos (tabela 2).

Tabela 1. Classificação TNM para melanoma.

Classificação		
T	Espessura de Breslow (mm)	Status da ulceração / mitoses
T _{is}	NA	NA
T1	≤ 1	a: sem ulceração e taxa mitótica < 1 / mm ² b: com ulceração e taxa mitótica ≥ 1 / mm ²
T2	1,01 – 2,00	a: sem ulceração b: com ulceração
T3	2,01 – 4,00	a: sem ulceração b: com ulceração
T4	> 4,00	a: sem ulceração b: com ulceração
N	Nº de linfonodos metastáticos	
N0	0	NA
N1	1	a: micrometástases ^a b: macrometástases ^b
N2	2 – 3	a: micrometástases ^a b: macrometástases ^b c: metástases em trânsito/satélites sem nodos metastáticos
N3	4 ou mais	
M	Sítio	LDH
M0	Sem metástases à distância	NA
M1a	Metástases na pele, subcutâneas ou nodais	Normal
M1b	Metástases no pulmão	Normal
M1c	Outras metástases viscerais	Normal
	Qualquer metástase à distância	Elevado

T_{is}, Tumor *in situ*; NA, não se aplica; LDH, Lactato desidrogenase.

^a Micrometástases são diagnosticadas depois da biópsia do linfonodo sentinela; ^b Macrometástases são definidas como metástases nodais clinicamente detectáveis e confirmadas patologicamente. Adaptado de (Balch *et al.*, 2009).

Tabela 2. Estadiamento do melanoma com base na classificação TNM.

Estadiamento clínico ^a				Estadiamento patológico ^b			
	T	N	M		T	N	M
0	T/s	N0	M0	0	T/s	N0	M0
IA	T1a	N0	M0	IA	T1a	N0	M0
IB	T1b	N0	M0	IB	T1b	N0	M0
	T2a	N0	M0		T2a	N0	M0
IIA	T2b	N0	M0	IIA	T2b	N0	M0
	T3a	N0	M0		T3a	N0	M0
IIB	T3b	N0	M0	IIB	T3b	N0	M0
	T4a	N0	M0		T4a	N0	M0
IIC	T4b	N0	M0	IIC	T4b	N0	M0
III	Qualquer T	N > 0	M0	IIIA	T1-4 ^a	N1a	M0
					T1-4 ^a	N2a	M0
				IIIB	T1-4b	N1a	M0
					T1-4b	N2a	M0
					T1-4 ^a	N1b	M0
					T1-4 ^a	N2b	M0
					T1-4 ^a	N2c	M0
				IIIC	T1-4b	N1b	M0
					T1-4b	N2b	M0
					T1-4b	N2c	M0
					Qualquer T	N3	M0
IV	Qualquer T	Qualquer N	M1	IV	Qualquer T	Qualquer N	M1

^a Estadiamento clínico inclui o microestadiamento do melanoma primário e avaliação clínica / radiológica de metástases.

^b Estadiamento patológico é o microestadiamento do melanoma primário e informações patológicas do linfonodo regional após linfadenectomia parcial ou completa. Pacientes em estágios patológicos 0 ou IA são exceções, pois não necessitam de avaliação patológica de seus linfonodos. *Adaptado de* (Balch *et al.*, 2009).

Nos estágios iniciais não-metastáticos do câncer, a cirurgia é a principal conduta para o tratamento. Quimioterapia e imunoterapia têm sido estudadas como outras formas de terapia. A quimioterapia tem seus efeitos limitados na sobrevivência geral dos pacientes devido a mecanismos quimioresistentes (La Porta, 2007; Rass e Hassel, 2009). Apenas uma pequena subpopulação de pacientes atinge benefícios duráveis a partir da imunoterapia (Qin *et al.*, 2013).

1.2 Epidemiologia

Nas últimas cinco décadas, a incidência de melanoma vem aumentando de forma constante entre caucasianos (Erdmann *et al.*, 2013). Aproximadamente 250.000 novos casos são diagnosticados por ano no mundo e são responsáveis por cerca de 60.000 mortes (Internacional Agency for Research on Cancer. http://globocan.iarc.fr/Pages/burden_sel.aspx [acessado em 21 de novembro de 2015]).

Além de representar a causa mais comum de morte por câncer de pele em escala global (Siegel *et al.*, 2014), o melanoma atinge indivíduos no auge de suas vidas (idade média de 52 anos), quase uma década antes do surgimento da maioria dos tumores sólidos (Erdei e Torres, 2010).

A incidência aumentada dessa neoplasia pode ser atribuída a uma maior exposição à radiação UV do sol (Erdmann *et al.*, 2013) assim como a um melhor entendimento da doença, permitindo diagnósticos mais precisos e precoces (Lens e Dawes, 2004; Solomon *et al.*, 2004; Siegel *et al.*, 2014). Adicionalmente, autores sustentam que o longo período de latência entre a iniciação da carcionogênese do melanoma até a apresentação clínica pode se estender por décadas, fato este que ajudaria a explicar porque as suas taxas de incidência continuam aumentando em muitos países (Erdmann *et al.*, 2013).

Segundo o Instituto Nacional de Câncer (INCA), a incidência estimada da doença no Brasil para o ano de 2016 é baixa (3.000 casos novos em homens e 2.670 em mulheres), no entanto, a sua letalidade é elevada. Este

mesmo levantamento mostrou que as maiores taxas de incidência encontram-se na região sul do Brasil, sendo 8,02 casos para cada 100 mil homens e 7,06 casos para cada 100 mil mulheres no Rio Grande do Sul.

A patogênese do melanoma é influenciada pela interação de fatores genéticos e ambientais, uma vez que a incidência desta neoplasia é fortemente afetada pela cor da pele e localização geográfica (Balch, 2002). Deste modo, o melanoma é uma doença que acomete, predominantemente, caucasianos e, a sua incidência, é de 5 a 20 vezes menor entre populações de pele escura (Houghton e Polsky, 2002).

Atualmente, a grande maioria dos pacientes diagnosticados com melanoma é curada devido ao diagnóstico nos estágios iniciais da doença, o que permite intervenção cirúrgica prevenindo a progressão da doença (Erdei e Torres, 2010). Por outro lado, quando diagnosticado tardiamente, o melanoma pode atingir uma grande variedade de órgãos à distância. Como consequência, o diagnóstico nos estágios mais avançados da doença está associado a um mau prognóstico (Houghton e Polsky, 2002).

Pacientes com tumores primários espessos ou metástases linfonodais frequentemente desenvolvem metástases a distância e neste estágio, a média de sobrevida é de 6 a 8 meses (Cummins *et al.*, 2006) e a taxa de sobrevida em 5 anos é de apenas 15% (Finn *et al.*, 2012; Siegel *et al.*, 2013).

1.3 Metástases

O melanoma é conhecido pela sua rápida progressão, metástases e mau prognóstico. É responsável por mais de 80% das mortes relacionadas à neoplasias cutâneas (Miller e Mihm, 2006). Cerca de 1 em 5 pacientes (20%) com melanoma desenvolverão metástases (Siegel *et al.*, 2013).

O tumor tem capacidade de metastatizar muito cedo no curso da doença e por isto, reconhecer os pacientes em risco de progressão tumoral é crucial. Aproximadamente 5% dos pacientes que apresentam lesões finas (<1,0 mm de espessura); 25-40% com lesões de 2,0-4,0 mm; e 50-75% com tumores >4,0 mm de espessura desenvolverão doença metastática (Balch *et al.*, 2001).

As metástases tanto podem ser regionais quanto podem atingir órgãos distantes. Os sítios mais comuns das metástases regionais são: a pele nas imediações do tumor primário, o tecido subcutâneo e os linfonodos (Damsky *et al.*, 2010). Dependendo da distância entre a metástase e o tumor primário, as metástases de pele podem ser classificadas como satelitoses (mais próximas à lesão primária) ou metástase em trânsito (mais distantes do melanoma primário) (Balch *et al.*, 2009). O surgimento de metástases cutâneas é o melhor indicativo de disseminação linfática ou hematogênica da doença (Wolf *et al.*, 2004). Os órgãos preferenciais para ocorrência de metástases de melanoma à distância são: pele, pulmão, cérebro, fígado, ossos e intestino. Metástases pulmonares são prevalentes e, geralmente, o primeiro local onde surgem às metástases viscerais. Outros sítios metastáticos como ossos e intestino ocorrem mais tardiamente na progressão da doença (Damsky *et al.*, 2010).

Atualmente, é reconhecido que múltiplos mecanismos celulares estão desregulados na iniciação e progressão do tumor, como proteínas reguladoras do ciclo celular, apoptoses, transdução de sinais, adesão das células e digestão da matriz extracelular (Meyer *et al.*, 2012).

Em resumo, as etapas para a progressão tumoral de lesões oriundas de nevos displásicos incluem a conversão de uma lesão pigmentada para a forma de melanoma *in situ*, que cresce lateralmente na epiderme (fase de crescimento radial) (Haass e Smalley, 2009). Se não tratado adequadamente, o melanoma pode progredir para a fase de crescimento vertical, a qual está associada com invasão da derme e, frequentemente, com o aparecimento de metástases (Clark, 1991; Sondak *et al.*, 2001).

A perda da adesão celular, a repressão da expressão de E-caderinas e o aumento da motilidade da célula são fenômenos que em conjunto denominam-se “transição epitélio-mesenquimal (EMT)” (Their, 2002; Huber *et al.*, 2005). Tal mecanismo está envolvido na invasividade e metástase de MM. Os melanócitos estão aderidos à queratinócitos basais na epiderme através da expressão de E-caderinas (Hsu *et al.*, 1996). O escape inicial de células neoplásicas de seu local de origem está associado com alterações transcriptômicas e proteômicas de um perfil epitelial para mesenquimal. Sendo assim, moléculas típicas do epitélio como as E-caderinas são substituídas por N-caderinas do mesênquima o que aumenta a motilidade celular, culminando na invasão e migração dos melanócitos transformados para órgãos distantes (Poser *et al.*, 2001; Thiery, 2009).

O melanoma é frequentemente fatal quando a doença progride para o estágio metastático e geralmente é resistente às terapias disponíveis, além destes tratamentos possuírem limitações, custo elevado e desencadearem reações adversas (Slominski e Carlson, 2014).

A dacarbazina, o único agente quimioterápico aprovado pelo *Food and Drug Administration* (FDA) para o tratamento do melanoma metastático está associado com uma resposta de 7 a 12% e uma sobrevida global de 5,6 a 7,8 anos depois do início do tratamento (Chapman *et al.*, 1999).

Várias estratégias para estimular a resposta imune antineoplásica tem sido exploradas. Historicamente, altas doses de interleucina 2 (IL-2) e interferon α eram administrados para pacientes com melanoma metastático e para melanomas ressecados (estágios clínicos II e III) com alto risco de recorrência, respectivamente. Cerca de 15 a 20% dos pacientes com melanoma metastáticos que recebiam altas doses de IL-2 obtinham resposta terapêutica e 6 a 8% experimentavam remissão completa da doença por mais de três anos após o tratamento (Atkins *et al.*, 1999). No entanto, efeitos tóxicos agudos incluindo disfunção de múltiplos órgãos e comprometimento hemodinâmico são observados a partir da administração desta terapia (Schwartzentruber, 2001). Mais recentemente, o uso de ipilimumab, um anticorpo monoclonal que bloqueia o antígeno 4 de linfócito T citotóxico (CTLA4) nos linfócitos, tem sido associado com o aumento da sobrevida dos pacientes metastáticos (Hodi *et al.*, 2010). Outro anticorpo monoclonal que tem sido desenvolvido para o tratamento do melanoma avançado é o nivolumab, o qual tem como alvo terapêutico o receptor de morte programada 1 (PD-L1) e

seus ligantes, promovendo a regulação negativa da ativação das células T e assim, a imunidade antitumoral (Ott *et al.*, 2013). Ensaios clínicos fase I, II e III tem observado eficácia e segurança promissoras de nivolumab em comparação e associação com outras terapias (Johnson *et al.*, 2015).

Aproximadamente 40 a 60% dos melanomas cutâneos apresentam mutações no gene BRAF que leva a ativação constitutiva da via da MAP-quinase. Entre as mutações de BRAF, a mais freqüente, em 90% dos casos, é a V600E, a qual resulta na substituição do aminoácido glutamina pela valina no códon 600 (Chapman *et al.*, 2011). Os resultados de um ensaio clínico de fase III mostraram que o Vemurafenib, um inibidor específico do BRAF mutado, foi capaz de aumentar a taxa de sobrevida global e livre de progressão em pacientes com melanoma metastático em comparação com o grupo tratado com dacarbazina (Chapman *et al.*, 2011).

Embora, nos últimos anos, o tratamento para melanoma metastático tenha avançado significativamente, a média da sobrevida dos pacientes tratados é menor do que um ano (Eggermont *et al.*, 2014). Sendo assim, a detecção e o diagnóstico dos tumores nos estágios iniciais da doença, ainda constituem uma abordagem crucial para aumentar a taxa de sobrevivência e qualidade de vida do paciente (Geller *et al.*, 2007).

1.4 Biomarcadores no prognóstico do melanoma

Biomarcadores são definidos como indicadores de processos biológicos, condições patológicas ou respostas farmacológicas que podem ser

quantificados e avaliados, permitindo o diagnóstico de uma doença específica, o acesso ao prognóstico da doença ou o monitoramento da resposta ao tratamento (Colburn *et al.*, 2001).

É consenso na literatura que o sistema de classificação e prognosticação definido pelo *American Joint Committee on Cancer* (AJCC, 2009) (Balch *et al.*, 2010; Gershenwald *et al.*, 2010), não é suficiente para acessar o risco de mortalidade para cada paciente individualmente dentro de cada estágio (Friedman *et al.*, 2012).

Os principais preditores de desfecho são: espessura de Breslow, taxa mitótica, presença de ulceração e envolvimento do linfonodo sentinela (Balch *et al.*, 2001). No entanto, existe um subgrupo de lesões finas, não ulceradas e sem atividade mitótica que apresentam comportamento altamente agressivo e que escapam da identificação dos fatores histopatológicos de prognóstico (Abbas *et al.*, 2014).

O melanoma tem sido reconhecido, há vários anos, como sendo altamente heterogêneo, de tal modo que aproximadamente 5% dos tumores em estágio I desenvolvem metástases precocemente podendo levar o paciente a óbito (Gimotty *et al.*, 2005; Balch *et al.*, 2009), enquanto determinadas lesões espessas não exibem este comportamento (Ratzinger *et al.*, 2014).

Muitos são os esforços dispensados pelos pesquisadores para conhecer os mecanismos moleculares envolvidos na progressão do melanoma, bem como para o desenvolvimento de ferramentas que possam ajudar no

desenvolvimento de modelos com capacidade de prever o curso provável da doença (Raskin *et al.*, 2013). Para tanto, diversos biomarcadores moleculares de tecido e sanguíneos tem sido investigados (Hoshimoto *et al.*, 2012; Jakob *et al.*, 2012; Xu *et al.*, 2012; Brunner *et al.*, 2013).

No momento, apenas a quantificação dos níveis de LDH sérico foi incorporada na rotina clínica como parte do sistema de estadiamento do melanoma (Hill *et al.*, 2015). Entretanto, estudos mostram que este marcador possui algumas limitações quanto à sua sensibilidade nos estágios iniciais do melanoma, porém, tem valor preditivo negativo para recidivas metastáticas (Hofmann *et al.*, 2009; Hofmann *et al.*, 2011).

Segundo (Saldanha *et al.*, 2016)), um dos maiores problemas enfrentados na busca por marcadores relevantes é que as macromoléculas são mal preservadas nas amostras de tecido de melanoma uma vez que, normalmente, todo o tecido tumoral é fixado em formalina e embocado em parafina. Além do mais, a maioria dos estudos possui um tamanho amostral reduzido dificultando a inclusão dos biomarcadores avaliados na prática clínica (El Hajj *et al.*, 2013).

A urgência na busca pelo desenvolvimento de biomarcadores complementares com potencial para o prognóstico específico do paciente com melanoma se justifica na medida em que poderia implicar na personalização de seu tratamento com melhoria no desfecho clínico (Schramm e Mann, 2011). Marcadores capazes de identificar pacientes com alto e baixo risco de desenvolver metástase poderiam fornecer informações adicionais, tais como

indicar aqueles com necessidade de monitoramento intensificado (Spatz *et al.*, 2010; Murali *et al.*, 2011; Murali *et al.*, 2012).

1.5 Cell-free DNA e inflamação

1.5.1 Liberação de DNA na corrente sanguínea

O DNA livre circulante (*cell-free* DNA ou cfDNA) é definido como moléculas de DNA extracelulares que ocorrem no sangue. O cfDNA pode ser detectado no plasma sanguíneo tanto de pacientes com câncer ou com outras doenças (p. e. processos inflamatórios severos) (Jiang e Pisetsky, 2005), quanto em pessoas saudáveis (Jung *et al.*, 2010).

A maior parte do DNA circulante apresenta-se no sangue como moléculas dupla-fita com peso molecular inferior ao DNA genômico e arranjadas em nucleossomos (Jung *et al.*, 2010). Entretanto, os mecanismos responsáveis pela liberação de cfDNA na circulação, em ambas condições fisiológica e patológica, ainda não são completamente entendidos.

Foi sugerido que, em indivíduos saudáveis, a presença de cfDNA ocorre devido a processos hematopoiéticos, onde as células liberam fragmentos de DNA de baixo peso molecular chamados de fração de DNA metabólico (Stroun *et al.*, 2001). O DNA metabólico pode formar um complexo com glicoproteínas que, na corrente sanguínea, atuaria como mensageiro, sinalizando funções entre células e tecidos (Gahan, 2006).

Além dos processos normais de liberação de cfDNA na circulação, portadores de câncer possuem vias adicionais de liberação desse tipo de DNA extracelular. São elas a apoptose e a necrose, processos que ocorrem com frequência em células tumorais com taxas elevadas de proliferação celular (Li *et al.*, 2003). Células mortas em excesso levam à saturação do processo de fagocitose pelos macrófagos e fragmentos de DNA degradado são liberados na circulação, o que acaba por elevar os níveis de cfDNA no sangue (Stroun *et al.*, 2001).

Alternativamente, uma baixa parcela de cfDNA origina-se da lise de células neoplásicas frágeis que se desprendem do tumor e entram na corrente sanguínea durante a disseminação do tumor. No entanto, este processo é dependente do tipo de câncer (Stroun *et al.*, 2000).

Concentrações mais elevadas de cfDNA frequentemente são encontradas em pacientes com câncer em relação àqueles saudáveis (Van Der Vaart e Pretorius, 2010). Sendo assim, vários estudos atribuem valor prognóstico à quantificação de DNA livre no sangue (Jung *et al.*, 2004; Pathak *et al.*, 2006; Ren *et al.*, 2006; Catarino *et al.*, 2008; Yoon *et al.*, 2009; Kamat *et al.*, 2010; Ulivi e Silvestrini, 2013; Spindler *et al.*, 2015). Em pacientes portadores de câncer de mama (Catarino *et al.*, 2008), fígado (Ren *et al.*, 2006), pulmão (Yoon *et al.*, 2009), ovário (Kamat *et al.*, 2010) e próstata (Jung *et al.*, 2004), altas concentrações mostraram-se – na maioria dos casos – como fatores indicativos para um pior desfecho quanto à sobrevida global ou livre de doença.

1.5.2 O papel da inflamação no câncer

Evidências sugerem que pelo menos 20% de todas as neoplasias surgem associadas a infecções e inflamações crônicas e, até mesmo aqueles tumores que não se desenvolvem como consequência destes processos, exibem extensos infiltrados inflamatórios com a expressão de níveis elevados de citocinas no microambiente tumoral (Coussens e Werb, 2002; Grivennikov e Karin, 2010).

Citocinas inflamatórias são pequenas proteínas mediadoras que desempenham papel-chave na resposta imunológica. São secretadas principalmente por linfócitos e macrófagos e possuem efeito específico na interação e comunicação entre células. Durante a resposta inflamatória, as citocinas influenciam na atividade, diferenciação, proliferação e sobrevivência de células imunológicas, bem como regulam a produção e atividade de outras citocinas que podem aumentá-la (citocinas pró-inflamatórias) ou reduzi-la (citocinas anti-inflamatórias) (Oliveira *et al.*, 2011). São exemplos de citocinas pró-inflamatórias (resposta Th1) as interleucinas 1 e 6 (IL-1 e IL-6), fator de necrose tumoral alfa (TNF- α) e interferon gama (IFN- γ). Entretanto, a resposta persistente e exacerbada de citocinas pró-inflamatórias pode levar a danos nos tecidos-alvo. Sendo assim, citocinas anti-inflamatórias, tais como IL-4, IL-10 e IL-13, podem regular a resposta imunossupressora, minimizando os efeitos indesejáveis da inflamação (Curfs *et al.*, 1997; Lin, E. *et al.*, 2000).

As citocinas podem atuar nas células que as secretam (ação autócrina), em células adjacentes (ação parácrina) ou, menos comumente, em células

distantes (ação endócrina) (Lin, E. *et al.*, 2000). Além disto, estas proteínas apresentam redundância nas atividades que desempenham, significando que funções similares podem ser estimuladas por citocinas diferentes. Geralmente são produzidas em cascata, à medida que uma citocina estimula a célula-alvo a produzir citocinas adicionais (Zhang e An, 2007).

Uma vez que as citocinas não podem ser agrupadas de acordo com as suas células de origem ou sua função biológica, elas são classificadas em interleucinas (IL; numeradas da IL-1 a IL-35), fatores de necrose tumoral (TNF), quimiocinas (citocinas quimiotáticas), interferons (IFN) e fatores de crescimento mesenquimal (Curfs *et al.*, 1997). Além disso, as citocinas podem ser classificadas em tipo I (p. e. TNF e IFN- γ), as quais estão envolvidas na resposta imunológica Th1 (T *helper* 1) e tipo II (p. e. IL-4, IL-10 e IL-13), envolvidas na resposta imune Th2 (T *helper* 2). A resposta Th1 é mediada por citocinas pró- inflamatórias expressas por células T CD4+ que induzem as células do sistema imune a atuarem contra infecções e malignidades (Dranoff, G., 2004; Smyth *et al.*, 2004). Já a resposta Th2, que também é mediada por citocinas produzidas pelas células T CD4+, promove imunidade humoral contra tumores e/ou desvio do sistema imune para uma resposta não protetora (tolerância) (Dranoff, Glenn, 2004; Smyth *et al.*, 2004).

As citocinas podem estar envolvidas tanto na ativação de mecanismos imunes que limitam o crescimento tumoral, quanto atuar como fatores de crescimento e sobrevivência de células pré-malignas (Karin e Greten, 2005), uma vez que são capazes de estimular a angiogênese, auxiliar na progressão

tumoral e metástases e também, manter a inflamação promotora do tumor (Karin, 2006; Yu *et al.*, 2009; Grivennikov e Karin, 2010).

As citocinas são produzidas pelo estroma do hospedeiro e por células do sistema imune em resposta a moléculas secretadas pelas células cancerígenas ou como parte do processo inflamatório que freqüentemente acompanha o crescimento do tumor. Além disto, as células malignas também contribuem secretando citocinas no ambiente tumoral (Smyth *et al.*, 2004). Neste ambiente, os níveis e a composição do conjunto de citocinas sofrem flutuações nos vários estágios do desenvolvimento do tumor (Smyth *et al.*, 2004). Essas proteínas podem atuar de modo a desempenhar um efeito aditivo, sinérgico ou antagonista. Portanto, é a integração destes efeitos na resposta antitumoral que determinará o desfecho geral (Lin, Edward *et al.*, 2000).

De acordo com a literatura, três cenários são possíveis no que se refere ao envolvimento de células imunitárias na patogênese tumoral. Densos infiltrados intratumorais de linfócitos nos estágios iniciais da tumorigênese estão fortemente correlacionados com freqüências reduzidas de metástases e maior sobrevida em vários tipos de câncer (Clark *et al.*, 1989; Clemente *et al.*, 1996; Naito *et al.*, 1998; Nakano *et al.*, 2001; Zhang *et al.*, 2003), indicando que a resposta do hospedeiro é capaz de atenuar a progressão da doença (Dunn *et al.*, 2002). Alternativamente, a inflamação crônica pode aumentar o risco de transformação maligna quando não resolvida (Ames *et al.*, 1995) e, portanto, a resposta imune do hospedeiro pode promover o desenvolvimento tumoral (Coussens e Werb, 2002). Por último, estudos mostram que certas amostras de tumores, particularmente aquelas obtidas durante o estabelecimento da fase

disseminativa da doença, não apresentam infiltrado de células imunológicas, indicando que estas neoplasias podem permanecer indetectáveis pelo sistema imune (Ochsenbein *et al.*, 2001).

O melanoma é um tipo de câncer fortemente associado com processos inflamatórios devido à secreção de altos níveis de citocinas pelas células tumorais (Richmond *et al.*, 1988; Claffey *et al.*, 1996; Amiri e Richmond, 2005; Leslie e Bar-Eli, 2005). A imunogenicidade do melanoma é atribuída aos seus antígenos, tais como: MART-1, gp100, gp75 e tirosinase (Hussein, 2005).

Apesar da tentativa do hospedeiro de eliminar a doença, a qual é representada através da resposta imune massiva contra antígenos do melanoma, é rara a sua regressão espontânea mediada pelo sistema imunológico (Tikoo e Haass, 2015). De fato, evidências sugerem que as citocinas secretadas pelas células do melanoma (p.e. interleucina 8, GRO α , GRO β , GRO γ , entre outras) exercem controle autócrino durante o crescimento e promoção da angiogênese no microambiente tumoral (Richmond *et al.*, 1988; Claffey *et al.*, 1996; Amiri e Richmond, 2005; Leslie e Bar-Eli, 2005).

1.6 Polimorfismo da enzima MnSOD

1.6.1 Espécies reativas de oxigênio e o estresse oxidativo

O termo espécies reativas de oxigênio (EROs) compreende moléculas derivadas de oxigênio com capacidade de receber elétrons extras e que podem oxidar outras moléculas (Cross *et al.*, 1987). As EROs contêm um ou mais

elétrons desemparelhados, o que faz com que estas moléculas ou elementos moleculares se tornem altamente reativos na busca por estabilização eletrônica (Halliwell e Gutteridge, 2015). As três formas primárias de EROs são: ânion radical superóxido ($O_2^{\cdot-}$), derivado da redução de um único elétron de oxigênio; peróxido de hidrogênio (H_2O_2), que resulta da conversão de duas moléculas superóxido em uma molécula de ERO não-radical e uma molécula de água através de enzimas superóxido dismutases; radical hidroxila ($\cdot OH$), formado por meio da reação de Fenton, na qual o peróxido de hidrogênio é reduzido pela doação de um elétron de Fe^{2+} (Sullivan e Chandel, 2014) (figura 3).

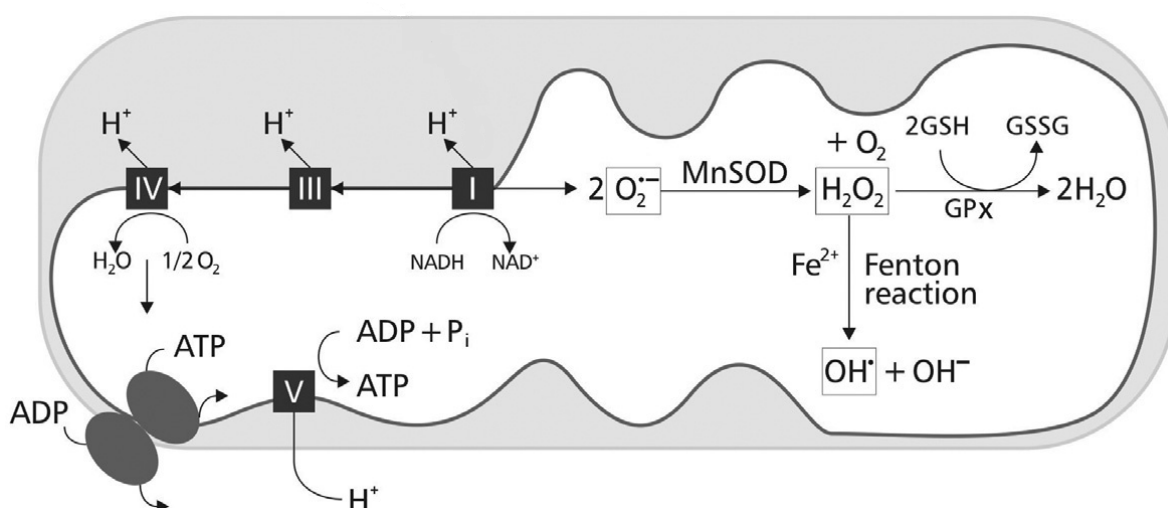


Figura 3. Produção de EROs na mitocôndria. NADH coleta elétrons de moléculas de oxigênio (O_2). Os elétrons fluem através da cadeia mitocondrial de transportes de elétrons para síntese de ATP. Uma pequena quantidade de O_2 sofre uma redução incompleta dando origem ao ânion superóxido ($O_2^{\cdot-}$), a primeira espécie reativa de oxigênio dos organismos aeróbicos. Posteriormente, a enzima superóxido dismutase dependente de manganês (MnSOD) dismuta $O_2^{\cdot-}$ em peróxido de hidrogênio (H_2O_2) e O_2 ; H_2O_2 é ainda neutralizado pela glutatona peroxidase (GPX) em H_2O com glutatona (GSH) como substrato. Igualmente, H_2O_2 pode ser convertido no radical

hidroxila ($\cdot\text{OH}$) através da reação de Fenton com ferro (Fe^{2+}). *Adaptado de* (Edeas, 2011).

EROs são produzidos durante o metabolismo aeróbico normal como produto de diversos processos celulares, principalmente na cadeia de transportes de elétrons na mitocôndria, bem como por agentes tóxicos ambientais (p.e. radiação, poluição do ar e xenobióticos) (Sosa *et al.*, 2013). Certos níveis de EROs são indispensáveis a medida que atuam como mediadores em processos celulares, tais como, diferenciação, controle da progressão do ciclo celular, apoptose e imunidade (Gonzales *et al.*, 1984; Abdel-Aziz e El-Naggar, 1997). No entanto, a produção excessiva de espécies reativas de oxigênio – tanto por questões patológicas quanto fisiológicas – acima da capacidade do sistema antioxidante de defesa do organismo, pode acabar resultando em estresse oxidativo (Ambrosone *et al.*, 1999).

As espécies reativas de oxigênio reagem com diferentes alvos intracelulares, incluindo lipídios, proteínas e DNA. Sendo assim, o estresse oxidativo pode resultar na quebra da molécula de DNA, na peroxidação de lipídios, na modificação de proteínas, na ruptura de membranas e no dano mitocondrial (Schwartz *et al.*, 1993; Emerit, 1994). Frequentemente, essa interação provoca danos às macromoléculas, o que pode resultar na morte das células, mutação ou carcinogênese (Ambrosone *et al.*, 1999).

Evidências sugerem que as EROs estão envolvidas em vários estágios do desenvolvimento em múltiplos passos do câncer (Marnett, 2000). O principal mecanismo pelo qual as EROs contribuem para a carcinogênese é através de

mutações e alterações nas expressões gênicas geradas a partir de lesões ao DNA provenientes do estresse oxidativo (Feig *et al.*, 1994; Cooke *et al.*, 2003). O radical hidroxila reage com todos os componentes da molécula de DNA, danificando tanto as bases púricas e pirimídicas quanto a desoxirribose (Halliwell e Gutteridge, 2015). A lesão oxidativa do DNA mais proeminente é a 8-Hidroxi guanina (8-OH-G), representando o primeiro passo nos processos de mutagênese, carcinogênese e envelhecimento (Kasai e Nishimura, 1991). Durante a replicação celular, 8-OH-G pode causar a transversão de GC→TA pelo pareamento incorreto com uma adenina (Cheng *et al.*, 1992; Moriya, 1993; Kamiya *et al.*, 1995).

1.6.2 Estresse oxidativo no melanoma

No que se refere às neoplasias dermatológicas, a radiação ultravioleta (UV) é um fator de risco bem-estabelecido que pode contribuir na geração de dano oxidativo e, conseqüentemente, no processo de carcinogênese (Nishigori *et al.*, 2004; Sander *et al.*, 2004). EROs são geradas indiretamente após a absorção de raios UVA (320-400 nm) pelos cromóforos das células da pele (Brash, 1997; Kielbassa *et al.*, 1997). Os raios UVB (280-320 nm), em uma menor proporção, também são capazes de induzir o estresse oxidativo (Kvam e Tyrrell, 1997).

No melanoma, sabe-se que espécies reativas de oxigênio estão presentes em todas as fases do seu desenvolvimento e que há um desequilíbrio no estado redox dos pacientes (Kvam e Tyrrell, 1997; Gadjeva *et al.*, 2008). Sander *et al.* (2003) demonstram a expressão excessiva de enzimas

antioxidantes em biópsias de pacientes com melanoma quando comparadas aos tecidos adjacentes ao tumor, aos nevos melanocíticos benignos e aos indivíduos controle. Adicionalmente, os autores concluíram que o aumento do estresse oxidativo encontrado nas células de melanoma poderia danificar os tecidos que circundam o tumor e então, auxiliar na sua progressão metastática.

1.6.3 Detoxificação enzimática de espécies reativas de oxigênio

Devido ao efeito deletério das EROs, diversos mecanismos de defesa evoluíram para proteger as células. Estes mecanismos incluem os sistemas de reparo, as enzimas antioxidantes (superóxido dismutases, SOD; catalase, CAT; e glutathione peroxidase, GPX) e pequenas moléculas “scavengers”, tais como a glutathione (GSH) (Gadjeva *et al.*, 2008).

Na pele, a modulação enzimática dos níveis de EROs é realizada pela superóxido dismutase dependente de cobre e zinco (CuZnSOD), superóxido dismutase dependente de manganês (MnSOD) e catalase, as quais mantêm o balanço redox dentro das células (Sander *et al.*, 2003). Entre estas enzimas, a MnSOD tem despertado interesse, uma vez que localiza-se na mitocôndria e é a primeira linha de defesa contra os radicais superóxido produzidos como produto da fosforilação oxidativa oriunda da exposição aos raios UV e de processos inflamatórios (Poswig *et al.*, 1999).

Na mitocôndria, a MnSOD atua convertendo EROs em peróxido de hidrogênio, o qual é catalisado em água, na sequência, pela catalase e pela glutathione peroxidase (Wallace, 1999; Li *et al.*, 2005). Além do mais, tem sido

demonstrado que a MnSOD tem capacidade de atuar como supressor tumoral, possivelmente modulando as vias apoptóticas e proliferativas *in situ*. (Oberley e Buettner, 1979; Li *et al.*, 1998). A expressão aumentada desta enzima foi observada na supressão do fenótipo maligno de células tumorais, até mesmo no melanoma (Church *et al.*, 1993).

A enzima MnSOD é codificada pelo gene superóxido dismutase 2 (SOD2), um gene de cópia única que contém cinco éxons espaçados por quatro introns e está localizado no cromossomo 6q25.3 (Wan *et al.*, 1994). O éxon 2 (códon 16) do gene SOD2 abriga um polimorfismo de nucleotídeo único (SNP), ocorrendo a substituição de T (timina) por C (citosina). Extensivamente estudado, este polimorfismo denominado Ala16Val, resulta na troca do aminoácido valina (Val, GTT) pelo aminoácido alanina (Ala, GCT) (Rosenblum *et al.*, 1996).

A substituição de aminoácidos ocorre na porção N-terminal da sequência que serve para direcionar o transporte da proteína para dentro da mitocôndria (*mitochondrial targeting sequence*, MTS), uma vez que o precursor da enzima é sintetizado no citoplasma (Pfanner e Geissler, 2001). Portanto, sugere-se que este polimorfismo altere a estrutura secundária da proteína implicando na eficiência do transporte da MnSOD para o interior da mitocôndria (Rosenblum *et al.*, 1996). O alelo variante Ala (A-MnSOD) permite uma importação mais eficiente da enzima para a matriz mitocondrial, devido à sua tradução em uma cadeia de aminoácidos precursora da MnSOD com estrutura em alfa-hélice, o que resulta em uma maior atividade enzimática quando comparado ao alelo Val (V-MnSOD), organizada em uma estrutura beta-folha que encontra dificuldade

em atravessar os poros da membrana interna da mitocôndria, sendo parcialmente destruída no proteossoma (Sutton *et al.*, 2003) (figura 4). Desse modo, sugere-se que indivíduos portadores do genótipo homozigoto AA (Ala/Ala) possuem maior atividade da MnSOD do que os homozigotos VV (Val/Val) (Sutton *et al.*, 2003).

Embora ainda muito controversa, diversos estudos sugerem haver associação entre a susceptibilidade ao câncer e o genótipo AA (Ambrosone *et al.*, 1999; Cheng *et al.*, 2005; Li *et al.*, 2005). Como, por exemplo, no câncer de mama, onde o risco de desenvolvimento da doença está aumentado em duas vezes, aproximadamente, nos indivíduos AA (Mitrinen *et al.*, 2001; Cai *et al.*, 2004; Slanger *et al.*, 2006), porém o genótipo VV pode estar correlacionado com tumores com maior potencial metastático (Bica *et al.*, 2010). De modo semelhante, foi demonstrada a associação entre o polimorfismo do SOD2 e o câncer de próstata, onde o alelo Ala estava associado com um risco maior a essa neoplasia (Woodson *et al.*, 2003). Já no caso das neoplasias cutâneas, não foi observada associação significativa entre o genótipo AA e o risco de melanoma, carcinoma basocelular e carcinoma epidermóide (Han *et al.*, 2007). De acordo com nossa busca na literatura, até o momento, o estudo de Han *et al.* (2007) foi o único que avaliou a associação entre o polimorfismo Ala16Val do gene SOD2 no melanoma.

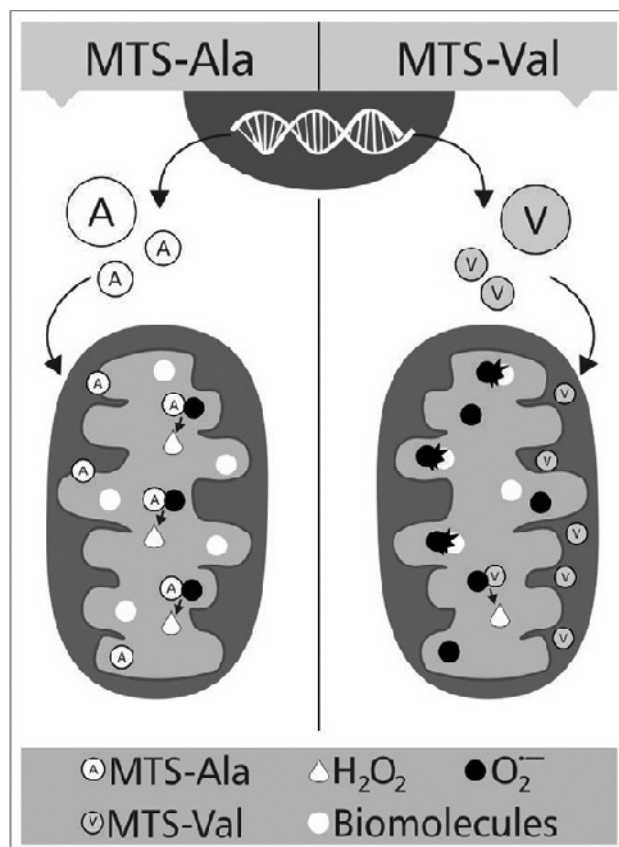


Figura 4. Conteúdo de MnSOD na mitocôndria de acordo com os diferentes precursores originados do polimorfismo da enzima: alelo variante Ala e alelo variante Val. Enquanto que o precursor da MnSOD Ala-variante (MTS-Ala) é corretamente transportado através das duas membranas mitocôndrias, o precursor Val-variante (MTS-Val) é parcialmente contido na membrana interna da mitocôndria. O aumento do precursor da MnSOD contendo o alelo Ala é traduzido em uma detoxificação mais eficiente do superóxido, enquanto que o precursor Val permite o acúmulo de EROs na mitocôndria. *Adaptado de (Kurmi et al., 2013).*

1.7 Justificativa

Dada a natureza agressiva do melanoma, é grande o esforço despendido por investigadores na busca por biomarcadores moleculares para complementar o diagnóstico e prognóstico de pacientes em risco potencial de desenvolver metástases (Larson *et al.*, 2009). A identificação de tais pacientes ainda nos estágios iniciais da doença poderia beneficiá-los por meio da complementação do tratamento padrão com terapias adjuvantes, além de um acompanhamento mais rigoroso.

Atualmente, os principais fatores prognósticos aceitos na prática clínica são a espessura de Breslow, a taxa mitótica, a presença de ulceração e o status do linfonodo sentinela (Balch *et al.*, 2009; Abbas *et al.*, 2014). Entretanto, sabe-se que tumores semelhantes – clínico e histopatologicamente – em indivíduos diferentes podem ter comportamento distinto quanto à progressão da doença.

A avaliação dos níveis de cfDNA no sangue de pacientes com melanoma, bem como de seu status imunológico, vem de encontro a esta tendência de identificação precoce de pacientes com mau prognóstico. Além de estabelecer a melhor estratégia terapêutica, é importante que estes marcadores possam auxiliar na identificação de recidivas e metástases sub-clínicas, permitindo o seu tratamento imediato e o aumento da sobrevida (Larson *et al.*, 2009).

Quanto ao estresse oxidativo no melanoma, sabe-se que as espécies reativas de oxigênio geradas como consequência da incidência de radiação UV sob a pele têm um papel importante no processo de carcinogênese (125, 126). No entanto, até onde temos conhecimento, apenas um artigo se propôs a avaliar a associação do polimorfismo da MnSOD no melanoma e nenhuma associação foi observada. Sendo assim, consideramos que esta questão ainda não foi completamente elucidada e merece atenção.

1.8 Referências bibliográficas

ABBAS, O.; MILLER, D. D.; BHAWAN, J. Cutaneous malignant melanoma: update on diagnostic and prognostic biomarkers. **Am J Dermatopathol**, v. 36, n. 5, p. 363-79, May 2014. ISSN 0193-1091.

ABDEL-AZIZ, A.; EL-NAGGAR, M. Superoxide dismutase activities in serum and white blood cells of patients with some malignancies. **Cancer letters**, v. 113, n. 1-2, p. 61-64, 1997. ISSN 0304-3835.

AMBROSONE, C. B. et al. Manganese superoxide dismutase (MnSOD) genetic polymorphisms, dietary antioxidants, and risk of breast cancer. **Cancer Research**, v. 59, n. 3, p. 602-606, 1999. ISSN 0008-5472.

AMES, B. N.; GOLD, L. S.; WILLETT, W. C. The causes and prevention of cancer. **Proceedings of the National Academy of Sciences**, v. 92, n. 12, p. 5258-5265, 1995. ISSN 0027-8424.

AMIRI, K. I.; RICHMOND, A. Role of nuclear factor- κ B in melanoma. **Cancer and Metastasis Reviews**, v. 24, n. 2, p. 301-313, 2005. ISSN 0167-7659.

ATKINS, M. B. et al. High-dose recombinant interleukin 2 therapy for patients with metastatic melanoma: analysis of 270 patients treated between 1985 and 1993. **Journal of clinical oncology**, v. 17, n. 7, p. 2105-2105, 1999. ISSN 0732-183X.

BALCH, C. M. et al. Final version of the American Joint Committee on Cancer staging system for cutaneous melanoma. **J Clin Oncol**, v. 19, n. 16, p. 3635-48, Aug 15 2001. ISSN 0732-183X.

BALCH, C. M. et al. Final version of 2009 AJCC melanoma staging and classification. **J Clin Oncol**, v. 27, n. 36, p. 6199-206, Dec 20 2009. ISSN 0732-183x.

BALCH, C. M. et al. Multivariate analysis of prognostic factors among 2,313 patients with stage III melanoma: comparison of nodal micrometastases versus macrometastases. **J Clin Oncol**, v. 28, n. 14, p. 2452-9, May 10 2010. ISSN 0732-183x.

BICA, C. G. et al. Polymorphism (ALA16VAL) correlates with regional lymph node status in breast cancer. **Cancer genetics and cytogenetics**, v. 196, n. 2, p. 153-158, 2010. ISSN 0165-4608.

BRASH, D. E. Sunlight and the onset of skin cancer. **Trends in genetics**, v. 13, n. 10, p. 410-414, 1997. ISSN 0168-9525.

BRUNNER, G. et al. A nine-gene signature predicting clinical outcome in cutaneous melanoma. **J Cancer Res Clin Oncol**, v. 139, n. 2, p. 249-58, Feb 2013. ISSN 0171-5216.

CAI, Q. et al. Genetic polymorphism in the manganese superoxide dismutase gene, antioxidant intake, and breast cancer risk: results from the Shanghai Breast Cancer Study. **Breast Cancer Research**, v. 6, n. 6, p. 1, 2004. ISSN 1465-542X.

CATARINO, R. et al. Quantification of Free Circulating Tumor DNA as a Diagnostic Marker for Breast Cancer. **DNA and Cell Biology**, v. 27, n. 8, p. 415-421, 2008.

CHAPMAN, P. B. et al. Phase III multicenter randomized trial of the Dartmouth regimen versus dacarbazine in patients with metastatic melanoma. **J Clin Oncol**, v. 17, n. 9, p. 2745-51, Sep 1999. ISSN 0732-183X.

CHAPMAN, P. B. et al. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. **N Engl J Med**, v. 364, n. 26, p. 2507-16, Jun 30 2011. ISSN 0028-4793.

CHENG, K. C. et al. 8-Hydroxyguanine, an abundant form of oxidative DNA damage, causes G----T and A----C substitutions. **Journal of Biological Chemistry**, v. 267, n. 1, p. 166-172, 1992. ISSN 0021-9258.

CHENG, T. C. et al. Breast cancer risk associated with genotype polymorphism of the catechol estrogen-metabolizing genes: A multigenic study on cancer susceptibility. **International journal of cancer**, v. 113, n. 3, p. 345-353, 2005. ISSN 1097-0215.

CHIN, L.; MERLINO, G.; DEPINHO, R. **Malignant melanoma: modern black plague and genetic black box**. DEPINHO, R. 12: 3467-3481 p. 1998.

CHURCH, S. L. et al. Increased manganese superoxide dismutase expression suppresses the malignant phenotype of human melanoma cells. **Proceedings of the National Academy of Sciences**, v. 90, n. 7, p. 3113-3117, 1993. ISSN 0027-8424.

CLAFFEY, K. P. et al. Expression of vascular permeability factor/vascular endothelial growth factor by melanoma cells increases tumor growth, angiogenesis, and experimental metastasis. **Cancer Res**, v. 56, n. 1, p. 172-81, Jan 01 1996. ISSN 0008-5472.

CLARK, W. Human cutaneous malignant melanoma as a model for cancer. **Cancer Metast Rev**, Dordrecht, v. 10, n. 2, p. 83-88, 1991. ISSN 0167-7659.

CLARK, W. H. et al. Model predicting survival in stage I melanoma based on tumor progression. **Journal of the National Cancer Institute**, v. 81, n. 24, p. 1893-1904, 1989. ISSN 0027-8874.

CLARK, W. H.; ELDER, D. E.; VAN HORN, M. The biologic forms of malignant melanoma. **Human Pathology**, v. 17, n. 5, p. 443-450, 1986. ISSN 0046-8177.

CLARK, W. H. et al. The histogenesis and biologic behavior of primary human malignant melanomas of the skin. **Cancer research**, v. 29, n. 3, p. 705, 1969. ISSN 0008-5472.

CLEMENTE, C. G. et al. Prognostic value of tumor infiltrating lymphocytes in the vertical growth phase of primary cutaneous melanoma. **Cancer**, v. 77, n. 7, p. 1303-1310, 1996. ISSN 0008-543X.

COLBURN, W. et al. Biomarkers and surrogate endpoints: Preferred definitions and conceptual framework. Biomarkers Definitions Working Group. **Clinical Pharmacol & Therapeutics**, v. 69, p. 89-95, 2001.

COOKE, M. S. et al. Oxidative DNA damage: mechanisms, mutation, and disease. **The FASEB Journal**, v. 17, n. 10, p. 1195-1214, 2003. ISSN 0892-6638.

COUSSENS, L. M.; WERB, Z. Inflammation and cancer. **Nature**, v. 420, n. 6917, p. 860-867, 2002. ISSN 0028-0836.

CROSS, C. E. et al. Oxygen radicals and human disease. **Annals of internal medicine**, v. 107, n. 4, p. 526-545, 1987. ISSN 0003-4819.

CUMMINS, D. L. et al. Cutaneous malignant melanoma. Mayo Clinic Proceedings, 2006, Elsevier. p.500-507.

CURFS, J. H.; MEIS, J. F.; HOOGKAMP-KORSTANJE, J. A. A primer on cytokines: sources, receptors, effects, and inducers. **Clin Microbiol Rev**, v. 10, n. 4, p. 742-80, Oct 1997. ISSN 0893-8512.

DAMSKY, W. E.; ROSENBAUM, L. E.; BOSENBERG, M. Decoding melanoma metastasis. **Cancers**, v. 3, n. 1, p. 126-163, 2010.

DRANOFF, G. Cytokines in cancer pathogenesis and cancer therapy. **Nat Rev Cancer**, v. 4, n. 1, p. 11-22, Jan 2004. ISSN 1474-175X.

DRANOFF, G. Cytokines in cancer pathogenesis and cancer therapy. **Nature Reviews Cancer**, v. 4, n. 1, p. 11-22, 2004. ISSN 1474-175X.

DUNN, G. P. et al. Cancer immunoediting: from immunosurveillance to tumor escape. **Nature immunology**, v. 3, n. 11, p. 991-998, 2002.

EDEAS, M. Strategies to target mitochondria and oxidative stress by antioxidants: key points and perspectives. **Pharmaceutical research**, v. 28, n. 11, p. 2771-2779, 2011. ISSN 0724-8741.

EGGERMONT, A. M.; SPATZ, A.; ROBERT, C. Cutaneous melanoma. **Lancet**, v. 383, n. 9919, p. 816-27, Mar 1 2014. ISSN 0140-6736.

EL HAJJ, P. et al. Tyrosinase-related protein 1 mRNA expression in lymph node metastases predicts overall survival in high-risk melanoma patients. **Br J Cancer**, v. 108, n. 8, p. 1641-7, Apr 30 2013. ISSN 0007-0920.

EMERIT, I. Reactive oxygen species, chromosome mutation, and cancer: possible role of clastogenic factors in carcinogenesis. **Free Radical Biology and Medicine**, v. 16, n. 1, p. 99-109, 1994. ISSN 0891-5849.

ERDEI, E.; TORRES, S. M. A new understanding in the epidemiology of melanoma. **Expert Rev Anticancer Ther**, v. 10, n. 11, p. 1811-23, Nov 2010. ISSN 1473-7140.

ERDMANN, F. et al. International trends in the incidence of malignant melanoma 1953– 2008—are recent generations at higher or lower risk? **International Journal of Cancer**, Hoboken, v. 132, n. 2, p. 385-400, 2013. ISSN 0020-7136.

FEIG, D. I.; REID, T. M.; LOEB, L. A. Reactive oxygen species in tumorigenesis. **Cancer Research**, v. 54, n. 7 Supplement, p. 1890s-1894s, 1994. ISSN 0008-5472.

FINN, L.; MARKOVIC, S. N.; JOSEPH, R. W. Therapy for metastatic melanoma: the past, present, and future. **BMC medicine**, v. 10, n. 1, p. 1, 2012. ISSN 1741-7015.

FRIEDMAN, E. B. et al. Serum microRNAs as biomarkers for recurrence in melanoma. **J Transl Med**, v. 10, p. 155, 2012. ISSN 1479-5876.

FRIEDMAN, R. J.; RIGEL, D. S.; KOPF, A. W. Early detection of malignant melanoma: The role of physician examination and self- examination of the skin. **CA: A Cancer Journal for Clinicians**, Bristol, v. 35, n. 3, p. 130-151, 1985. ISSN 0007-9235.

GADJEVA, V.; DIMOV, A.; GEORGIEVA, N. Influence of therapy on the antioxidant status in patients with melanoma. **Journal of clinical pharmacy and therapeutics**, v. 33, n. 2, p. 179-185, 2008. ISSN 1365-2710.

GAHAN, P. B. **Circulating DNA: Intracellular and intraorgan messenger?** 1075: 21-33 p. 2006.

GALLAGHER, W. M.; BERGIN, O. E. Multiple markers for melanoma progression regulated by DNA methylation: insights from transcriptomic studies. **Carcinogenesis**, v. 26, n. 11, p. 1856-1867, 2005. ISSN 0143-3334.

GELLER, A. C. et al. Screening, early detection, and trends for melanoma: Current status (2000- 2006) and future directions. **Journal of the American Academy of Dermatology**, v. 57, n. 4, p. 555-572, 2007. ISSN 0190-9622.

GERSHENWALD, J. E.; SOONG, S.-J.; BALCH, C. M. 2010 TNM staging system for cutaneous melanoma... and beyond. **Annals of surgical oncology**, v. 17, n. 6, p. 1475-1477, 2010. ISSN 1068-9265.

GIMOTTY, P. A. et al. A population-based validation of the American Joint Committee on Cancer melanoma staging system. **Journal of Clinical Oncology**, v. 23, n. 31, p. 8065-8075, 2005. ISSN 0732-183X.

GONZALES, R. et al. Superoxide dismutase, catalase, and glutathione peroxidase in red blood cells from patients with malignant diseases. **Cancer research**, v. 44, n. 9, p. 4137-4139, 1984. ISSN 0008-5472.

GRIN, C. M. et al. Accuracy in the clinical diagnosis of malignant melanoma. **Arch Dermatol**, v. 126, n. 6, p. 763-6, Jun 1990. ISSN 0003-987X.

GRIVENNIKOV, S. I.; KARIN, M. Inflammation and oncogenesis: a vicious connection. **Current opinion in genetics & development**, v. 20, n. 1, p. 65-71, 2010. ISSN 0959-437X.

HAASS, N. K.; SMALLEY, K. S. Melanoma biomarkers: current status and utility in diagnosis, prognosis, and response to therapy. **Mol Diagn Ther**, v. 13, n. 5, p. 283-96, 2009. ISSN 1177-1062.

HALLIWELL, B.; GUTTERIDGE, J. M. **Free radicals in biology and medicine**. Oxford University Press, USA, 2015. ISBN 0198717482.

HAN, J.; COLDITZ, G. A.; HUNTER, D. J. Manganese superoxide dismutase polymorphism and risk of skin cancer (United States). **Cancer Causes Control**, v. 18, n. 1, p. 79-89, Feb 2007. ISSN 0957-5243.

HERMAN, C. Emerging technologies for the detection of melanoma: achieving better outcomes. **Clin Cosmet Investig Dermatol**, v. 5, p. 195-212, 2012. ISSN 1178-7015.

HILL, R. et al. TRIB2 as a biomarker for diagnosis and progression of melanoma. **Carcinogenesis**, v. 36, n. 4, p. 469-77, Apr 2015. ISSN 0143-3334.

HODI, F. S. et al. Improved survival with ipilimumab in patients with metastatic melanoma. **New England Journal of Medicine**, v. 363, n. 8, p. 711-723, 2010. ISSN 0028-4793.

HOFMANN, M. A. et al. Diagnostic value of melanoma inhibitory activity serum marker in the follow-up of patients with stage I or II cutaneous melanoma. **Melanoma Res**, v. 19, n. 1, p. 17-23, Feb 2009. ISSN 0960-8931.

HOFMANN, M. A. et al. Impact of lymph node metastases on serum level of melanoma inhibitory activity in stage III melanoma patients. **J Dermatol**, v. 38, n. 9, p. 880-6, Sep 2011. ISSN 0385-2407.

HOSHIMOTO, S. et al. AIM1 and LINE-1 epigenetic aberrations in tumor and serum relate to melanoma progression and disease outcome. **J Invest Dermatol**, v. 132, n. 6, p. 1689-97, Jun 2012. ISSN 0022-202x.

HOUGHTON, A. N.; POLSKY, D. Focus on melanoma. **Cancer Cell**, v. 2, n. 4, p. 275-8, Oct 2002. ISSN 1535-6108.

HSU, M. Y. et al. Shifts in cadherin profiles between human normal melanocytes and melanomas. **J Invest Dermatol Symp Proc**, v. 1, n. 2, p. 188-94, Apr 1996. ISSN 1087-0024.

HUBER, M. A.; KRAUT, N.; BEUG, H. Molecular requirements for epithelial–mesenchymal transition during tumor progression. **Current Opinion in Cell Biology**, v. 17, n. 5, p. 548-558, 2005. ISSN 0955-0674.

HUSSEIN, M. Tumour-infiltrating lymphocytes and melanoma tumorigenesis: an insight. **British Journal of Dermatology**, v. 153, n. 1, p. 18-21, 2005. ISSN 1365-2133.

JAKOB, J. A. et al. NRAS mutation status is an independent prognostic factor in metastatic melanoma. **Cancer**, v. 118, n. 16, p. 4014-23, Aug 15 2012. ISSN 0008-543x.

JIANG, N.; PISETSKY, D. S. The effect of inflammation on the generation of plasma DNA from dead and dying cells in the peritoneum. **Journal of leukocyte biology**, v. 77, n. 3, p. 296, 2005. ISSN 0741-5400.

JOHNSON, D. B.; PENG, C.; SOSMAN, J. A. Nivolumab in melanoma: latest evidence and clinical potential. **Ther Adv Med Oncol**, v. 7, n. 2, p. 97-106, Mar 2015. ISSN 1758-8340.

JUNG, K.; FLEISCHHACKER, M.; RABIEN, A. Cell- free DNA in the blood as a solid tumor biomarker—A critical appraisal of the literature. **Clinica Chimica Acta**, v. 411, n. 21, p. 1611-1624, 2010. ISSN 0009-8981.

JUNG, K. et al. Increased cell- free DNA in plasma of patients with metastatic spread in prostate cancer. **Cancer Letters**, v. 205, n. 2, p. 173-180, 2004. ISSN 0304-3835.

KAMAT, A. A. et al. Plasma cell- free DNA in ovarian cancer: an independent prognostic biomarker.(Clinical report). **Cancer**, v. 116, n. 8, p. 1918, 2010. ISSN 0008-543X.

KAMIYA, H. et al. 8-Hydroxyguanine (7, 8-dihydro-8-oxoguanine) in hot spots of the c-Ha-ras gene: effects of sequence contexts on mutation spectra. **Carcinogenesis**, v. 16, n. 4, p. 883-889, 1995. ISSN 0143-3334.

KARIN, M. Nuclear factor- κ B in cancer development and progression. **Nature**, v. 441, n. 7092, p. 431-436, 2006. ISSN 0028-0836.

KARIN, M.; GRETEN, F. R. NF- κ B: linking inflammation and immunity to cancer development and progression. **Nature Reviews Immunology**, v. 5, n. 10, p. 749-759, 2005. ISSN 1474-1733.

KASAI, H.; NISHIMURA, S. Formation of 8-hydroxydeoxyguanosine in DNA by oxygen radicals and its biological significance. **Oxidative stress: Oxidants and antioxidants**, p. 99-116, 1991.

KIELBASSA, C.; ROZA, L.; EPE, B. Wavelength dependence of oxidative DNA damage induced by UV and visible light. **Carcinogenesis**, v. 18, n. 4, p. 811-816, 1997. ISSN 0143-3334.

KURMI, O. et al. Oxidative potential of smoke from burning wood and mixed biomass fuels. **Free radical research**, v. 47, n. 10, p. 829-835, 2013. ISSN 1071-5762.

KVAM, E.; TYRRELL, R. M. Induction of oxidative DNA base damage in human skin cells by UV and near visible radiation. **Carcinogenesis**, v. 18, n. 12, p. 2379-2384, 1997. ISSN 0143-3334.

LA PORTA, C. A. M. Drug Resistance in Melanoma: New Perspectives. **Current Medicinal Chemistry**, v. 14, n. 4, p. 387-391, 2007. ISSN 0929-8673.

LANGLEY, R. G. B.; SOBER, A. J. Clinical recognition of melanoma and its precursors. **Hematology/Oncology Clinics of North America**, v. 12, n. 4, p. 699-715, 1998. ISSN 0889-8588.

LARSON, A. R.; KONAT, E.; ALANI, R. M. Melanoma biomarkers: current status and vision for the future. **Nat Clin Pract Oncol**, v. 6, n. 2, p. 105-17, Feb 2009. ISSN 1743-4254.

LENS, M. B.; DAWES, M. **Global perspectives of contemporary epidemiological trends of cutaneous malignant melanoma**. Oxford, UK. 150: 179-185 p. 2004.

LESLIE, M. C.; BAR-ELI, M. Regulation of gene expression in melanoma: new approaches for treatment. **Journal of cellular biochemistry**, v. 94, n. 1, p. 25-38, 2005. ISSN 1097-4644.

LI, C.-N. et al. Cell- free DNA is released from tumor cells upon cell death: a study of tissue cultures of tumor cell lines. **Journal of clinical laboratory analysis**, v. 17, n. 4, p. 103, 2003. ISSN 0887-8013.

LI, H. et al. Manganese superoxide dismutase polymorphism, prediagnostic antioxidant status, and risk of clinical significant prostate cancer. **Cancer research**, v. 65, n. 6, p. 2498-2504, 2005. ISSN 0008-5472.

LI, N. et al. Overexpression of manganese superoxide dismutase in DU145 human prostate carcinoma cells has multiple effects on cell phenotype. **The Prostate**, v. 35, n. 3, p. 221-233, 1998. ISSN 1097-0045.

LIN, E.; CALVANO, S. E.; LOWRY, S. F. Inflammatory cytokines and cell response in surgery. **Surgery**, v. 127, n. 2, p. 117-126, 2000. ISSN 0039-6060.

LIN, E.; CALVANO, S. E.; LOWRY, S. F. Inflammatory cytokines and cell response in surgery. **Surgery**, v. 127, n. 2, p. 117-26, Feb 2000. ISSN 0039-6060.

MARNETT, L. J. Oxyradicals and DNA damage. **Carcinogenesis**, v. 21, n. 3, p. 361-370, 2000. ISSN 0143-3334.

MCGOVERN, V. J. et al. The classification of malignant melanoma and its histologic reporting. **Cancer**, New York, v. 32, n. 6, p. 1446-1457, 1973. ISSN 0008-543X.

MEYER, S. et al. A Seven- Marker Signature and Clinical Outcome in Malignant Melanoma: A Large- Scale Tissue- Microarray Study with Two Independent Patient Cohorts (Seven- Marker Signature in Malignant Melanoma). **PLoS ONE**, San Francisco, USA, v. 7, n. 6, p. e38222, 2012.

MILLER, A. J.; MIHM, M. C., JR. Melanoma. **N Engl J Med**, v. 355, n. 1, p. 51-65, Jul 6 2006. ISSN 0028-4793.

MITRUNEN, K. et al. Association between manganese superoxide dismutase (MnSOD) gene polymorphism and breast cancer risk. **Carcinogenesis**, v. 22, n. 5, p. 827-829, 2001. ISSN 0143-3334.

MORIYA, M. Single-stranded shuttle phagemid for mutagenesis studies in mammalian cells: 8-oxoguanine in DNA induces targeted GC--> TA transversions in simian kidney cells. **Proceedings of the National Academy of Sciences**, v. 90, n. 3, p. 1122-1126, 1993. ISSN 0027-8424.

MURALI, R. et al. Factors predicting recurrence and survival in sentinel lymph node-positive melanoma patients. **Annals of surgery**, v. 253, n. 6, p. 1155-1164, 2011. ISSN 0003-4932.

MURALI, R. et al. Clinicopathologic features of incident and subsequent tumors in patients with multiple primary cutaneous melanomas. **Annals of surgical oncology**, v. 19, n. 3, p. 1024-1033, 2012. ISSN 1068-9265.

NAITO, Y. et al. CD8+ T cells infiltrated within cancer cell nests as a prognostic factor in human colorectal cancer. **Cancer research**, v. 58, n. 16, p. 3491-3494, 1998. ISSN 0008-5472.

NAKANO, O. et al. Proliferative activity of intratumoral CD8+ T-lymphocytes as a prognostic factor in human renal cell carcinoma clinicopathologic demonstration of antitumor immunity. **Cancer research**, v. 61, n. 13, p. 5132-5136, 2001. ISSN 0008-5472.

NISHIGORI, C.; HATTORI, Y.; TOYOKUNI, S. Role of reactive oxygen species in skin carcinogenesis. **Antioxidants and Redox Signaling**, v. 6, n. 3, p. 561-570, 2004. ISSN 1523-0864.

OBERLEY, L. W.; BUETTNER, G. R. Role of superoxide dismutase in cancer: a review. **Cancer research**, v. 39, n. 4, p. 1141-1149, 1979. ISSN 0008-5472.

OCHSENBEIN, A. F. et al. Roles of tumour localization, second signals and cross priming in cytotoxic T-cell induction. **Nature**, v. 411, n. 6841, p. 1058-1064, 2001. ISSN 0028-0836.

OLIVEIRA, C. M. B. D. et al. Citocinas e dor. **Revista Brasileira de Anestesiologia**, 2011. ISSN 0034-7094.

OTT, P. A.; HODI, F. S.; ROBERT, C. CTLA-4 and PD-1/PD-L1 blockade: new immunotherapeutic modalities with durable clinical benefit in melanoma patients. **Clin Cancer Res**, v. 19, n. 19, p. 5300-9, Oct 01 2013. ISSN 1078-0432.

PATHAK, A. K. et al. Circulating cell-free DNA in plasma/serum of lung cancer patients as a potential screening and prognostic tool. **Clin Chem**, v. 52, n. 10, p. 1833-42, Oct 2006. ISSN 0009-9147.

PEHAMBERGER, H.; STEINER, A.; WOLFF, K. In vivo epiluminescence microscopy of pigmented skin lesions. I. Pattern analysis of pigmented skin lesions. **J Am Acad Dermatol**, v. 17, n. 4, p. 571-83, Oct 1987. ISSN 0190-9622.

PFANNER, N.; GEISSLER, A. Versatility of the mitochondrial protein import machinery. **Nature Reviews Molecular Cell Biology**, v. 2, n. 5, p. 339-349, 2001. ISSN 1471-0072.

PHAN, A. et al. Acral lentiginous melanoma: histopathological prognostic features of 121 cases. **British Journal of Dermatology**, Oxford, UK, v. 157, n. 2, p. 311-318, 2007. ISSN 0007-0963.

POSER, I. et al. Loss of E- cadherin expression in melanoma cells involves up-regulation of the transcriptional repressor Snail. **The Journal of biological chemistry**, v. 276, n. 27, p. 24661, 2001. ISSN 0021-9258.

POSWIG, A. et al. Adaptive antioxidant response of manganese-superoxide dismutase following repetitive UVA irradiation. **Journal of investigative dermatology**, v. 112, n. 1, p. 13-18, 1999. ISSN 0022-202X.

QIN, Y. et al. Identification of unique sensitizing targets for anti-inflammatory CDDO-Me in metastatic melanoma by a large-scale synthetic lethal RNAi screening. **Pigment Cell & Melanoma Research**, v. 26, n. 1, p. 97-112, 2013. ISSN 1755-1471.

RASKIN, L. et al. Transcriptome profiling identifies HMGA2 as a biomarker of melanoma progression and prognosis. **J Invest Dermatol**, v. 133, n. 11, p. 2585-92, Nov 2013. ISSN 0022-202x.

RASS, K.; HASSEL, J. C. Chemotherapeutics, chemoresistance and the management of melanoma. **G Ital Dermatol Venereol**, v. 144, n. 1, p. 61-78, Feb 2009. ISSN 0392-0488.

RATZINGER, G. et al. Association of TNFRSF10D DNA-methylation with the survival of melanoma patients. **Int J Mol Sci**, v. 15, n. 7, p. 11984-95, 2014. ISSN 1422-0067.

REN, N. et al. Circulating DNA level is negatively associated with the long-term survival of hepatocellular carcinoma patients. **World Journal of Gastroenterology**, v. 12, n. 24, p. 3911-3914, 2006. ISSN 10079327.

RICHMOND, A. et al. Molecular characterization and chromosomal mapping of melanoma growth stimulatory activity, a growth factor structurally related to beta-thromboglobulin. **The EMBO journal**, v. 7, n. 7, p. 2025, 1988.

RIGEL, D. S. Epiluminescence microscopy in clinical diagnosis of pigmented skin lesions? **Lancet**, v. 349, n. 9065, p. 1566-7, May 31 1997. ISSN 0140-6736.

ROSENBLUM, J. S.; GILULA, N. B.; LERNER, R. A. On signal sequence polymorphisms and diseases of distribution. **Proceedings of the National Academy of Sciences**, v. 93, n. 9, p. 4471-4473, 1996. ISSN 0027-8424.

SALDANHA, G. et al. microRNA-10b is a prognostic biomarker for melanoma. **Mod Pathol**, v. 29, n. 2, p. 112-21, Feb 2016. ISSN 0893-3952.

SANDER, C. et al. Oxidative stress in malignant melanoma and non-melanoma skin cancer. **British Journal of Dermatology**, v. 148, n. 5, p. 913-922, 2003. ISSN 1365-2133.

SANDER, C. S. et al. Role of oxidative stress and the antioxidant network in cutaneous carcinogenesis. **International journal of dermatology**, v. 43, n. 5, p. 326-335, 2004. ISSN 1365-4632.

SCHRAMM, S. J.; MANN, G. J. Melanoma prognosis: a REMARK-based systematic review and bioinformatic analysis of immunohistochemical and gene microarray studies. **Mol Cancer Ther**, v. 10, n. 8, p. 1520-8, Aug 2011. ISSN 1535-7163.

SCHWARTZ, J. L.; ANTONIADES, D. Z.; ZHAO, S. Molecular and biochemical reprogramming of oncogenesis through the activity of prooxidants and antioxidants. **Annals of the New York Academy of Sciences**, v. 686, n. 1, p. 262-278, 1993. ISSN 1749-6632.

SCHWARTZENTRUBER, D. J. Guidelines for the safe administration of high-dose interleukin-2. **Journal of immunotherapy**, v. 24, n. 4, p. 287-293, 2001. ISSN 1524-9557.

SIEGEL, R. et al. Cancer statistics, 2014. **CA: A Cancer Journal for Clinicians**, v. 64, n. 1, p. 9-29, 2014. ISSN 0007-9235.

SIEGEL, R.; NAISHADHAM, D.; JEMAL, A. Cancer statistics, 2013. **CA: a cancer journal for clinicians**, v. 63, n. 1, p. 11-30, 2013. ISSN 1542-4863.

SLANGER, T. E.; CHANG-CLAUDE, J.; WANG-GOHRKE, S. Manganese superoxide dismutase Ala-9Val polymorphism, environmental modifiers, and risk of breast cancer in a German population. **Cancer Causes & Control**, v. 17, n. 8, p. 1025-1031, 2006. ISSN 0957-5243.

SLOMINSKI, A. T.; CARLSON, J. A. Melanoma resistance: a bright future for academicians and a challenge for patient advocates. **Mayo Clin Proc**, v. 89, n. 4, p. 429-33, Apr 2014. ISSN 0025-6196.

SMYTH, M. J. et al. Cytokines in cancer immunity and immunotherapy. **Immunological reviews**, v. 202, n. 1, p. 275-293, 2004. ISSN 1600-065X.

SOLOMON, C. et al. Melanoma and lifetime UV radiation. **Cancer Causes Control**, Dordrecht, v. 15, n. 9, p. 893-902, 2004. ISSN 0957-5243.

SONDAK, V.; ROSS, M.; SCHUCHTER, L. Early stage (I, II, III) melanoma. **Curr. Treat. Options in Oncol.**, Philadelphia, v. 2, n. 3, p. 183-191, 2001. ISSN 1527-2729.

SOSA, V. et al. Oxidative stress and cancer: an overview. **Ageing research reviews**, v. 12, n. 1, p. 376-390, 2013. ISSN 1568-1637.

SPATZ, A.; BATIST, G.; EGGERMONT, A. M. The biology behind prognostic factors of cutaneous melanoma. **Curr Opin Oncol**, v. 22, n. 3, p. 163-8, May 2010. ISSN 1040-8746.

SPINDLER, K. L. et al. Circulating free DNA as biomarker and source for mutation detection in metastatic colorectal cancer. **PLoS One**, v. 10, n. 4, p. e0108247, 2015. ISSN 1932-6203.

STROUN, M. et al. About the possible origin and mechanism of circulating DNA: Apoptosis and active DNA release. **Clin Chim Acta**, v. 313, p. 139-142 p. 2001.

STROUN, M. et al. The origin and mechanism of circulating DNA. **Circulating Nucleic Acids In Plasma Or Serum**, v. 906, p. 161-168, 2000. ISSN 0077-8923.

SULLIVAN, L. B.; CHANDEL, N. S. Mitochondrial reactive oxygen species and cancer. **Cancer & metabolism**, v. 2, n. 1, p. 1, 2014. ISSN 2049-3002.

SUTTON, A. et al. The Ala16Val genetic dimorphism modulates the import of human manganese superoxide dismutase into rat liver mitochondria. **Pharmacogenetics and Genomics**, v. 13, n. 3, p. 145-157, 2003. ISSN 1744-6872.

THEIR, J. P. Epithelial- mesenchymal transitions in tumor progression. **Nature Reviews Cancer**, v. 2, n. 6, p. 442-454, 2002. ISSN 1474175X.

THIERY, J. P. [Epithelial-mesenchymal transitions in cancer onset and progression]. **Bull Acad Natl Med**, v. 193, n. 9, p. 1969-78; discussion 1978-9, Dec 2009. ISSN 0001-4079.

TIKOO, S.; HAASS, N. K. Friends or foes: IL- 10 and TGF- β in melanoma. **Experimental Dermatology**, v. 24, n. 4, p. 254-255, 2015. ISSN 0906-6705.

TSAO, H. et al. Early detection of melanoma: Reviewing the ABCDEs. **Journal of the American Academy of Dermatology**, v. 72, n. 4, p. 717-723, 2015. ISSN 0190-9622.

ULIVI, P.; SILVESTRINI, R. Role of quantitative and qualitative characteristics of free circulating DNA in the management of patients with non-small cell lung cancer. **Cell Oncol (Dordr)**, v. 36, n. 6, p. 439-48, Dec 2013. ISSN 2211-3428.

VAN DER VAART, M.; PRETORIUS, P. J. Is the role of circulating DNA as a biomarker of cancer being prematurely overrated? **Clinical Biochemistry**, v. 43, n. 1, p. 26-36, 2010. ISSN 0009-9120.

WALLACE, D. C. Mitochondrial diseases in man and mouse. **Science**, v. 283, n. 5407, p. 1482-1488, 1999. ISSN 0036-8075.

WAN, X. S.; DEVALARAJA, M. N.; ST. CLAIR, D. K. Molecular structure and organization of the human manganese superoxide dismutase gene. **DNA and cell biology**, v. 13, n. 11, p. 1127-1136, 1994. ISSN 1044-5498.

WOLF, I. H. et al. Locoregional cutaneous metastases of malignant melanoma and their management. **Dermatol Surg**, v. 30, n. 2 Pt 2, p. 244-7, Feb 2004. ISSN 1076-0512.

WOODSON, K. et al. Manganese superoxide dismutase (MnSOD) polymorphism, α -tocopherol supplementation and prostate cancer risk in the Alpha-Tocopherol, Beta-Carotene Cancer Prevention Study (Finland). **Cancer Causes & Control**, v. 14, n. 6, p. 513-518, 2003. ISSN 0957-5243.

XU, Y. et al. Differential expression of microRNAs during melanoma progression: miR-200c, miR-205 and miR-211 are downregulated in melanoma and act as tumour suppressors. **Br J Cancer**, v. 106, n. 3, p. 553-61, Jan 31 2012. ISSN 0007-0920.

YOON, K. A. et al. Comparison of circulating plasma DNA levels between lung cancer patients and healthy controls. **J Mol Diagn**, v. 11, n. 3, p. 182-5, May 2009. ISSN 1525-1578.

YU, H.; PARDOLL, D.; JOVE, R. STATs in cancer inflammation and immunity: a leading role for STAT3. **Nature Reviews Cancer**, v. 9, n. 11, p. 798-809, 2009. ISSN 1474-175X.

ZHANG, J.-M.; AN, J. Cytokines, inflammation and pain. **International anesthesiology clinics**, v. 45, n. 2, p. 27, 2007.

ZHANG, L. et al. Intratumoral T cells, recurrence, and survival in epithelial ovarian cancer. **New England Journal of Medicine**, v. 348, n. 3, p. 203-213, 2003. ISSN 0028-4793.

2. OBJETIVOS

2.1 Objetivo geral

Avaliar o potencial prognóstico dos níveis de DNA livre circulante (*cfDNA*) como marcador relacionado à inflamação e do polimorfismo da superóxido dismutase dependente de manganês (MnSOD) em pacientes com melanoma.

2.2 Objetivos específicos

- a) Determinar a associação entre os níveis de *cfDNA* e o melanoma; através da comparação entre casos (pacientes com melanoma) e controles (indivíduos saudáveis);
- b) Avaliar o impacto do tratamento cirúrgico nos níveis de *cfDNA*;
- c) Analisar a associação entre a concentração de *cfDNA* e as características histopatológicas da doença, bem como fatores de risco reconhecidos para o melanoma;
- d) Validar o status inflamatório dos pacientes com melanoma por meio da quantificação de citocinas pró-inflamatórias e anti-inflamatórias;

- e) Determinar a frequência alélica e genotípica dos alelos Ala e Val do gene SOD2 na população em estudo e comparar estas frequências entre casos e controles;
- f) Correlacionar o genótipo da MnSOD aos fatores prognósticos descritos pela AJCC e fatores de risco para melanoma.

3. ARTIGOS CIENTÍFICOS

3.1 Artigo científico I: “Genetic and epigenetic biomarkers in melanoma prognosis: a last five years update”

Autores: Munique Mendonça Azevedo, Yuri Tomé Machado Strey, Bruna
Rudolfo Faraco, Amanda Carolina Aguiar Weigert, Luciana El Halal Schuch,
Felice Riccardi, Claudia Giuliano Bica

Enviado para publicação na Revista “Melanoma Research”

Genetic and epigenetic biomarkers in melanoma prognosis: a last five years update

Short title: Update on melanoma biomarkers

AUTHORS

Munique Mendonça Azevedo, *MS*¹, Yuri Tomé Machado Strey¹, Bruna Rudolfo Faraco¹, Amanda Carolina Aguiar Weigert¹, Luciana El Halal Schuch, *MD*¹, Felice Riccardi, *MD*², *MS*, Claudia Giuliano Bica, *PhD*^{1*}

AFFILIATIONS

¹Universidade Federal de Ciências da Saúde de Porto Alegre (UFCSPA), Porto Alegre, Brazil

²Hospital Santa Rita, Hospital Complex of Santa Casa de Misericórdia de Porto Alegre, Porto Alegre, Brazil

CORRESPONDING AUTHOR AND AUTHOR TO REQUEST REPRINTS

* Claudia G. Bica, PhD, Laboratory of Pathology, Universidade Federal de Ciências da Saúde de Porto Alegre (UFCSPA), 245 Sarmiento Leite, Porto Alegre/RS, Zip Code: 90050-170, Brazil. Tel.: +55(51) 3303-8794. E-mail: claudia@ufcspa.edu.br.

DECLARATIONS OF INTEREST

The authors declare that they have no conflict of interest including any financial, personal or other relationships with other people or organizations that could inappropriately influence, or be perceived to influence, the present work.

SOURCE OF FUNDING

This study was funded by the following research grants and fellowships: CNPq and CAPES.

ABSTRACT

Cutaneous melanoma is an aggressive malignancy due to its propensity to metastasize and intrinsic resistance to treatment. Melanoma is responsible for over 80% of deaths from skin cancer and its incidence is rising faster than any other cancer worldwide. Early-stage melanoma is frequently curable, in contrast, when the disease reaches stage IV, the prognosis is poor. Predicting the behaviour of melanoma in individual patients can be difficult once it has long been recognized as a highly heterogeneous disease. Currently, the general guideline for prognostication is AJCC (American Joint Committee of Cancer) staging system, which is based on clinical and histological parameters. Despite many attempts to identify complementary molecular markers to stratify patients into risk-subgroups in each AJCC, to date, none of them are recommended in clinical assessment. In this review, we analyzed the last five years state-of-the-art in genetic and epigenetic biomarkers in melanoma prognosis. Twenty-five publications have been analyzed. The majority comprises evaluation of microRNA profile, following by studies which compare mRNA expression of different biomarkers with BRAF and NRAS mutational status. Significant insights into melanoma progression have been achieved through genetic and epigenetic analysis, however, further studies are necessary to validate and incorporate prognostic molecular biomarkers in clinical practice.

KEY WORDS: cutaneous melanoma; biomarkers; prognosis; genetic; epigenetic

INTRODUCTION

Cutaneous melanoma represents a highly aggressive malignancy that accounts for only about 4% of all dermatological cancers, but it is responsible for the deaths of more than 80% of skin cancer patients [1]. Globally, the incidence of melanoma has risen faster than any other cancer in white populations in the last years [2,3]. This increase may be due to the tendency for early detection and surveillance, but also to the intense sun exposure [4-6].

Despite early-diagnosed primaries tumors can be cured by surgical excision, once spread to distant sites occurs the prognosis is poor [7]. Melanoma can metastasize very early in the disease process. Approximately 5% of patients with thin lesions (<1.00 mm thick), 25–40% presenting intermediary thickness (lesions between 2–4 mm) and 50–75% with thick lesions (> 4.00 mm thick) develop metastatic disease and die within 10 years [8]. The median overall survival (OS) in stage IV (metastatic disease) is 8–18 months [9] and only 15% live more than 5 years [10].

Treatment options for advanced melanoma are historically limited because it is considered a refractory disease [11]. The recent development of new therapies such as MAP-Kinase inhibitors and the immune checkpoint antibodies (anti-CTLA-4, anti-PD1/PDL1) [12-14] have prolonged life for a few months, but they have some limitations and the prognosis still poor [15].

The staging system proposed by American Joint Committee on Cancer [9] is currently used for melanoma classification, prognostic prediction and

management of patients. The system is based in histopathological features such as Breslow thickness, presence of ulceration and mitotic rate, as well the presence of nodal and/or systemic metastases to classify patients with primary melanoma into stages I/II and individuals with regional/distant metastases into stages III/IV, respectively. Also, serum LDH levels have been taken into account to stratify patients into the AJCC Stages [9].

However, predicting the behaviour of melanoma in individual patients can be difficult due to the disease heterogeneity [16,17]. There are a small subset of tumors exhibiting aggressive behaviour that are not identified by any of AJCC predictors, which limit the capability to screening and early treatment to patients at higher risk of metastasis [18,19]. This warrants the development of complementary biomarkers with prognostic significance to stratify patients into risk-subgroups in each AJCC stage as an attempt to improve outcomes [7]. Various studies have sought to assess molecular biomarkers of melanoma progression, but to date, none of them are recommended in clinical assessment [20].

Molecular biology of melanoma is complex and over 85,000 coding mutations have been reported [21]. Over the last years, melanoma research has been focused in underlying genetic alterations involved in acquisition of metastatic potential by tumor cells [22]. In this regard, determining genetic expression profiles could be utilized to evaluate risk of disease progression and patient survival in primary melanoma [23-25]. In addition to genetic, epigenetic alterations recently have provided new insights regarding to the melanoma progression [26].

In this review, we focus on studies published in the last 5 years that evaluated genetic and epigenetic biomarkers with predictive value for melanoma prognostication.

METHODOLOGY

An active virtual search of medical literature has been conducted on studies evaluating studies on genetic and epigenetic biomarkers for melanoma, published in the period between 2012 and 2016. We searched MEDLINE and COCHRANE LIBRARY using different combinations of the following descriptors: "Melanoma", "Melanomas", "Malignant Melanoma", "Malignant Melanomas", "Biological Markers", "Biomarkers", "Genetic Marker", "DNA Markers", "DNA Marker", "Prognosis", "Prognoses" and "Prognostic".

Initially, sorting of articles was conducted based on title followed by abstract and finally full texts were reviewed if they were categorized as relevant reports. We did not restrict the language in our research.

Studies were included if they met the following criteria for eligibility: having human beings as research subjects; having cutaneous melanoma tissue or blood sample from melanoma patient as biological sample; being an experimental study, suitable for the evaluation of prognostic biomarker; using molecular methods to access genetic and epigenetic alterations. Studies restricted to mucosal or uveal melanoma and in vitro reports, as well as studies whose purpose was to search for potential therapeutic targets or diagnosis biomarkers, have been excluded. The eligibility criteria were assessed

independently by 2 reviewers, and in case of disagreement, a third researcher was consulted.

RESULTS

This review summarizes the current existing genetic and epigenetic biomarkers in cutaneous melanoma patients. A total of 726 publications were retrieved from our electronic search. Of these, 80 abstracts were considered relevant and full texts were reviewed in detail. By the end of the review 25 studies met our inclusion criteria (Figure 1).

Figure 1 here.

In the last 5 years, many studies evaluating candidate biomarkers for melanoma prognostication have been conducted. Most of these markers are implicated in molecular pathways involved in cell proliferation and survival, including mutations in mitogen-activated protein kinase (MAPK) and cell cycle regulators [27]. However, until now, none of them have been recommended for stratification and management of melanoma patients in clinical practice.

In our research, several genetic and epigenetic biomarkers with prognostic significance have been identified and are listed in Table 1 [7,28-51].

Table 1 here.

DISCUSSION

Malignant melanoma is an aggressive skin cancer due to its rapid progression, propensity to metastasize, poor prognosis and intrinsic resistance to treatment. The incidence of melanoma is rising worldwide, being a major public-health problem [52]. Due to the disease heterogeneity, further biomarkers to stratifying melanoma patients and estimating probability of disease progression and survival are needed. Complementary biomarkers with specific prognostic potential could allow for more focused clinical surveillance of patients with increased risk of disease progression. However, until today, there are no validated molecular prognostic markers in melanoma [53].

Despite many attempts to identify complementary biomarkers predicting clinical outcome in melanoma, the majority of them is not robust enough and their utility in routine clinical practice still limited. Therefore, further studies are necessary in order to provide new biomarkers with predictive value.

Combinations of two or more biomarkers to predict prognosis have gained increasing importance in this field and many studies comprising in this review presented multiple biomarkers assays [33-35,43,47-49]. However, some studies still focusing on *BRAF* and *NRAS* polymorphism assessment or making comparisons between the mutational status of this ongenes and microRNA levels or gene expression, for example [36,38,40,46].

Furthermore, we could notice in this review that microRNA expression profile is rising as a promising tool for melanoma prognostication

and, in the last years, the number of publications using this biomarker in the attempt to predict melanoma outcome has been increased [7,45-51].

In addition, we have compiled the markers categories with greatest representation in this search for genetic and epigenetic biomarkers in melanoma prognosis.

MAPK pathway

The mitogen activated protein kinase (MAPK) pathway is one of the most important signaling pathways in regulation of cell proliferation and survival [38]. Mutations in upstream factors such as RAS and RAF lead to a constitutive activation of the MAPK pathway, giving growth advantage to cells in various cancers [54]. A majority of cutaneous melanomas show mutations in genes which encode proteins of MAPK signaling pathway and thus acting in regulation of melanocytes proliferation [55].

Indeed, approximately 40-50% of melanoma cases exhibiting *BRAF* mutations and 15-20% with mutated *NRAS* [56,57]. *BRAF* is a member of the RAF family of enzymes, which plays a critical role in mediating the cellular effects of growth factor signals through receptors on the cell surface to the nucleus [38].

The most prevalent *BRAF* mutation, accounting for ~85% *BRAF* mutations, is *V600E*, a T to A nucleotide transversion which results in a valine to glutamic amino acid substitution [58,59]. This single mutation implicates in a

growth factor-independent proliferation of malignant cells by increasing the catalytic activity of B-Raf [60].

Recent evidences suggest the value of testing *BRAF* mutational status in melanoma to access patient prognosis. Moreau *et al.* [38] reported that patients with *BRAF*-mutated melanoma had worse overall survival and distant metastases-free survival than wild-type *BRAF* melanoma patients [38]. In addition, a meta-analysis showed that *BRAF* mutation is a risk factor for patient survival in melanoma by increasing the risk of mortality in 1.7 times (95% confidence interval, 1.37 to 2.12) [61].

Besides *BRAF* mutations, the protocongene *NRAS* is susceptible to genetic alterations which also activate the MAPK pathway. This gene codes for a guanine triphosphate (GTP)-binding protein and mutations at hotspot in exon 2 (codon 61) lead to loss of GTPase activity and to a permanently activated conformation of RAS [62,63].

Regarding prognostic value of *NRAS*, Jakob *et al.* [40] provided the first evidence that it could be a biomarker for melanoma outcome independent of current validated prognostic markers. The results of this analysis pointing to a significantly shorter survival from patients diagnosed in stage IV of melanoma with an activating mutation in the *NRAS* gene than patients without a mutation in either the *BRAF* or the *NRAS* genes [40].

MicroRNA profile

Over the last few years, there is growing interest in evaluating the prognostic potential of epigenetic biomarkers in the search for melanoma outcome predictors. Epigenetic alterations, defined as changes in gene expression without modifications in DNA sequence, have been implicated in the pathogenesis of various human cancers, including melanoma [18]. These events are potentially reversible, occur at greater frequency than genetic mutations and they consist of DNA methylation, histone modifications and non-coding RNA [26,64]. Regarding DNA methylation in melanoma, hypomethylation of large areas across genome, for example, is recognized as the responsible to increased expression of genes normally silenced during development [65].

Another epigenetic mechanism becoming the focus in melanoma research is microRNA (miRNA), playing an important role in the regulation of protein expression [60]. MicroRNAs are a large family of short non-coding single-stranded RNAs (approximately 22 nucleotides), evolutionarily conserved, that negatively regulate gene expression at the posttranscriptional level [66] and act in cell differentiation, metabolism, and apoptosis [60]. Gene regulation occurred by base pairing within the 3'-untranslated region of messenger mRNA of its target gene to specify degradation or repression of translation [66]. Many studies currently demonstrated the miRNAs implication in tumorigenesis process – targeting oncogenes and tumor suppressors [67] – as well their relevance in melanoma progression [68]. miRNAs can be deregulated and

change their expression profiles in tissue and/or blood samples upon development and progression of cancer, leaving specific signatures [69,70].

The evidence thus far shows that miRNAs could represent a potential prognostic biomarker in melanoma, since they have technical advantages like stability, being resistant to RNase digestion [71], and they are not susceptible to degradation in FFPE samples because of their very small size [3]. Actually, the miRNA most differentially expressed between normal melanocytes and primary melanomas from patients identified is miR-211, which is encoded by an intronic region of TRPM1, a candidate suppressor of melanoma metastasis [72,73].

CONCLUSIONS

In the last decades, there was an advance concerning molecular investigations of melanoma biology. Biomarkers in oncology that provide useful tools of molecular biology to characterize cancer signatures are crucial to personalized treatment. Development of a prognostic assay that could identify with a high risk of relapse will be highly valuable for melanoma management and beneficial for melanoma patients. Significant insights into melanoma progression have been achieved through genetic and epigenetic analysis, however, further studies are necessary to validate and incorporate prognostics molecular biomarkers in clinical practice.

ACKNOWLEDGEMENTS

The authors are grateful for the help of the Laboratory of Pathology (Universidade Federal de Ciências da Saúde de Porto Alegre, Porto Alegre, Brazil).

REFERENCES

1. Miller AJ, Mihm MC, Jr. Melanoma. *N Engl J Med* 2006; 355 (1):51-65.
2. Giblin AV, Thomas JM. Incidence, mortality and survival in cutaneous melanoma. *J Plast Reconstr Aesthet Surg* 2007; 60 (1):32-40.
3. Xu Y, Brenn T, Brown ER, Doherty V, Melton DW. Differential expression of microRNAs during melanoma progression: miR-200c, miR-205 and miR-211 are downregulated in melanoma and act as tumour suppressors. *Br J Cancer* 2012; 106 (3):553-561.
4. Siegel R, Ma J, Zou Z, Jemal A. Cancer statistics, 2014. *CA: A Cancer Journal for Clinicians* 2014; 64 (1):9-29.
5. Solomon C, White E, Kristal A, Vaughan T. Melanoma and lifetime UV radiation. *Cancer Causes Control* 2004; 15 (9):893-902.
6. Lens MB, Dawes M. Global perspectives of contemporary epidemiological trends of cutaneous malignant melanoma. Oxford, UK2004. pp. 179-185.

7. Saldanha G, Elshaw S, Sachs P, Alharbi H, Shah P, Jothi A, et al. microRNA-10b is a prognostic biomarker for melanoma. *Mod Pathol* 2016; 29 (2):112-121.
8. Balch CM, Buzaid AC, Soong SJ, Atkins MB, Cascinelli N, Coit DG, et al. Final version of the American Joint Committee on Cancer staging system for cutaneous melanoma. *J Clin Oncol* 2001; 19 (16):3635-3648.
9. Balch CM, Gershenwald JE, Soong SJ, Thompson JF, Atkins MB, Byrd DR, et al. Final version of 2009 AJCC melanoma staging and classification. *J Clin Oncol* 2009; 27 (36):6199-6206.
10. Siegel R, Naishadham D, Jemal A. Cancer statistics, 2013. *CA: a cancer journal for clinicians* 2013; 63 (1):11-30.
11. Johnson DB, Peng C, Sosman JA. Nivolumab in melanoma: latest evidence and clinical potential. *Ther Adv Med Oncol* 2015; 7 (2):97-106.
12. Chapman PB, Hauschild A, Robert C, Haanen JB, Ascierto P, Larkin J, et al. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. *N Engl J Med* 2011; 364 (26):2507-2516.
13. Flaherty KT, Robert C, Hersey P, Nathan P, Garbe C, Milhem M, et al. Improved survival with MEK inhibition in BRAF-mutated melanoma. *N Engl J Med* 2012; 367 (2):107-114.

14. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 2010; 363 (8):711-723.
15. Slominski AT, Carlson JA. Melanoma resistance: a bright future for academicians and a challenge for patient advocates. *Mayo Clin Proc* 2014; 89 (4):429-433.
16. Schramm SJ, Mann GJ. Melanoma prognosis: a REMARK-based systematic review and bioinformatic analysis of immunohistochemical and gene microarray studies. *Mol Cancer Ther* 2011; 10 (8):1520-1528.
17. Spatz A, Batist G, Eggermont AM. The biology behind prognostic factors of cutaneous melanoma. *Curr Opin Oncol* 2010; 22 (3):163-168.
18. Abbas O, Miller DD, Bhawan J. Cutaneous malignant melanoma: update on diagnostic and prognostic biomarkers. *Am J Dermatopathol* 2014; 36 (5):363-379.
19. Dye DE, Medic S, Ziman M, Coombe DR. Melanoma Biomolecules: Independently Identified but Functionally Intertwined. *Front Oncol* 2013; 3:252.
20. Gould Rothberg BE, Rimm DL. Biomarkers: the useful and the not so useful--an assessment of molecular prognostic markers for cutaneous melanoma. *J Invest Dermatol* 2010; 130 (8):1971-1987.

21. Lawrence MS, Stojanov P, Polak P, Kryukov GV, Cibulskis K, Sivachenko A, et al. Mutational heterogeneity in cancer and the search for new cancer-associated genes. *Nature* 2013; 499 (7457):214-218.
22. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011; 144 (5):646-674.
23. Kabbarah O, Nogueira C, Feng B, Nazarian RM, Bosenberg M, Wu M, et al. Integrative genome comparison of primary and metastatic melanomas. *PLoS One* 2010; 5 (5):e10770.
24. Riker AI, Enkemann SA, Fodstad O, Liu S, Ren S, Morris C, et al. The gene expression profiles of primary and metastatic melanoma yields a transition point of tumor progression and metastasis. *BMC Med Genomics* 2008; 1:13.
25. Gould Rothberg BE, Bracken MB, Rimm DL. Tissue biomarkers for prognosis in cutaneous melanoma: a systematic review and meta-analysis. *J Natl Cancer Inst* 2009; 101 (7):452-474.
26. de Unamuno B, Palanca S, Botella R. Update on melanoma epigenetics. *Curr Opin Oncol* 2015; 27 (5):420-426.
27. Takata M, Saida T. Genetic alterations in melanocytic tumors. *J Dermatol Sci* 2006; 43 (1):1-10.

28. Brunner G, Reitz M, Heinecke A, Lippold A, Berking C, Suter L, et al. A nine-gene signature predicting clinical outcome in cutaneous melanoma. *J Cancer Res Clin Oncol* 2013; 139 (2):249-258.
29. Rapanotti MC, Suarez Viguria TM, Costanza G, Ricosi I, Pierantozzi A, Di Stefani A, et al. Sequential molecular analysis of circulating MCAM/MUC18 expression: a promising disease biomarker related to clinical outcome in melanoma. *Arch Dermatol Res* 2014; 306 (6):527-537.
30. Raskin L, Fullen DR, Giordano TJ, Thomas DG, Frohm ML, Cha KB, et al. Transcriptome profiling identifies HMGA2 as a biomarker of melanoma progression and prognosis. *J Invest Dermatol* 2013; 133 (11):2585-2592.
31. Reid AL, Millward M, Pearce R, Lee M, Frank MH, Ireland A, et al. Markers of circulating tumour cells in the peripheral blood of patients with melanoma correlate with disease recurrence and progression. *Br J Dermatol* 2013; 168 (1):85-92.
32. El Hajj P, Journe F, Wiedig M, Laios I, Sales F, Galibert MD, et al. Tyrosinase-related protein 1 mRNA expression in lymph node metastases predicts overall survival in high-risk melanoma patients. *Br J Cancer* 2013; 108 (8):1641-1647.
33. Falkenius J, Lundeberg J, Johansson H, Tuominen R, Frostvik-Stolt M, Hansson J, et al. High expression of glycolytic and pigment proteins is

associated with worse clinical outcome in stage III melanoma. *Melanoma Res* 2013; 23 (6):452-460.

34. Ito T, Wada M, Nagae K, Nakano-Nakamura M, Nakahara T, Hagihara A, et al. Triple-marker PCR assay of sentinel lymph node as a prognostic factor in melanoma. *J Eur Acad Dermatol Venereol* 2015; 29 (5):912-918.

35. Lengyel Z, Lovig C, Kommedal S, Keszthelyi R, Szekeres G, Battyani Z, et al. Altered expression patterns of clock gene mRNAs and clock proteins in human skin tumors. *Tumour Biol* 2013; 34 (2):811-819.

36. Mitchell B, Leone D, Feller JK, Bondzie P, Yang S, Park HY, et al. Correlation of chemokine receptor CXCR4 mRNA in primary cutaneous melanoma with established histopathologic prognosticators and the BRAF status. *Melanoma Res* 2014; 24 (6):621-625.

37. Campillo JA, Legaz I, Lopez-Alvarez MR, Bolarin JM, Las Heras B, Muro M, et al. KIR gene variability in cutaneous malignant melanoma: influence of KIR2D/HLA-C pairings on disease susceptibility and prognosis. *Immunogenetics* 2013; 65 (5):333-343.

38. Moreau S, Saiag P, Aegerter P, Bosset D, Longvert C, Helias-Rodzewicz Z, et al. Prognostic value of BRAF(V(6)(0)(0)) mutations in melanoma patients after resection of metastatic lymph nodes. *Ann Surg Oncol* 2012; 19 (13):4314-4321.

39. Nagore E, Heidenreich B, Rachakonda S, Garcia-Casado Z, Requena C, Soriano V, et al. TERT promoter mutations in melanoma survival. *Int J Cancer* 2016.
40. Jakob JA, Bassett RL, Jr., Ng CS, Curry JL, Joseph RW, Alvarado GC, et al. NRAS mutation status is an independent prognostic factor in metastatic melanoma. *Cancer* 2012; 118 (16):4014-4023.
41. Kaehler KC, Politz O, Henderson D, Ulbrich HF, Hauschild A, Mund C, et al. Novel DNA methylation markers with potential prognostic relevance in advanced malignant melanoma identified using COBRA assays. *Melanoma Res* 2015; 25 (3):225-231.
42. Ratzinger G, Mitteregger S, Wolf B, Berger R, Zelger B, Weinlich G, et al. Association of TNFRSF10D DNA-methylation with the survival of melanoma patients. *Int J Mol Sci* 2014; 15 (7):11984-11995.
43. Hoshimoto S, Kuo CT, Chong KK, Takeshima TL, Takei Y, Li MW, et al. AIM1 and LINE-1 epigenetic aberrations in tumor and serum relate to melanoma progression and disease outcome. *J Invest Dermatol* 2012; 132 (6):1689-1697.
44. Wang H, Lee S, Nigro CL, Lattanzio L, Merlano M, Monteverde M, et al. NT5E (CD73) is epigenetically regulated in malignant melanoma and associated with metastatic site specificity. *Br J Cancer* 2012; 106 (8):1446-1452.

45. Margue C, Reinsbach S, Philippidou D, Beaume N, Walters C, Schneider JG, et al. Comparison of a healthy miRNome with melanoma patient miRNomes: are microRNAs suitable serum biomarkers for cancer? *Oncotarget* 2015; 6 (14):12110-12127.
46. Tembe V, Schramm SJ, Stark MS, Patrick E, Jayaswal V, Tang YH, et al. MicroRNA and mRNA expression profiling in metastatic melanoma reveal associations with BRAF mutation and patient prognosis. *Pigment Cell Melanoma Res* 2015; 28 (3):254-266.
47. van Kempen LC, van den Hurk K, Lazar V, Michiels S, Winnepenninckx V, Stas M, et al. Loss of microRNA-200a and c, and microRNA-203 expression at the invasive front of primary cutaneous melanoma is associated with increased thickness and disease progression. *Virchows Arch* 2012; 461 (4):441-448.
48. Fleming NH, Zhong J, da Silva IP, Vega-Saenz de Miera E, Brady B, Han SW, et al. Serum-based miRNAs in the prediction and detection of recurrence in melanoma patients. *Cancer* 2015; 121 (1):51-59.
49. Friedman EB, Shang S, de Miera EV, Fog JU, Teilum MW, Ma MW, et al. Serum microRNAs as biomarkers for recurrence in melanoma. *J Transl Med* 2012; 10:155.
50. Lin N, Zhou Y, Lian X, Tu Y. Expression of microRNA-106b and its clinical significance in cutaneous melanoma. *Genet Mol Res* 2015; 14 (4):16379-16385.

51. Shiiyama R, Fukushima S, Jinnin M, Yamashita J, Miyashita A, Nakahara S, et al. Sensitive detection of melanoma metastasis using circulating microRNA expression profiles. *Melanoma Res* 2013; 23 (5):366-372.
52. Forsea AM, Del Marmol V, de Vries E, Bailey EE, Geller AC. Melanoma incidence and mortality in Europe: new estimates, persistent disparities. *Br J Dermatol* 2012; 167 (5):1124-1130.
53. Rendleman J, Shang S, Dominianni C, Shields JF, Scanlon P, Adaniel C, et al. Melanoma risk loci as determinants of melanoma recurrence and survival. *J Transl Med* 2013; 11:279.
54. Malumbres M, Barbacid M. RAS oncogenes: the first 30 years. *Nat Rev Cancer* 2003; 3 (6):459-465.
55. Jemal A, Siegel R, Ward E, Hao Y, Xu J, Thun MJ. Cancer statistics, 2009. *CA Cancer J Clin* 2009; 59 (4):225-249.
56. Davies H, Bignell GR, Cox C, Stephens P, Edkins S, Clegg S, et al. Mutations of the BRAF gene in human cancer. *Nature* 2002; 417 (6892):949-954.
57. Hocker T, Tsao H. Ultraviolet radiation and melanoma: a systematic review and analysis of reported sequence variants. *Hum Mutat* 2007; 28 (6):578-588.

58. Shinozaki M, Fujimoto A, Morton DL, Hoon DS. Incidence of BRAF oncogene mutation and clinical relevance for primary cutaneous melanomas. *Clinical Cancer Research* 2004; 10 (5):1753-1757.
59. Long GV, Menzies AM, Nagrial AM, Haydu LE, Hamilton AL, Mann GJ, et al. Prognostic and clinicopathologic associations of oncogenic BRAF in metastatic melanoma. *Journal of Clinical Oncology* 2011; 29 (10):1239-1246.
60. Voller D, Ott C, Bosserhoff A. MicroRNAs in malignant melanoma. *Clin Biochem* 2013; 46 (10-11):909-917.
61. Safaee Ardekani G, Jafarnejad SM, Tan L, Saeedi A, Li G. The prognostic value of BRAF mutation in colorectal cancer and melanoma: a systematic review and meta-analysis. *PLoS One* 2012; 7 (10):e47054.
62. Lee JH, Choi JW, Kim YS. Frequencies of BRAF and NRAS mutations are different in histological types and sites of origin of cutaneous melanoma: a meta-analysis. *Br J Dermatol* 2011; 164 (4):776-784.
63. Ruoslahti E, Giancotti FG. Integrins and tumor cell dissemination. *Cancer Cells* 1989; 1 (4):119-126.
64. Richards HW, Medrano EE. Epigenetic marks in melanoma. *Pigment Cell Melanoma Res* 2009; 22 (1):14-29.

65. Rubinstein JC, Tran N, Ma S, Halaban R, Krauthammer M. Genome-wide methylation and expression profiling identifies promoter characteristics affecting demethylation-induced gene up-regulation in melanoma. *BMC Med Genomics* 2010; 3:4.
66. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004; 116 (2):281-297.
67. Ventura A, Jacks T. MicroRNAs and cancer: short RNAs go a long way. *Cell* 2009; 136 (4):586-591.
68. Caramuta S, Egyhazi S, Rodolfo M, Witten D, Hansson J, Larsson C, et al. MicroRNA expression profiles associated with mutational status and survival in malignant melanoma. *J Invest Dermatol* 2010; 130 (8):2062-2070.
69. Healy NA, Heneghan HM, Miller N, Osborne CK, Schiff R, Kerin MJ. Systemic mirnas as potential biomarkers for malignancy. *Int J Cancer* 2012; 131 (10):2215-2222.
70. Weiland M, Gao XH, Zhou L, Mi QS. Small RNAs have a large impact: circulating microRNAs as biomarkers for human diseases. *RNA Biol* 2012; 9 (6):850-859.
71. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanyan EL, et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A* 2008; 105 (30):10513-10518.

72. Mazar J, DeYoung K, Khaitan D, Meister E, Almodovar A, Goydos J, et al. The regulation of miRNA-211 expression and its role in melanoma cell invasiveness. PLoS One 2010; 5 (11):e13779.

73. Levy C, Khaled M, Iliopoulos D, Janas MM, Schubert S, Pinner S, et al. Intronic miR-211 assumes the tumor suppressive function of its host gene in melanoma. Mol Cell 2010; 40 (5):841-849.

LEGEND OF TABLE AND FIGURE

Table 1. Genetic and epigenetic biomarkers in melanoma.

Figure 1. Flowchart representing the process of publications selection based on eligibility criteria.

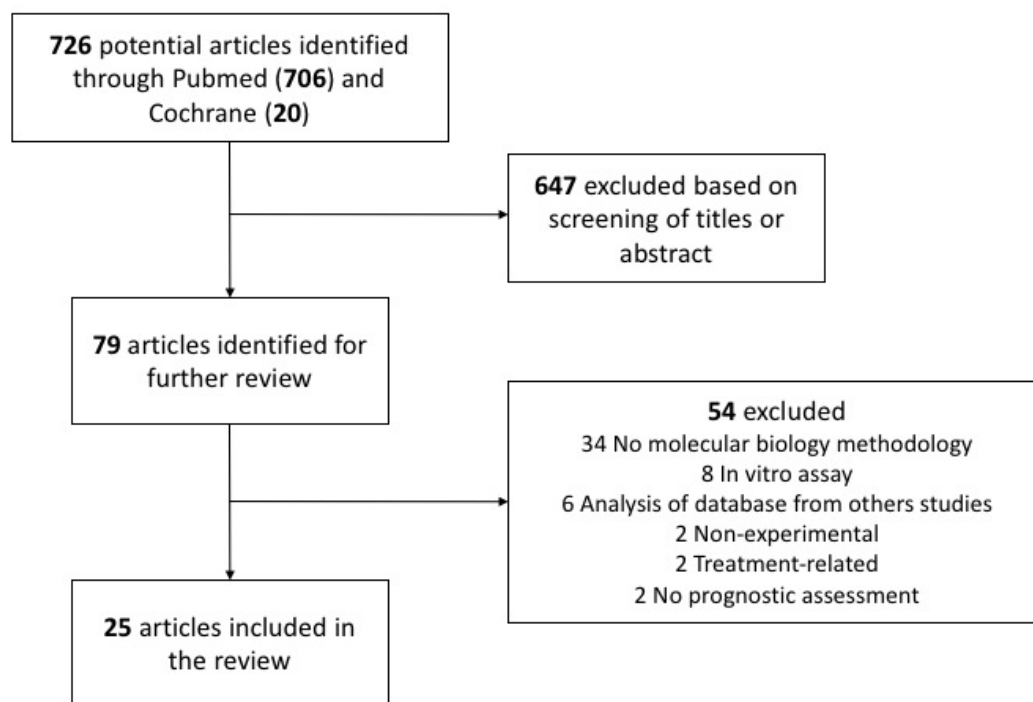


Figure 1. Flowchart representing the process of publications selection based on eligibility criteria.

Table 1. Genetic and epigenetic biomarkers in melanoma.

Biomarker	Description	N	Findings	Reference
<u>mRNA expression</u>				
<i>KRT9</i>	Structural constituent of cytoskeleton	44	Protective gene; expression correlated with OS.	Brunner <i>et al.</i> [28]
<i>DCD</i>	Peptidase activity	44	Protective gene; expression correlated with OS.	Brunner <i>et al.</i> [28]
<i>PIP</i>	IgG binding	44	Protective gene; expression correlated with OS.	Brunner <i>et al.</i> [28]
<i>SCGB1D2</i>	Lipophilin gene	44	Protective gene; expression correlated with OS.	Brunner <i>et al.</i> [28]
<i>SCGB2A2</i>	Secretoglobin	44	Protective gene; expression correlated with OS.	Brunner <i>et al.</i> [28]
<i>COL6A6</i>	Potentially involved in tumor cell invasion	44	Protective gene; expression correlated with OS.	Brunner <i>et al.</i> [28]
<i>KBTBD10</i>	Potentially involved in tumor cell invasion	44	Risk gene; expression inversely correlated with OS.	Brunner <i>et al.</i> [28]
<i>ECRG2</i>	Inhibits tumor cell invasion	44	Risk gene; expression inversely correlated with OS.	Brunner <i>et al.</i> [28]
<i>HES6</i>	Might act as a regulator of melanoma cell differentiation	44	Risk gene; expression inversely correlated with OS.	Brunner <i>et al.</i> [28]
<i>MCAM/MUC18</i>	Cell adhesion	175	Significantly associated with poor prognosis and death, regardless of the AJCC stages; positive expression associated with melanoma progression.	Rapanotti <i>et al.</i> [29]
<i>HMGA2</i>	Transcription factor	421	Transcription factor HMGA2 is up-regulated in primary and metastatic melanoma compared with normal skin; HMGA2 is independently associated with OS, DFS and DMFS.	Raskin <i>et al.</i> [30]
<i>MLANA</i>	Melanoma cell marker	382	Expressed in 45% of patients who experienced disease recurrence.	Reid <i>et al.</i> [31]
<i>ABCB5</i>	Melanoma cell marker	382	Expressed in 49% of patients who experienced disease recurrence. Co-expression of <i>MLANA</i> and <i>ABCB5</i> produced a positive predictive value of 56% and a negative predictive value of 72%.	Reid <i>et al.</i> [31]
<i>TYRP1</i>	Pigment-related	104	<i>TYRP1/S100B</i> ratio correlated with reduced DFS and OS, increased Breslow thickness and presence of ulceration of primaries.	El Hajj <i>et al.</i> [32]
		84	Median mRNA expression was lower for those stage III patients with good outcome compared with those with poor outcome.	Falkenius <i>et al.</i> [33]
<i>GAPDHS</i>	Glycolytic gene	84	A significant difference was observed in <i>GAPDHS</i> expression between short- and long-term survivors (stage III patients); the median mRNA ratio between long- and short-term survivors was 0.19.	Falkenius <i>et al.</i> [33]
<i>GAPDH</i>	Glycolytic gene	84	Median mRNA expression was lower for those stage III patients with good outcome compared with those with poor outcome (the median mRNA ratio between long- and short-term survivors was 0.65).	Falkenius <i>et al.</i> [33]
<i>PKM2</i>	Glycolytic gene	84	Median mRNA expression was lower for those stage III patients with good outcome compared with those with poor outcome (the median mRNA ratio between long- and short-term survivors was 0.78).	Falkenius <i>et al.</i> [33]
Triple-marker: Tyrosinase, GP-100 and MART-1	Melanocyte-related	165	Patients with SLN presenting positivity for the 3 markers had significantly poor MSS and DFS when compared to those with positivity for 1 or 2 markers or PCR negativity.	Ito <i>et al.</i> [34]
<i>PER 1</i>	Clock gene	32	<i>PER1</i> showed significantly decreased levels in melanoma tumors when compared with adjacent nontumorous skin tissues. The magnitude of decrease in mRNA level was 39%.	Lengyel <i>et al.</i> [35]
<i>PER2</i>	Clock gene	32	<i>PER2</i> showed significantly decreased levels in melanoma tumors when compared with adjacent nontumorous skin tissues. The magnitude of decrease in mRNA level was 40%.	Lengyel <i>et al.</i> [35]
<i>CLOCK</i>	Clock gene	32	<i>CLOCK</i> showed significantly decreased levels in melanoma tumors when compared with adjacent nontumorous skin tissues. The magnitude of decrease in mRNA level was 48%. Reciprocal association was found between <i>CLOCK</i> mRNA levels in tumor and Breslow thickness.	Lengyel <i>et al.</i> [35]
<i>CRY1</i>	Clock gene	32	<i>CRY1</i> expression showed reciprocal associations with Breslow thickness and ulceration.	Lengyel <i>et al.</i> [35]
<i>CXCR4</i>	Chemokine receptor activity	87	Correlation between elevated <i>CXCR4</i> expression and the presence of BRAF mutation; <i>CXCR4</i> mRNA was significantly lower in AJCC stage 2 compared with stage 1; absence of a brisk host response was associated with <i>CXCR4</i> mRNA expression.	Mitchell <i>et al.</i> [36]
<u>Genetic polymorphism</u>				
<i>KIR</i>	Immune protection	387	<i>KIR2DL3/C1</i> genotype as a protective marker for melanoma patients and for patients with SLN metastasis; <i>KIR2DL1(+)/S1(-)/C2C2</i> genotype associated with susceptibility to melanoma and SLN metastasis.	Campillo <i>et al.</i> [37]
<i>BRAF</i>	Oncogene	105	Death occurred in 83.3% (OS of 1.4 years) and 60.3% (OS of 2.8 years) of stage III patients with and without BRAF mutations, respectively.	Moreau <i>et al.</i> [38]
<i>TERT</i> promoter	Telomerase RNA reverse transcriptase activity	300	<i>TERT</i> promoter mutations were associated with histopathological features of tumor; patients with <i>TERT</i> mutations had shorter DFS than patients without those mutations; poor MSS compared to those without <i>TERT</i> mutations.	Nagore <i>et al.</i> [39]

Table 1. (continued)

Biomarker	Description	N	Findings	Reference
<i>NRAS</i>	Oncogene	677	Patients with <i>NRAS</i> mutations had a median survival of 8.2 months from stage IV diagnosis (wild type patients = 15.1 months); <i>NRAS</i> mutations are associated with decreased OS independently of age, sex, metastases category and LDH level.	Jakob <i>et al.</i> [40]
<u>DNA methylation</u>				
<i>CDH11</i>	Cell-cell adhesion	28	Increased values either in <i>CDH11</i> or age lead to a shorter expected time to death in metastatic melanoma patients.	Kaehler <i>et al.</i> [41]
<i>BASP1</i>	Membrane bound	28	Increased <i>BASP1</i> values seem to correlate with time to death in metastatic melanoma patients.	Kaehler <i>et al.</i> [41]
<i>TNFRSF10D</i>	Member of the TNF-receptor superfamily	68	<i>TNFRSF10D</i> DNA methylation status significantly associated with poor OS and RFS.	Ratzinger <i>et al.</i> [42]
<i>LINE-1</i>	Retrotransposable element	113	<i>LINE-1</i> U index (hypomethylation) level was significantly higher in melanoma patients than that of normal skin and nevi, and was elevated with increasing AJCC stage; associated with DFS and OS in stage I/II patients.	Hoshimoto <i>et al.</i> [43]
<i>AIM1</i>	Tumor-suppressor gene	92	<i>AIM1</i> promoter hypermethylation was found in higher frequency in metastatic than in primary melanoma; associated with DFS and OS in stage I/II patients.	Hoshimoto <i>et al.</i> [43]
<i>NT5E</i>	5'-nucleotidase activity	52	Relapse with metastatic disease is more common in primary melanomas lacking <i>NT5E</i> methylation.	Wang <i>et al.</i> [44]
<u>microRNA profile</u>				
miR-10B	N/A	79	Increased expression of miR-10B (3.7-fold) in thick primaries melanomas with metastasis (cases) compared with thick primaries melanomas non metastatic (controls) and a trend of increasing expression between primaries tumors and their matched metastases.	Saldanha <i>et al.</i> [7]
Cell-free miRNAs	N/A	126	Several miRNAs were down-regulated in late stage melanoma patients (e.g. miR-200c-3p, -204-5p, -182-5p, -301a-3p) while others were up-regulated (e.g. miR-211-5p, -193b-3p, -720, -205-5p).	Margue <i>et al.</i> [45]
miR-142-3p	N/A	84	Biomarker expression >1.5-fold in good compared with poor prognosis patients; expression was significantly lower in patients with stage IV compared with stage III of the disease.	Tembe <i>et al.</i> [46]
miR-142-5p	N/A	84	Biomarker expression >1.5-fold in good compared with poor prognosis patients	Tembe <i>et al.</i> [46]
miR-150-5p	N/A	84	Biomarker expression >1.5-fold in good compared with poor prognosis patients; reduced expression in BRAF mutation positive tumors.	Tembe <i>et al.</i> [46]
miR-200a	N/A	23	Correlation between tumor thickness and loss of expression of miR-200a, miR-200c and miR-203, and their progressive loss in a series of matched primary tumors and associated metastases suggests a possible association with disease progression.	van Kempen <i>et al.</i> [47]
miR-200c	N/A	23	Correlation between tumor thickness and loss of expression of miR-200a, miR-200c and miR-203, and their progressive loss in a series of matched primary tumors and associated metastases suggests a possible association with disease progression.	van Kempen <i>et al.</i> [47]
miR-203	N/A	23	Correlation between tumor thickness and loss of expression of miR-200a, miR-200c and miR-203, and their progressive loss in a series of matched primary tumors and associated metastases suggests a possible association with disease progression.	van Kempen <i>et al.</i> [47]
miR-150	N/A	283	Expression combined with stage separated patients by RFS and Os and improved prediction of recurrence over stage alone.	Fleming <i>et al.</i> [48]
miR-30b	N/A	283	Expression combined with stage separated patients by RFS and Os and improved prediction of recurrence over stage alone.	Fleming <i>et al.</i> [48]
miR-15B	N/A	283	Expression combined with stage separated patients by RFS and Os and improved prediction of recurrence over stage alone; serum levels significantly increased over time in recurrent patients.	Fleming <i>et al.</i> [48]
miR-425	N/A	283	Expression combined with stage separated patients by RFS and Os and improved prediction of recurrence over stage alone.	Fleming <i>et al.</i> [48]
miRNA signature: miR150, miR-15b; miR-199a-5p, miR33a and miR-424	N/A	130	Melanoma patient's classification into high and low recurrence risk groups with significant separation of RFS.	Friedman <i>et al.</i> [49]
miR-106b	N/A	129	High miR-106b expression was correlated with Breslow thickness, tumor ulceration and advanced clinical stage; patients with shorter 5-years OS showed high miR-106b expression than those with low expressions.	Lin <i>et al.</i> [50]
microRNA profile: miR-9, miR-145, miR-150, miR-155 and miR-205	N/A	41	The combination of miR-9, miR-145, miR-150, miR-155 and miR-205 was capable to distinguish the patients with metastasis from those without it.	Shiyyama <i>et al.</i> [51]

OS = overall survival, DFS = disease-free survival, DMFS = distant metastases-free survival, SLN = sentinel lymph node, MSS = melanoma-specific survival, RFS = recurrence-free survival, N/A = not applicable.

3.2 Artigo científico II: “Cell-free DNA as an inflammatory marker in post-surgical monitoring of melanoma patients”

Autores: Munique Mendonça Azevedo, Claudia Giuliano Bica, Ivana Beatrice Mânica da Cruz, Luciana El Halal Schuch, Fernanda Barbisan, Bruna Rudolfo Faraco, Rebeca Kollar Vieira da Silva, Yuri Thomé Machado Strey, Marta Medeiros Frescura Duarte, Thiago Duarte, Felice Riccardi

Enviado para publicação na Revista “Biomarkers”

Cell-free DNA as an inflammatory marker in post-surgical monitoring of melanoma patients

Munique Mendonça Azevedo, M.S.¹, Claudia Giuliano Bica, Ph.D^{1*}, Ivana Beatrice Mânica da Cruz, Ph.D², Luciana El Halal Schuch, M.D.¹, Fernanda Barbisan, M.S.², Bruna Rudolfo Faraco¹, Rebeca Kollar Vieira da Silva¹, Yuri Thomé Machado Strey¹, Marta Medeiros Frescura Duarte², Thiago Duarte², Felice Riccardi, M.S., M.D.³

¹ *Universidade Federal de Ciências da Saúde de Porto Alegre (UFCSPA), Porto Alegre, Brazil*

² *Universidade Federal de Santa Maria (UFSM), Santa Maria, Brazil*

³ *Hospital Santa Rita, Santa Casa de Misericórdia de Porto Alegre, Porto Alegre, Brazil*

* Corresponding author: CG Bica, 245 Sarmento Leite Street, Porto Alegre/RS, Brazil. Zip Code: 90050170. Phone: +5551 3303-8794, e-mail: claudia@ufcspa.edu.br

Cell-free DNA as an inflammatory marker in post-surgical monitoring of melanoma patients

Introduction: Cell-free DNA (cfDNA) is extracellular DNA detectable in blood plasma and increased levels have been reported in cancer patients. **Material and Methods:** A case-control study was conducted to determine potential association between cfDNA levels and melanoma. A total of 136 individuals (68 cases and 68 controls) had the plasmatic cfDNA concentration measured by an ultra-sensitive fluorescent dye and spectrofluorometer. In order to validate chronic inflammatory state of melanoma patients, the levels of proinflammatory cytokines (IL-1, IL-6, TNF- α , IFN- γ) and anti-inflammatory cytokine (IL-10) were also quantified. **Results:** Plasma cfDNA and proinflammatory cytokines levels were significantly higher in pre-surgical patients group compared with controls. In contrast, levels of IL-10 are higher in controls than in cases. A cutoff point was established and the individuals were categorized in high levels of cfDNA (≥ 46 ng/mL) and low levels of cfDNA (< 46 ng/mL) groups. Among melanoma patients, 23.5% of the subjects presented cfDNA high levels, while in healthy control, only 2.8% showed. Furthermore, high levels of cfDNA trended to be significantly associated with distant metastasis. There was a significant reduction of about 76% of cfDNA levels after tumor excision. **Conclusion:** cfDNA can be a potential biomarker for melanoma inflammatory status and a tool for help in post-surgical monitoring.

Keywords: Cell-free DNA – Biomarker – Inflammatory cytokines – Post-surgical follow-up – Prognostic factors

Cell-free DNA as an inflammatory marker in post-surgical monitoring of melanoma patients

Introduction

Cutaneous melanoma, arising from the transformation of melanocytes, represents the most aggressive skin cancer with rapid progression, intrinsic resistance to treatment and high mortality rate (1). The incidence of melanoma has increased over the past decades, faster than all other cancers (2), potentially due to improved surveillance, but also to increased sun exposure (3). When diagnosed early, melanoma can be cured by surgical excision. However, approximately 1 in 35 patients develop metastatic tumors and only 15% of those survive for 5 years (4, 5).

Concerning prognosis, the main clinical and histopathologic predictors of outcome are Breslow thickness, presence of ulceration, mitotic rate, as well as sentinel node status (6). However, melanoma has long been recognized as a highly heterogeneous disease which, in some cases, may show an unpredictable course (7, 8). For example, there is a subset of early-stage tumors exhibiting high-risk behavior that are not identified through histological prognostic factors in primary tumor (9). Despite many advances in melanoma molecular knowledge, the identification of patients at risk of developing advanced disease remains a valid medical issue that warrants the search for biomarkers with prognostic significance.

Cell-free DNA (cfDNA) is represented by fragments of double-stranded DNA that left the nucleus of the cells and is present in plasma, serum and others body fluids (10). Although mechanisms of cfDNA release into the blood stream are not completely elucidated, the most common hypothesis is that it arises from dead or dying immune cells (11). Evidences suggest that cfDNA

levels are elevated in both cancer patients (12-15) and in various non-malignant pathological conditions, such as chronic inflammatory diseases or tissue trauma (16, 17), compared to healthy individuals. Some studies even suggest the clinical relevance of this biomarker for the diagnosis, prognosis and monitoring of cancer (18).

Hypothetically, large amounts of cfDNA are expected in cancer patients as a result of increased apoptosis and necrosis process, characteristic of tumor cells turnover, leading to an overload of phagocytic cells responsible for clearing such debris (19, 20). Therefore, inflammatory processes could be triggered by tumoral areas, which exhibit chronic necrosis and apoptosis events. In consequence, waves of cell death are observed in cancer patients releasing cfDNA fragments to blood stream (21-23).

In this context, which cfDNA could be considered a marker related to inflammatory conditions and melanoma is a highly immunogenic cancer associated with massive immune cell infiltration, we performed a case-control study to determine potential association between cfDNA levels and melanoma. We also quantified the levels of proinflammatory cytokines, interleukin 1 (IL-1), IL-6, tumor necrosis factor alpha (TNF- α), gamma interferon (IFN- γ) and levels of IL-10, an anti-inflammatory cytokine, in order to validate chronic inflammatory state of melanoma patients. Additionally, we evaluated the impact of surgical treatment on cfDNA levels and the association with disease progression.

Clinical significance

- Cell-free DNA levels are higher in melanoma patients than healthy individuals;

- Proinflammatory cytokines levels are increased in melanoma cases compared to controls;
- Anti-inflammatory cytokine level are reduced in melanoma cases compared to controls;
- Cell-free DNA levels dropped after surgery in melanoma patients;
- Surgical process decreased inflammatory background of melanoma patients and cfDNA can be a potential tool for help in post-surgical monitoring.

Methods

Study population

The study was approved by two Ethical Board Committees (Universidade Federal de Ciências da Saúde de Porto Alegre and Santa Casa de Misericórdia de Porto Alegre, Porto Alegre, Brazil) where the study was conducted (approval number 465145 and 436339, respectively). Written informed consent was obtained from all subjects included in the study.

A case-control epidemiological study was conducted in order to evaluate the association between cfDNA levels and melanoma. One hundred thirty-six subjects were included, 68 cases and 68 healthy controls, sex and age matched. The cases were selected from patients who had received, for the first time melanoma diagnosis, and who had applied for treatment at the Melanoma Nucleus of Hospital Santa Rita (Santa Casa de Misericórdia de Porto Alegre) between the years 2013 and 2015. Healthy subjects were selected from academic community.

In order to exclude co-morbidities that could affect cfDNA levels, such as chronic inflammatory, autoimmune diseases and recent trauma, the medical records of all subjects were checked and also blood tests and imaging exams were analyzed. The patients compared with healthy subjects were identified as pre-surgical group.

In addition, a complementary analysis was conducted to evaluate the cfDNA levels of patients that reached a year of follow-up after the surgical treatment, designated as the post-surgical group. From 68 subjects enrolled in the study, 61 returned and participated of this second analysis.

The histological sections of all patients were reviewed by the institution's senior pathologist. Histopathological data from primary melanoma tumor were obtained from pathology report of biopsy and patients' medical records. Information from exposition to main risk factors for melanoma (sunburn episodes, familiar and personal melanoma history, phenotypic characteristics and skin's phototype) was collected through standardized questionnaire.

Blood sample collection

A total of 5mL of peripheral blood was collected from the study population in a sterile tube containing EDTA. The blood collection was carried out one single time in individuals of the control group. Among melanoma patients, two blood collections were performed: the first before the surgical procedure (pre-surgical group), meaning the primary tumor had not been completely excised yet at this point; and a second one year after surgical treatment (post-surgical group). In order to separate the blood plasma, the samples were centrifuged at 3.000 rpm

for 10 minutes and stored at -80°C until the quantification of cfDNA levels be run.

Quantitative analysis of cfDNA

cfDNA analysis was performed similar to described in Cichota *et al.*(24). Briefly, the commercial kit Quant-iT™ PicoGreen® dsDNA (Invitrogen) was used to quantify cfDNA. The Quant-iT™ PicoGreen® dsDNA is an ultra-sensitive fluorescent stain which has affinity for double-stranded DNA solutions, detecting and quantifying even low concentrations (from 25 pg/ml of DNA). The stain dilution was made according to the instructions of fabricant and the reactions were performed in 96 well plates. On each plate were added 10µL of plasma, 10 µL of fluorescent stain and 80 µL 1X TE. Calf thymus DNA was used for internal control of reaction. After 5 minutes, cfDNA concentration was reading on spectrofluorometer (SpectraMax® M2e Multimode Microplate Reader) with 480nm excitation and 520nm emission. Each sample was quantified on triplicate and the mean values were calculated to obtain cfDNA final concentration.

Inflammatory cytokines quantification

To validate the inflammatory state of melanoma patients, cytokine levels (IL-1, IL-6, TNFα, INFγ and IL-10) were quantified using the ELISA capture, according to the manufacturer's instructions (Abcam's kit, Cambridge, MA, USA).

Statistical Analysis

The data was analyzed using SPSS 23.0 software (Chicago, USA). To compare case and control groups, initially a Kolmogorov-Smirnov analysis was

performed indicating non-normal cfDNA distribution. For this reason we compared quantitative values between case and control groups by Mann-Whitney test and we presented results as mean $\pm$ standard deviation (SD), median and 95% confident interval (CI). From results obtained we performed a percentile distribution analysis and categorized our sample by 75th percentile value. Categorical analysis were performed by Chi-square or Fisher's exact test and results presented by relative frequencies (%). From univariate analysis the potential influence of sex and age was evaluated by multivariate analysis of logistic regression (Backward Wald method). A complementary analysis compared the cfDNA levels between pre and post-surgical moment by paired-Student *t* analysis. The statistical significance was established as $P < 0.05$.

Results

Cases and controls included in the study were caucasian, aged with 54.46 ± 14.84 and 55.62 ± 16.36 years old, respectively. In both cases and controls individuals, 51.5% (35/68) were men and 48.5% (33/68) were women. The baseline clinical characteristics referring to the primary tumor from melanoma patients are described in Table 1.

Table 1 here.

Table 2 shows the median concentration of cfDNA and proinflammatory cytokines in samples of melanoma patients without previous treatment and healthy controls. Plasma cfDNA levels were significantly higher in pre-surgical patients group (42.61 ± 32.96 ng/mL) compared with controls (17.36 ± 10.64 ng/mL) ($P < 0.005$). In the same way, levels of proinflammatory cytokines are significantly increased in melanoma patients when compared to healthy

individuals, except IL-10, which higher measures were found in controls than in cases ($P < 0.001$).

Table 2 here.

As cfDNA and cytokines distribution did not present a normal pattern, additional percentile analysis was performed. A cutoff point was determined from the percentile distribution analysis comparing the cfDNA levels of cases and controls. The percentiles analysis showed differences between groups in 75th cfDNA value, equivalent to 46 ng/ml. From these data, the samples were categorized in high levels of cfDNA group (≥ 46 ng/mL) and low levels of cfDNA group (< 46 ng/mL) and were statistically compared.

Among melanoma patients, 23.5% of the subjects (16/68) presented cfDNA levels above the cutoff point, while in healthy control, only 2.8% (2/68) ($\chi^2_1 = 13.219$; $P < 0.001$) showed. Additionally, the multivariate logistic regression analysis (Backward Wald method) showed that the association between melanoma and high levels of cfDNA (≥ 46 ng/mL) was independent of sample gender and age.

High levels of cfDNA trended to be significantly associated with distant metastasis ($\chi^2_1 = 3.878$; $P = 0.089$). Metastasis to distant organs were more frequent in group of melanoma patients with cfDNA levels ≥ 46 ng/mL than group with lower levels of cfDNA, 18.8% and 3.9% respectively (table 3).

Table 3 here.

Concerning to the others prognostic (Breslow thickness, mitotic rate, presence of ulceration and SLN status) and risk factors analyzed (previous skin cancer, another prior cancer, familial melanoma, skin phototype, eye color,

sunburn episodes and childhood freckles), no significant relationship with high and low cfDNA levels were identified.

Finally, when comparing average cfDNA levels before ($42.61 \text{ ng/mL} \pm 32.96$) and after primary tumor complete excision ($27.13 \text{ ng/mL} \pm 11.68$), there was a significant reduction of these levels after surgical treatment ($t_3 = 24.2$; $P < 0.05$). Furthermore, a significant difference was found between cfDNA concentration melanoma patients samples collected one year after surgery and control group (Figure 1).

Figure 1 here.

The impact of surgery on cfDNA was evaluated by percentage of cfDNA found in post-surgical group in comparison with pre-surgical group (100%). The mean of cfDNA dropped 76.9% indicating that surgical process decreased inflammatory background of melanoma patients.

Discussion

The results of the present investigation indicate that blood cfDNA levels quantified by Picogreen dye are increased in melanoma patients prior surgery. Moreover, was observed a lowering in cfDNA values when patients were submitted to tumor excision. For our best knowledge, this is the first report with association between cfDNA levels and melanoma patients pre and post-surgery.

The presence of cfDNA in human plasma was described, for the first time, over six decades ago (25). Nonetheless, only in 1977 the presence of abnormally high levels of cfDNA was identified in the blood stream of cancer patients compared with healthy controls (26). Since then, the potential

diagnostic, prognostic and monitoring significance of cfDNA has been demonstrated for various types of cancer (27-29).

The progression in the isolation of cfDNA was only possible due to the development of sensitive molecular procedures using specific fluorescent dyes and PCR techniques (30). Nowadays, cfDNA quantification is easy, low cost and minimally invasive technique sensible enough to detect few quantities of DNA released in the blood.

Corroborating our results, plenty of studies have been demonstrating increased levels of cfDNA in cancer patients compared with those with non-malignant disease (31-33). In addition, we showed that there is association between increased levels of cfDNA and melanoma, regardless of age or sex. It is believed that DNA release in the blood of patients with malignant disease is associated with neoplastic cellular death process in tumoral environment (34). Associated with necrosis and apoptosis, inflammation process can also coexist in tumors, leading to an elaboration of cytokines and inflammatory mediators (21-23).

Prior evidences reported that melanoma is strongly associated with the inflammatory process, which can be observed by high levels of proinflammatory cytokine secretion (35). For this reason, we quantified proinflammatory cytokines IL-1, IL-6, TNF- α , IFN- γ and IL-10 in the pre-surgical group to confirm the inflammatory background of melanoma patients. Actually, our results showed higher levels of proinflammatory cytokines, and lower levels of IL-10, an anti-inflammatory cytokine, in case group than control. Corroborating our findings, Wiguna and Walden (36) showed that the expression of IL-10 gene is lower in melanoma metastases than normal skin.

It is well established that inflammatory process is mediated by a balance between proinflammatory (Th1) and anti-inflammatory (Th2) cytokines expressed by CD4⁺ T cells (37). In this way, significant association or with proinflammatory cytokines or anti-inflammatory cytokines may indicate that cfDNA levels could be an emergent marker of melanoma inflammatory status. In fact, subjects with higher cfDNA levels (> 46 ng/mL) showed a significant association with lower IL-10 levels.

Potential association between cfDNA levels and other clinical aspects of melanoma diseases was also evaluated, but no statistically significant was found. However, our data showed a tendency of association between higher cfDNA levels and distant metastasis development in patients' follow-up during one year. Unfortunately, statistical analysis was not significant probably due to a low number of patients including in our study and/or the short follow-up time that the patients were observed. In the literature, it is recognized that distance metastasis also has the potential to collaborate with increasing levels of cfDNA in individuals (38).

We are conscious that number of patients was low affecting the analysis of results. However, melanoma patients with a recent diagnosis, without prior treatment or with other non-inflammatory diseases and dysfunctions are very hard to be selected. For this reason, we believe that our results could be considered epidemiologically relevant.

Finally, after surgery for tumor excision the cfDNA plasmatic levels of melanoma patients decreased significantly, although it has not reached a concentration as low as those present in healthy individuals. Therefore, it is probable that cfDNA levels decrease after the surgical treatment is

consequence of local inflammation reduction. The decrease of cfDNA levels after anticancer therapies has already been reported in the literature (34) and it has been observed that the maintenance of high levels of this biomarker may suggest the presence of residual tumor cells (39).

All this results combined propound that the quantification of cfDNA has the potential to be consider an emergent marker of melanoma inflammatory status, as well as its potential to become an additional monitoring method for surgical success of melanoma patients.

Conclusion

The cfDNA proved to be a significant biomarker for melanoma inflammatory status regardless of sex and age, as well as its potential as a complementary test in the postoperative monitoring of the patients. In this manner, changes in cfDNA can be traced by repeated blood samples during patient's follow-up allowing for early relapse detection. The major challenge is melanoma management is the lack of an efficient marker for detecting disease progression at an early stage and/or for monitoring patients' treatment response. The employing of non-invasive blood biomarker assays during follow-up allow for monitoring of disease dynamic associated with treatment effectiveness is crucial in order to improve patients' outcome.

Acknowledgments

The authors are grateful for the help of the Laboratory of Biogenomics (Universidade Federal de Santa Maria, Santa Maria, Brazil) and of the Laboratory of Pathology (Universidade Federal de Ciências da Saúde de Porto Alegre, Porto Alegre, Brazil).

Declaration of interest

The authors declare that they have no conflict of interest including any financial, personal or other relationships with other people or organizations that could inappropriately influence, or be perceived to influence, the present work.

Funding information

This study was funded by the research grants from Conselho Nacional de Desenvolvimento Científico e Tecnológico - CNPq (Grant number 441727/2014) and fellowships from Comissão de Aperfeiçoamento de Pessoal de Nível Superior - CAPES.

References

1. Tsao H, Chin L, Garraway LA, Fisher DE. Melanoma: from mutations to medicine. *Genes & development*. 2012;26(11):1131-55.
2. Lens MB, Dawes M. Global perspectives of contemporary epidemiological trends of cutaneous malignant melanoma. Oxford, UK2004. p. 179-85.
3. Dye DE, Medic S, Ziman M, Coombe DR. Melanoma biomolecules: independently identified but functionally intertwined. *Front Oncol*. 2013;3:252.
4. Siegel R, Naishadham D, Jemal A. Cancer statistics, 2013. *CA: a cancer journal for clinicians*. 2013;63(1):11-30.
5. Finn L, Markovic SN, Joseph RW. Therapy for metastatic melanoma: the past, present, and future. *BMC medicine*. 2012;10(1):1.

6. Balch CM, Gershenwald JE, Soong SJ, Thompson JF, Atkins MB, Byrd DR, et al. Final version of 2009 AJCC melanoma staging and classification. *J Clin Oncol*. 2009;27(36):6199-206.
7. Meguerditchian A-N, Asubonteng K, Young C, Lema B, Wilding G, Kane JM. Thick primary melanoma has a heterogeneous tumor biology: an institutional series. *World journal of surgical oncology*. 2011;9(1):1.
8. Gimotty PA, Elder DE, Fraker DL, Botbyl J, Sellers K, Elenitsas R, et al. Identification of high-risk patients among those diagnosed with thin cutaneous melanomas. *Journal of Clinical Oncology*. 2007;25(9):1129-34.
9. Cheng Y, Lu J, Chen G, Ardekani GS, Rotte A, Martinka M, et al. Stage-specific prognostic biomarkers in melanoma. *Oncotarget*. 2015;6(6):4180-9.
10. Su YH, Wang M, Brenner DE, Norton PA, Block TM. Detection of Mutated K-ras DNA in Urine, Plasma, and Serum of Patients with Colorectal Carcinoma or Adenomatous Polyps. *Annals of the New York Academy of Sciences*. 2008;1137(1):197-206.
11. Mittra I, Nair NK, Mishra PK. Nucleic acids in circulation: Are they harmful to the host? *Journal of biosciences*. 2012;37(2):301-12.
12. Tangvarasittichai O, Jaiwang W, Tangvarasittichai S. The plasma DNA concentration as a potential breast cancer screening marker. *Indian J Clin Biochem*. 2015;30(1):55-8.
13. Ulivi P, Silvestrini R. Role of quantitative and qualitative characteristics of free circulating DNA in the management of patients with non-small cell lung cancer. *Cell Oncol (Dordr)*. 2013;36(6):439-48.

14. Kamat AA, Baldwin M, Urbauer D, Dang D, Han LY, Godwin A, et al. Plasma cell- free DNA in ovarian cancer: an independent prognostic biomarker.(Clinical report). *Cancer*. 2010;116(8):1918.
15. Jung K, Stephan C, Lewandowski M, Klotzek S, Jung M, Kristiansen G, et al. Increased cell- free DNA in plasma of patients with metastatic spread in prostate cancer. *Cancer Letters*. 2004;205(2):173-80.
16. Galeazzi M, Morozzi G, Piccini M, Chen J, Bellisai F, Fineschi S, et al. Dosage and characterization of circulating DNA: present usage and possible applications in systemic autoimmune disorders. *Autoimmunity Reviews*. 2003;2(1):50-5.
17. Laktionov PP, Tamkovich SN, Rykova EY, Bryzgunova OE, Starikov AV, Kuznetsova NP, et al. Extracellular circulating nucleic acids in human plasma in health and disease. *Nucleosides, nucleotides & nucleic acids*. 2004;23(6-7):879.
18. Larson AR, Konat E, Alani RM. Melanoma biomarkers: current status and vision for the future. *Nat Clin Pract Oncol*. 2009;6(2):105-17.
19. Alonso SR, Ortiz P, Pollan M, Perez-Gomez B, Sanchez L, Acuna MJ, et al. Progression in cutaneous malignant melanoma is associated with distinct expression profiles: a tissue microarray-based study. *Am J Pathol*. 2004;164(1):193-203.
20. Gormally E, Caboux E, Vineis P, Hainaut P. Circulating free DNA in plasma or serum as biomarker of carcinogenesis: Practical aspects and

biological significance. *Mutation Research-Reviews in Mutation Research*. 2007;635(2):105-17.

21. Duffield JS. The inflammatory macrophage: a story of Jekyll and Hyde. *Clinical science*. 2003;104(1):27-38.

22. Gordon S. Alternative activation of macrophages. *Nature reviews immunology*. 2003;3(1):23-35.

23. Fadok VA, Bratton DL, Guthrie L, Henson PM. Differential effects of apoptotic versus lysed cells on macrophage production of cytokines: role of proteases. *J Immunol*. 2001;166(11):6847-54.

24. Cichota LC, Bochi GV, Tatsch E, Torbitz VD, Agnol P, Zanardo JC, et al. Circulating Double-Stranded DNA in Plasma of Hemodialysis Patients and its Association with Iron Stores. *Clinical laboratory*. 2014;61(8):985-90.

25. Mandel P, Metais P. Les acides nucléiques du plasma sanguin chez l'homme. *C R Seances Soc Biol Fil*. 1948;142(3-4):241-3.

26. Leon SA, Shapiro B, Sklaroff DM, Yaros MJ. Free DNA in the serum of cancer patients and the effect of therapy. *Cancer research*. 1977;37(3):646.

27. Papadopoulou E, Davilas E, Sotiriou V, Georgakopoulos E, Georgakopoulou S, Koliopanos A, et al. Cell- free DNA and RNA in Plasma as a New Molecular Marker for Prostate and Breast Cancer. *Annals of the New York Academy of Sciences*. 2006;1075(1):235-43.

28. Frattini M, Gallino G, Signoroni S, Balestra D, Battaglia L, Sozzi G, et al. Quantitative analysis of plasma DNA in colorectal cancer patients: a novel prognostic tool. *Annals of the New York Academy of Sciences*. 2006;1075:185.
29. Kamat AA, Sood AK, Dang D, Gershenson DM, Simpson JL, Bischoff FZ. Quantification of total plasma cell-free DNA in ovarian cancer using real-time PCR. *Annals of the New York Academy of Sciences*. 2006;1075:230.
30. Jung K, Fleischhacker M, Rabien A. Cell-free DNA in the blood as a solid tumor biomarker—A critical appraisal of the literature. *Clinica Chimica Acta*. 2010;411(21):1611-24.
31. Ren N, Ye Q-H, Qin L-X, Zhang B-H, Liu Y-K, Tang Z-Y. Circulating DNA level is negatively associated with the long-term survival of hepatocellular carcinoma patients. *World Journal of Gastroenterology*. 2006;12(24):3911-4.
32. Spindler KL, Pallisgaard N, Andersen RF, Brandslund I, Jakobsen A. Circulating free DNA as biomarker and source for mutation detection in metastatic colorectal cancer. *PLoS One*. 2015;10(4):e0108247.
33. Yoon KA, Park S, Lee SH, Kim JH, Lee JS. Comparison of circulating plasma DNA levels between lung cancer patients and healthy controls. *J Mol Diagn*. 2009;11(3):182-5.
34. Schwarzenbach H, Hoon DS, Pantel K. Cell-free nucleic acids as biomarkers in cancer patients. *Nature Reviews Cancer*. 2011;11(6):426-37.
35. Melnikova V, Bar-Eli M. Inflammation and melanoma growth and metastasis: the role of platelet-activating factor (PAF) and its receptor. *Cancer and Metastasis Reviews*. 2007;26(3-4):359-71.

36. Wiguna AP, Walden P. Role of IL-10 and TGF- β in melanoma. *Experimental dermatology*. 2015;24(3):209-14.
37. Smyth MJ, Cretney E, Kershaw MH, Hayakawa Y. Cytokines in cancer immunity and immunotherapy. *Immunological reviews*. 2004;202(1):275-93.
38. Schwarzenbach H, Alix-Panabières C, Müller I, Letang N, Vendrell J-P, Rebillard X, et al. Cell- free tumor DNA in blood plasma as a marker for circulating tumor cells in prostate cancer. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2009;15(3):1032.
39. Wimberger P, Roth C, Pantel K, Kasimir-Bauer S, Kimmig R, Schwarzenbach H. Impact of platinum- based chemotherapy on circulating nucleic acid levels, protease activities in blood and disseminated tumor cells in bone marrow of ovarian cancer patients. *International journal of cancer Journal international du cancer*. 2011;128(11):2572.

Figure captions

Figure 1: Comparison of cfDNA levels between cases at pre-surgical moment and after one year from surgery and healthy controls. The surgical treatment decreased cfDNA levels in melanoma patients, leading to concentration almost as low as those observed in controls.

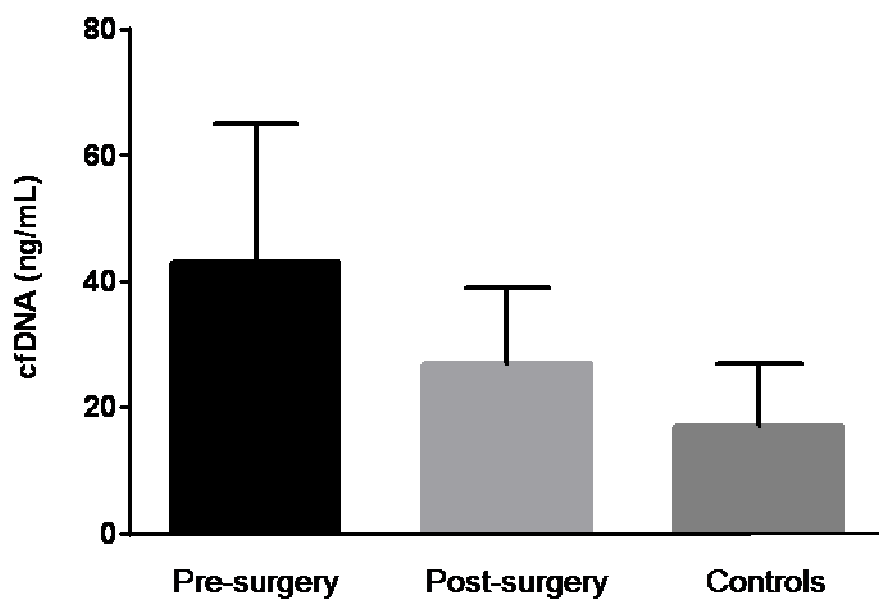


Table 1. Clinicopathological features from primary tumor of melanoma patients.

Variable	<i>N</i>	%
Tumor Site		
Head/neck	11	16.2
Trunk	36	52.9
Upper limb	12	17.7
Lower limb	9	13.2
Histological subtype		
Superficial spreading	51	75
Lentigo maligna	7	10.3
Nodular	7	10.3
Acral lentiginous	2	2.9
Desmoplastic	1	1.5
Clark's level		
I	16	23.6
II	13	19.1
III	17	25
IV	20	29.4
V	2	2.9
Breslow thickness		
<i>Tis</i>	16	23.5
≤ 1 mm	32	47.1
1.01 - 2 mm	12	17.6
2.01 - 4 mm	3	4.4
> 4 mm	5	7.4
Presence of ulceration		
Yes	8	11.8
No	60	88.2
Mitotic rate		
Mean ± SD	1.63 ± 2.51	
Satellitosis		
Yes	2	2.9
No	66	97.1

Tis = tumor *in situ*; SD = standard deviation

Table 2. Comparison of cfDNA and inflammatory cytokines levels from melanoma patients before surgical treatment (cases) and healthy controls.

Inflammatory markers	Cases (<i>N</i> = 68)			Controls (<i>N</i> = 68)			<i>P</i> value*
	Mean ± SD**	Median	95% CI (Lower-Upper)	Mean ± SD**	Median	95% CI (Lower-Upper)	
cfDNA (ng/μL)	42.61 ± 32.96	34.94	34.44 – 52.16	17.36 ± 10.64	14	14 – 19.3	< 0.005
IL-1 (pg/mL)	175.92 ± 51,27	166	161.34 – 190.49	101.45 ± 16.69	100	97.3 – 105.62	< 0.001
IL-6 (pg/mL)	185 ± 72,69	180	185 – 226.31	126.64 ± 14.81	129	122 – 130.84	< 0.001
TNFα (pg/mL)	221.38 ± 71.6	212	201.03 – 241.72	144.5 ± 26.6	149	137.44 – 152.31	< 0.001
INFγ (pg/mL)	277.24 ± 101.5	294	248.39 – 306.08	200.26 ± 37.16	213	189.64 – 210	< 0.001
IL-10 (pg/mL)	32.3 ± 12.12	33.5	28.85 – 35.74	67.2 ± 22.65	59.5	59.5 – 73.68	< 0.001

* Mann-Whitney test was used in comparisons.

**SD = standard error.

Table 3. Association of melanoma risk and prognostic factors with cfDNA levels.

Variable	N	cfDNA level (%)		P value
		≥ 46 ng/mL	< 46 ng/mL	
Previous skin carcinoma				0.58
Yes	10	0	19.2	
No	58	100	80.8	
Another prior cancer				0.064
Yes	3	0	3	
No	65	35	30	
Familial melanoma				0.835
Yes	14	18.8	21.2	
No	54	81.3	78.8	
Skin phototype I/II				0.722
Yes	45	62.5	67.3	
No	23	37.5	32.7	
Eye color				0.788
Light	36	50	53.8	
Dark	32	50	47.1	
Sunburn episodes				0.184
Childhood	10	25	11.5	
Another periods of life	58	75	88.5	
Childhood freckles				0.897
Yes	13	7.7	92.3	
No	55	27.3	72.7	
Presence of ulceration				0.346
Yes	9	6.3	15.4	
No	59	93.8	84.6	
SLN status				0.933
Positive	4	14.3	13	
Negative	26	85.7	87	
Metastasis				0.089
Yes	5	18.8	3.9	
No	63	81.3	96.1	
Breslow thickness				0.634*
Mean ± SD	68	1.04 ± 1.31	1.27 ± 2.46	
Mitotic rate				0.934*
Mean ± SD	68	1.66 ± 2.55	1.60 ± 2.51	

SLN = sentinel lymph node; SD = standard deviation; *Statistical test: Student's *t* test (the others variables were analyzed with Chi-square test); Statistical significance considered when $P < 0.05$.

3.3 Artigo científico II: “Gender and age-specific association between manganese superoxide dismutase polymorphism (Ala16Val-SOD2) and melanoma risk”

Autores: Munique Azevedo, Ivana Cruz, Luciana Schuch, Fernanda Barbisan, Bruna Faraco, Rebeca da Silva, Amanda Weigert, Yuri Strey, Marta Duarte, Thiago Duarte, Felice Riccardi e Claudia Bica

Enviado para publicação na Revista “Pathology & Oncology Research”

Gender and age-specific association between manganese superoxide dismutase polymorphism (Ala16Val-SOD2) and melanoma risk

Short title: Ala16Val-SOD2 polymorphism and melanoma risk

Munique Mendonça Azevedo^{1, †}, Ivana Beatrice Mânica Cruz^{2, †}, Luciana El Halal Schuch^{1, †}, Fernanda Barbisan^{2, †}, Bruna Rudolfo Faraco^{2, †}, Rebeca Kollar Vieira da Silva^{1, †}, Amanda Carolina Aguiar Weigert^{1, †}, Yuri Thomé Machado Strey^{1, †}, Marta Medeiros Frescura Duarte^{2, †}, Thiago Duarte^{2, †}, Felice Riccardi^{3, †}, Claudia Giuliano Bica^{2, *}

¹ Universidade Federal de Ciências da Saúde de Porto Alegre, Porto Alegre, 90050170, Brazil

² Universidade Federal de Santa Maria, Santa Maria, 97105900, Brazil

³ Hospital Santa Rita, Santa Casa de Misericórdia de Misericórdia de Porto Alegre, Porto Alegre, 90020090, Brazil

* Claudia Bica. Tel: +55 51 33038794; E-mail: claudia@ufcspa.edu.br

Abstract

Reactive oxygen species (ROS) inadequately detoxified may result in oxidative stress, which is associated with cancer development and progression, including skin carcinogenesis. Enzymatic detoxification is one of the defense mechanisms to protect cells against ROS. We evaluated the association

between melanoma and the functional polymorphism of SOD2 (Ala16Val-SOD2), which encodes the manganese superoxide dismutase (MnSOD). The polymorphism was determined in 62 melanoma patients and 500 healthy subjects. Genotyping from blood of cases and controls was carried out through PCR-RFLP method using specific primers and *Hae III* restriction enzyme. The genetic frequencies of Ala16Val-SOD2 polymorphism were in Hardy-Weinberg equilibrium. Allele frequencies were A = 0.55 and V = 0.45 in the control group and A = 0.56 and V = 0.44 in melanoma group. Comparison between genotypes showed a higher frequency of AA in melanoma patients when compared to healthy subjects. For the first time in literature, our results provide evidences about an increased melanoma risk associated with SOD2 polymorphism. However this association is not independent of sex and age, meaning that women and elderly carrying AA genotype are more susceptibility to melanoma.

Keywords: Oxidative stress, antioxidant enzymes, Ala16Val polymorphism, melanoma risk

Acknowledgements

The authors are grateful for the help of the Laboratory of Biogenomics (Universidade Federal de Santa Maria, Santa Maria, Brazil) and of the Laboratory of Pathology (Universidade Federal de Ciências da Saúde de Porto Alegre, Porto Alegre, Brazil).

Gender and age-specific association between manganese superoxide dismutase polymorphism (Ala16Val-SOD2) and melanoma risk

Introduction

Cutaneous melanoma, arising from abnormal proliferation of melanocytes, is an aggressive and lethal form of skin cancer due to its propensity to metastasize and intrinsic resistance to treatment [1]. Although melanoma represents only 4% of all dermatological neoplasias, it is responsible by 75% of skin cancer-related deaths [2]. The disease can be cured by surgical removal when diagnosed in early stage. However, only 14% of metastatic melanoma patients survive for 5 years [2]. Currently, the American Joint Committee on Cancer (AJCC) staging system, based on histopathological features of primary tumor, is widely used to predict melanoma prognosis [3]. Patients are classified into four stages according Breslow thickness, presence of ulceration, extent of nodal involvement for primary and locally metastatic melanoma, metastasis to distant organs and serum lactate dehydrogenase (LDH) level [3].

In the last decades, the incidence of melanoma is increasing worldwide, faster than any other malignancy [4]. This increase can be explained both by changes in diagnostic criteria and early detection due to improved surveillance, as by increased sun exposure [5]. Consider the major etiological factor in skin cancer, the ultraviolet radiation [UVR] of the sun can directly damage DNA or generate reactive oxygen species (ROS) [6].

ROS, such as superoxide anion radical ($O_2^{\cdot-}$), hydrogen peroxide (H_2O_2), or hydroxyl radical ($\cdot OH$), are generated during normal aerobic metabolism or exposure to toxicants [7,8]. ROS inadequately detoxified may result in oxidative stress which is associated with cancer development and progression, including skin carcinogenesis [9,10,11,12]. Concerning to the melanoma, ROS are present in all stages of its development and systemic and tumor redox deregulation has been reported in patients [13-16].

ROS are known to react with macromolecules inducing DNA oxidative damage which can result in cell death or mutations [17]. One of the defense mechanisms to protect cells from ROS includes detoxifying enzymes [15]. Manganese superoxide dismutase (MnSOD), a family of superoxide dismutases (SODs), is an endogenous antioxidant enzyme that catalyze the dismutation of superoxide into oxygen and hydrogen peroxide in the mitochondria [18]. This enzyme is encoded by the superoxide dismutase (SOD2) gene (locus 6q25.3). A polymorphism occurs at codon 16 of SOD2 gene where a single nucleotide change from T to C leading to valine-to-alanine substitution (Ala16Val)[19]. The Ala variant allows more efficient SOD2 uptake into the mitochondria and generates more active SOD2 compared with the Val variant [32, 33]. Despite extensively researched, Ala16Val-SOD2 polymorphism association with cancer remains controversial. Several studies have suggested increased cancer risk associated with the AA [Ala/Ala] genotype, contrary to might be predicted [22-26].

As far as we know, there are few studies evaluating SOD2 polymorphism in melanoma [27]. Therefore, in this study, we investigated the association between the Ala16Val-SOD2 polymorphism and melanoma. In addition, we

evaluated the potential association between this genetic variant and melanoma clinicopathological prognostic factors, as well as the relationship with phototype and sunburn episodes related with risk factors on melanoma.

Methods

Study population

A case-control study was conducted in order to evaluate the association between MnSOD polymorphism and melanoma. A total of 562 subjects were included, from which 62 were melanoma patients and 500 were healthy subjects.

Eligible for inclusion as cases in the present study were patients diagnosed with melanoma who applied for treatment at the Melanoma Nucleus of Hospital Santa Rita (Porto Alegre, Brazil) between the years 2013 and 2015. Medical records of patients were checked and blood tests and imaging exams were analyzed to exclude possible co-morbidities and other cancer diagnosis prior to the date of blood draw. Histological sections from biopsies, surgery and sentinel lymph nodes were analyzed by the institution's senior pathologist. Information about the primary tumor was recollected from pathology report of biopsy and patients' medical records. Information from exposition to main risk factors for melanoma was collected through standardized questionnaire.

Control group was composed by healthy volunteers from an academic community and were paired by sex and age with cases. Controls were eligible for selection if they had no pathologic condition such cancer, chronic

inflammation and recent trauma. Institutional review board approved the study. Written informed consent was obtained from all subjects included in the study.

DNA extraction and Ala16Val-SOD2 polymorphism genotyping

Peripheral blood was collected from all participants by venous puncture and was stored at -20°C until analysis. Genomic DNA from blood leukocytes was extracted using a salting-out method [28]. Cases and controls were genotyped as described by Bica *et al* [29]. Briefly, the method is based in a Polymerase Chain Reaction followed by Restriction Fragment Length Polymorphism (PCR-RFLP). Initially, a fragment of the human SOD2 gene (110bp) was amplified with primers 5'-ACCAGCAGGCAGCTGGCGCCGG-3' (sense-strand) and 5'-GCGTTGATGTGAGGTTCCAG-3' (antisense-strand). The PCR reaction was carried out in a total volume of 50 µL containing 5 µL 10X PCR buffer, 1.25 µL 10mM dNTPs, 1 µL 25mM MgCl₂, 0.5 µL Taq polymerase (Gibco Inc, Co.), 10 ng genomic DNA, 0.4 pM each primer and enough ddH₂O for complete the reaction volume. PCR reaction consisted of an initial denaturation at 95 °C for 5 min followed by 35 cycles at 95 °C for 1 min, 61 °C for 1 min and 30 s of extension at 72 °C. An additional final extension of 7 min at 72°C was performed at the end of final cycle. Amplicons (10 µL) were digested with specific endonuclease *Hae III* (15 U, Gibco. Inc, Co.) for 6 h at 37°C. Digestion products were analyzed by electrophoresis in 4% agarose gel stained with ethidium bromide. Different patterns of restriction are observed in agarose gel electrophoresis according to genotype: AA (2 fragments; 23 and 85 bp); AV (3 fragments; 23, 85 and 110 bp); and VV (110 bp).

Statistical analysis

The data was analyzed using SPSS 23.0 software (Chicago, USA). Student's t test was carried out to compare quantitative baseline variables between case and control subjects, while Chi-square (χ^2) was performed to compare categorical variables between the groups. The allele frequencies were estimated by the gene-counting method. Chi-square analysis was used to estimate the Hardy–Weinberg equilibrium. The allelic and genotype frequencies were compared among groups using chi-square or the Fisher's exact test. The association between prognostic/risk factors of melanoma and Ala16Val-SOD2 polymorphism was evaluated by ANOVA or chi-square test. The statistical significance was established as $P < 0.05$.

Results

Clinicopathological features of melanoma cases are listed in Table 1. Melanoma patients and healthy individual included in the study were Caucasian and there were no significant differences in age and sex in both groups. Table 2 shows baseline characteristics of the study population and Ala16Val-SOD2 genotype frequencies.

Table 1 here.

The genetic frequencies of Ala16Val-SOD2 polymorphism were in Hardy-Weinberg equilibrium. Allele frequencies were $A = 0.55$ and $V = 0.45$ in the

control group and $A = 0.56$ and $V = 0.44$ in melanoma group. Comparison between genotypes showed a higher frequency of AA in melanoma patients when compared to healthy subjects.

Table 2 here.

However, the association between AA frequency and melanoma was influenced by sex and age. Women with AA genotype were at higher melanoma risk ($P < 0.05$; OR = 2.49; 95% CI = 1.15 – 4.41). Similarly, elderly (≥ 60 years old) homozygote AA were more susceptible to melanoma development ($P < 0.001$; OR = 4.0; 95% CI = 1.79 – 8.97) (Figure 1 and 2).

In addition, the association between tumoral aggressiveness at the time of diagnosis and Ala16Val-SOD2 polymorphism was evaluated in melanoma patients. There was no significant association between genetic polymorphism and Breslow thickness ($P = 0.51$), mitosis rate ($P = 0.89$), presence of ulceration ($P = 0.53$) or metastasis ($P = 0.88$). Moreover, no significant interaction was observed between this polymorphism and patients' phototype ($P = 0.88$) or sunburn history ($P = 0.88$).

Figure 1 here.

Figure 2 here.

DISCUSSION

The results of the present case–control study indicate that the genotypes frequencies of Ala16Val-SOD2 polymorphism were different when melanoma

patients were compared to controls. A significant association was found between AA genotype and melanoma risk. Nevertheless, this association is not independent of sex and age, meaning that women and elderly carrying AA genotype are more susceptible to melanoma. To our knowledge, this is the first report in medical literature to suggest an association of melanoma and SOD2 polymorphism.

Previous results described by Han *et al* in skin cancer have shown no significant association between Ala16Val-SOD2 polymorphism and the risk of each type of skin cancer, including melanoma. Similar to our finding, they did not observe significant interaction between this polymorphism and sunburn history, well known as an etiological factor in dermatological neoplasias[27, 30].

Ultraviolet radiation can induce photooxidative damage in the skin and it is an important issue since sun exposure has been increasing [5, 31]. The oxidative stress induced by inflammatory response to UV exposure causes damage to cellular components and changes in pattern of gene expression leading to skin pathologies such as cancer [12].

Concerning antioxidant mechanism, manganese superoxide dismutase (MnSOD) is considered the first line of defense against reactive oxygen species produced by oxidative phosphorylation upon UV radiation and inflammation [32]. Evidences suggest that SOD activities are altered in blood of melanoma patients, indicating oxidative stress associated to the disease [15].

Several epidemiological studies have been evaluating the association between the functional polymorphism Ala16Val-SOD2 and cancer risk [33]. It has been postulated that this polymorphism, which alters the secondary structure of MnSOD, affects the efficiency of mitochondrial transport of the enzyme [19]. In

fact, Ala variant allows more efficient MnSOD uptake into the mitochondria than Val variant, suggesting that AA homozygous may have higher SOD2 antioxidant activity, which could suppress carcinogenesis process [20, 21, 33]. However, this hypothesis is controversial in the literature, with an increase in the number of studies that have observed association between Ala allele and cancer risk [23,34]. Bica *et al* in a case-control study found that the risk of breast cancer was increased in men and women who carried AA genotype, as well as in prostate cancer, when compared to controls[29].

Our results also suggest that AA genotype significantly increase the baseline risk of melanoma in women and elderly. Although confirmation of this study is needed due to the small number of melanoma patients included, these findings suggest an important role of Ala16Val-SOD2 polymorphism in melanoma susceptibility related to sex and age. Therefore, complementary studies with a larger sample size are needed to confirm these results.

Funding

This study was funded by the research grants from Conselho Nacional de Desenvolvimento Científico e Tecnológico - CNPq (Grant number 441727/2014) and fellowships from Comissão de Aperfeiçoamento de Pessoal de Nível Superior - CAPES.

Conflict of Interest

The authors declare that they have no conflict of interest.

References

1. Tsao H, Chin L, Garraway LA *et al.* (2012) Melanoma: from mutations to medicine. *Gen Dev*, **26**, 1131-1155.
2. Miller AJ, Mihm MC Jr. (2006) Melanoma. *N Engl J Med*, **355**, 51-65.
3. Balch CM, Gershenwald JE, Soong SJ *et al.* (2009) Final version of 2009 AJCC melanoma staging and classification. *J Clin Oncol*, **27**, 6199-206.
4. Garbe C, Peris K, Hauschild A *et al.* (2010) Diagnosis and treatment of melanoma: European consensus-based interdisciplinary guideline. *Eur J Cancer*, **46**, 270-283.
5. Dye DE, Medic S, Ziman M, Coombe DR (2013) Melanoma biomolecules: independently identified but functionally intertwined. *Front Oncol*, **3**, 252.
6. de Gruijl FR (2002) Photocarcinogenesis: UVA vs. UVB radiation. *Skin Pharmacol Appl Physiol*, **15**, 316-320.
7. Freeman BA, Crapo JD (1982) Biology of disease: free radicals and tissue injury. *Laboratory Invest*, **47**, 412-426.
8. Matés JM, Pérez-Gómez C, Núñez de Castro I (1999) Antioxidant enzymes and human diseases. *Clin Biochem*, **32**, 595-603.
9. Halliwell B (2007) Oxidative stress and cancer: have we moved forward? *Biochem J*, **401**, 1-11.
10. Klaunig, JE, Kamendulis LM (2004) The role of oxidative stress in carcinogenesis. *Annu Rev Pharmacol Toxicol*, **44**, 239-267.
11. Nishigori C, Hattori Y, Toyokuni S (2004) Role of reactive oxygen species in skin carcinogenesis. *Antioxid Redox Signal*, **6**, 561-570.

12. Sander CS, Chang H, Hamm F, *et al.* (2004) Role of oxidative stress and the antioxidant network in cutaneous carcinogenesis. *Int J Dermatol*, **43**, 326-335.
13. Meyskens FL, McNulty SE, Buckmeier JA, *et al.* (2001) Aberrant redox regulation in human metastatic melanoma cells compared to normal melanocytes. *Free Rad Biol Med*, **31**, 799-808.
14. Sander C, Hamm F, Elsner P, Thiele JJ (2003) Oxidative stress in malignant melanoma and non-melanoma skin cancer. *Br J Dermatol*, **148**, 913-922.
15. Gadjeva V, Dimov A, Georgieva N (2008) Influence of therapy on the antioxidant status in patients with melanoma. *J C PharmTher*, **33**, 179-185.
16. Picardo M, Grammatico P, Roccella F, Roccella M, Grandinetti M, Del Porto G, Passi S (1996) Imbalance in the antioxidant pool in melanoma cells and normal melanocytes from patients with melanoma. *J Invest Dermatol*, **107**, 322-326.
17. Wiener H, Perry RT, Chen Z, Harrell LE, Go RC (2007) A polymorphism in SOD2 is associated with development of Alzheimer's disease. *Genes Brain Behav*, **6**, 770-776.
18. Borgstahl GE, Parge HE, Hickey MJ, *et al.* (1996) Human mitochondrial manganese superoxide dismutase polymorphic variant Ile58Thr reduces activity by destabilizing the tetrameric interface. *Biochemistry*, **35**, 4287-4297.

19. Rosenblum JS, Gilula NB, Lerner RA (1996) On signal sequence polymorphisms and diseases of distribution. *Proc Natl Acad Sci U S A*, **93**, 4471-4473.
20. Sutton, A. Hania K, Prip-Buus C, *et al.* (2003) The Ala16Val genetic dimorphism modulates the import of human manganese superoxide dismutase into rat liver mitochondria. *Pharmacogenetics*, **13**, 145-157.
21. Sutton A, Imbert A, Igoudjil A, *et al.* (2005) The manganese superoxide dismutase Ala16Val dimorphism modulates both mitochondrial import and mRNA stability. *Pharmacogenetics*, **15**, 311-319.
22. Woodson K, Tangrea JA, Lehman TA, *et al.* (2003) Manganese superoxide dismutase (MnSOD) polymorphism, α -tocopherol supplementation and prostate cancer risk in the Alpha-Tocopherol, Beta-Carotene Cancer Prevention Study (Finland). *Cancer Causes Control*, **14**, 513-518.
23. Li H, Kantoff PW, Giovannucci E, *et al.* (2005) Manganese superoxide dismutase polymorphism, prediagnostic antioxidant status, and risk of clinical significant prostate cancer. *Cancer Res*, **65**, 2498-2504.
24. Cai Q, Shu X, Wen W, *et al.* (2004) Genetic polymorphism in the manganese superoxide dismutase gene, antioxidant intake, and breast cancer risk: results from the Shanghai Breast Cancer Study. *Breast Cancer Res*, **6**, 1.
25. Slanger TE, Chang-Claude J, Wang-Gohrke S (2006) Manganese superoxide dismutase Ala-9Val polymorphism, environmental modifiers, and risk of breast cancer in a German population. *Cancer Causes Control*, **17**, 1025-1031.

26. Mitrunen K, Sillanpää P, Kataja V, *et al.* (2001) Association between manganese superoxide dismutase (MnSOD) gene polymorphism and breast cancer risk. *Carcinogenesis*, **22**, 827-829.
27. Han, J. Colditz GA, Hunter, DJ (2007) Manganese superoxide dismutase polymorphism and risk of skin cancer (United States). *Cancer Causes Control*, **18**, 79-89.
28. Miller S, Dykes DD, Polesky HF (1988) A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Res*, **16**, 1215.
29. Bica CG, de Moura da Silva LL, Toscani NV, *et al.* (2009) MnSOD gene polymorphism association with steroid-dependent cancer. *Pathol Oncol Res*, **15**, 19-24.
30. Black H, deGrujil FR, Forbes PD, *et al.* (1997) Photocarcinogenesis: an overview. *J Photochem Photobiol B*, **40**, 29-47.
31. Darr D, Fridovich I (1994) Free radicals in cutaneous biology. *J Invest Dermatol*, **102**, 671-675.
32. Poswig A, Wenk J, Brenneisen P, *et al.* (1999) Adaptive antioxidant response of manganese-superoxide dismutase following repetitive UVA irradiation. *J Invest Dermatol*, **112**, 13-18.
33. Wang S, Wang F, Shi X, *et al.* (2009) Association between manganese superoxide dismutase (MnSOD) Val-9Ala polymorphism and cancer risk—A meta-analysis. *Eur J Cancer*, **45**, 2874-2881.
34. Kang D, Lee KM, Park SK, *et al.* (2007) Functional variant of manganese superoxide dismutase (SOD2 V16A) polymorphism is associated with

prostate cancer risk in the prostate, lung, colorectal, and ovarian cancer study. *Cancer Epidemiol Biomarkers Prev*, **16**, 1581-1586.

TABLE AND FIGURES LEGENDS

Table 1. Clinicopathological features of the case group ($n = 62$).

Table 2. Baseline characteristics and Ala16Val-SOD2 genetic frequencies of melanoma patients and controls.

Fig.1 Genotype frequencies comparison between cases and controls. (A) Significant differences between genotypes frequencies were observed when cases were compared to controls ($P < 0.05$); Ala16Val-SOD2 polymorphism was significantly associated with melanoma risk, but this association was not independent of (B) sex and (C) age. Women and elderly carrying AA genotype are more susceptibility to melanoma.

Fig.2 Relative risk of AA genotype and melanoma associated with sex and age. The bars are representing the odds ratio and 95 % confidence interval. The total population of the study was stratified by sex and age (older ≥ 60 years old; younger < 60 years old) to perform the comparisons of genotype frequencies. Women with AA genotype had 2.49-fold increased melanoma risk, while elderly who carried AA genotype were 4-fold more susceptible to melanoma.

Figure 1

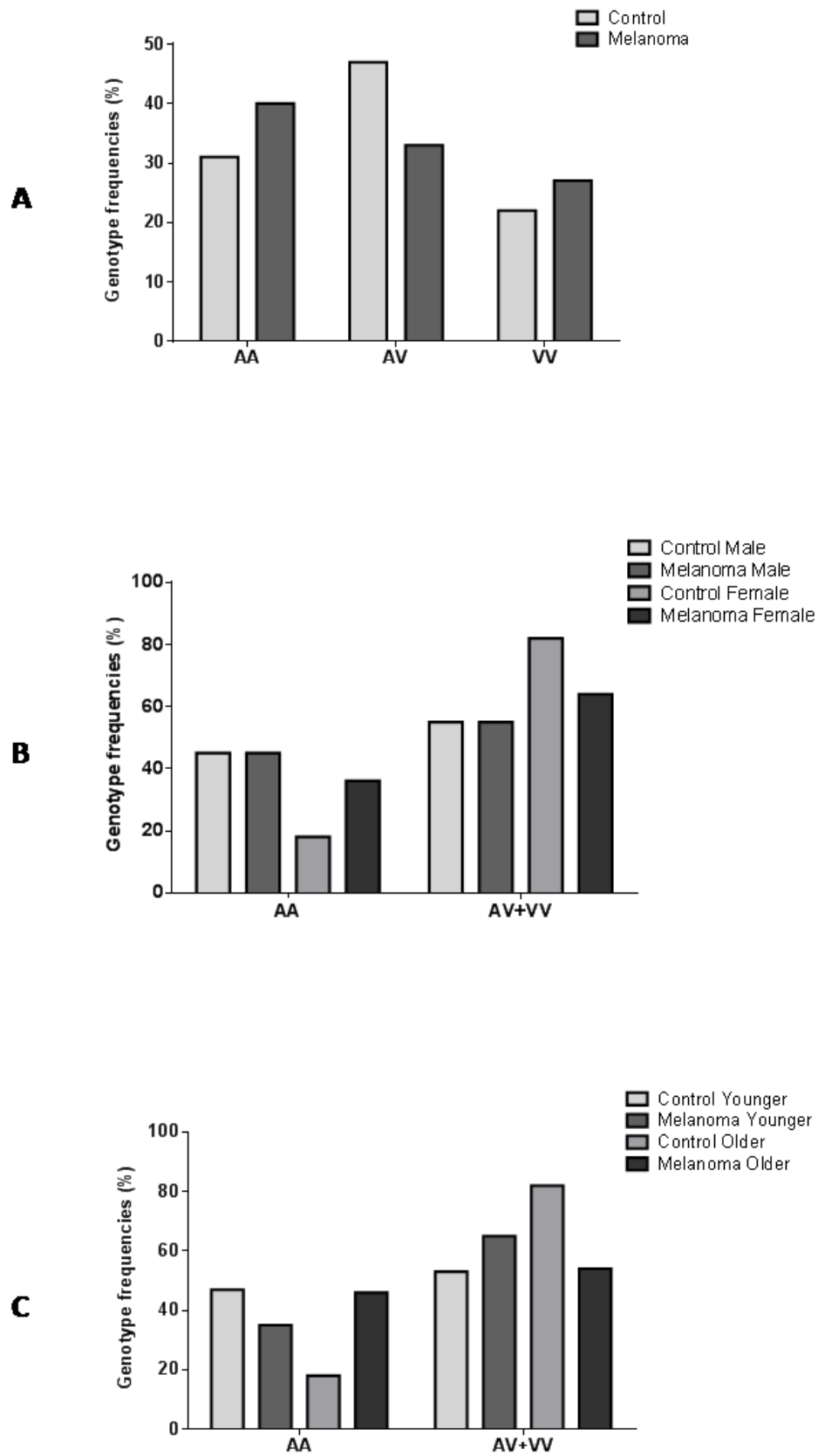


Figure 2

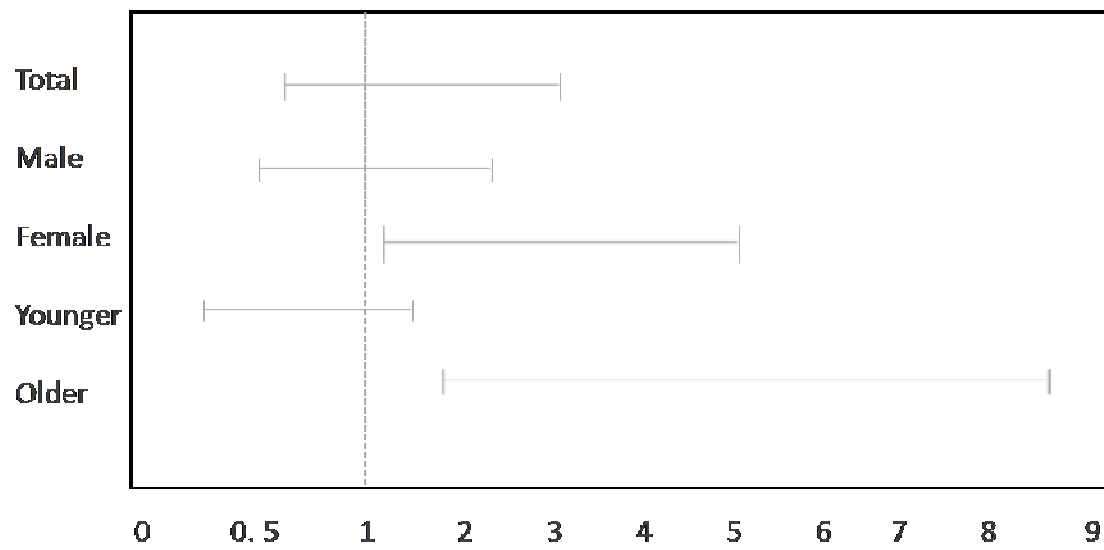


Table 1

Characteristics	N	%
Tumor Site		
Head/neck	13	21
Trunk	32	51.6
Upper limb	10	16.1
Lower limb	7	11.3
Histological subtype		
Superficial spreading	47	75.8
Lentigo maligna	7	11.3
Nodular	5	8.1
Acral lentiginous	2	3.2
Desmoplastic	1	1.6
Clark's level		
I	13	21
II	10	16.1
III	14	22.6
IV	22	35.5
V	3	4.8
Breslow thickness		
T _{is}	13	21
≤ 1 mm	25	40.3
1.01 - 2 mm	10	16.1
2.01 - 4 mm	10	16.1
> 4 mm	4	6.5
Presence of ulceration		
Yes	7	11.3
No	55	88.7
Mitotic rate		
Mean ± SD	1.94 ± 2.47	
Metastasis		
Yes	16	25.8
No	46	74.2

T_{is} = tumor *in situ*; SD = standard deviation

Table 2

Variables	Cases	Controls	<i>P</i>
Age (years, mean $\pm$ SD)	57.55 $\pm$ 11.98	56.17 $\pm$ 15.32	0.49
Sex (<i>n</i>)			0.55
Male	29	237	
Female	33	263	
Genotype <i>n</i> (%)			*0.04
AA	25 (40.3)	156 (31.2)	
VV	17 (27.4)	110 (22)	
AV	20 (32.3)	234 (46.8)	

SD = standard deviation; Statistical test: age comparison between groups was analyzed with Student's *t* test; other variables were analyzed with Chi-square test; Statistical significance was considered when *P* < 0.05.

4. CONSIDERAÇÕES FINAIS

O presente estudo faz parte de um projeto maior intitulado “**Avaliação de Marcadores Prognósticos no Melanoma**”, aprovado pelos Comitês de Ética em Pesquisa (CEP) da UFCSPA (Parecer N° 465.145) e da ISCMPA (Parecer do projeto original N° 436.339; Parecer da emenda N° 843.196). Além da presente tese de doutorado, também está sendo desenvolvida uma dissertação de mestrado como parte integrante deste projeto.

O Núcleo de Melanoma do Hospital Santa Rita da ISCMPA é um centro de referência no tratamento de pacientes com melanoma e coordenado pelo Dr. Felice Riccardi, colaborador deste projeto. A parceria deste serviço com os pesquisadores pertencentes ao Grupo de Pesquisa de Marcadores Tumoriais de Diagnóstico e Prognóstico da UFCSPA teve início em 2013. Um objetivo comum aos dois grupos é o desenvolvimento de biomarcadores de prognóstico para o melanoma capaz de serem incorporados na prática clínica visando melhor classificar os pacientes com alto e baixo risco de doença metastática.

O desenvolvimento de marcadores de prognóstico é extremamente importante para o manejo desta doença. O biomarcador ideal deve ser sensível, específico, confiável, de rápida análise, de baixo custo e deve "agregar valor", em termos prognósticos ou terapêuticos, às ferramentas atuais de avaliação.

A vantagem da triagem precoce de pacientes com alto risco é a oportunidade de identificar aqueles que necessitam de um acompanhamento clínico intensificado ou mesmo tratamentos adjuvantes, enquanto que, por outro lado, minimizaria intervenções desnecessárias em pacientes de baixo risco.

Na busca por artigos para compor a revisão sistemática de marcadores genéticos e epigenéticos no melanoma (artigo I da tese), ficou claro que embora tenha havido recente progresso no entendimento dos mecanismos genéticos por trás do melanoma, poucos biomarcadores provaram serem clinicamente significantes e, por isto, nenhum deles foi incorporado à rotina clínica até hoje. Muitos pesquisadores da área vêm apostando na abordagem de múltiplos marcadores para acessar o desfecho dos pacientes portadores de melanoma, principalmente aqueles que avaliam a expressão de microRNAs no sangue e tecido, por se tratarem de amostras facilmente obtidas de materiais biológicos escassos e/ou mal-preservados.

Devido a um maior conhecimento da doença por parte da população, nos últimos anos há uma tendência de diagnóstico precoce da doença. Na experiência obtida durante a condução deste estudo, foi notória a identificação de um grande número de pacientes com lesões finas ou *in situ*, usualmente curáveis cirurgicamente. No entanto, houve um subgrupo de pacientes com melanomas igualmente finos que vieram a óbito após o desenvolvimento de metástases sistêmicas, salientando o caráter heterogêneo desta neoplasia que não pode ser acessado em sua totalidade pelo sistema de classificação da AJCC. Portanto, acreditamos que este fato justifique o desenvolvimento de novos marcadores de prognóstico.

Biomarcadores sanguíneos são atrativos por se tratarem de métodos fáceis de obtenção de amostras e minimamente invasivos comparados com biópsias de tecido, por exemplo. Por esta razão, optamos por avaliar o potencial da quantificação de *cfDNA* no prognóstico do melanoma.

Em uma análise preliminar apresentada em forma de resumo e pôster no Congresso Brasileiro de Melanoma (2015), o trabalho intitulado “Potencial marcador plasmático de *screening* em pacientes com melanoma” foi nomeado como um dos destaques do evento e escolhido para apresentação oral, a qual foi realizada pelo Dr. Felice Riccardi. Nosso estudo foi reconhecido como segundo melhor trabalho do evento, além de recebermos premiação financeira.

Com a inclusão de novos casos, além da incorporação da análise para avaliar o status inflamatório dos pacientes, escrevemos o artigo II da tese que foi submetido para publicação na revista científica *Biomarkers*. Como principais resultados observados, destacamos: (1) níveis de *cfDNA* significativamente aumentado nos pacientes com melanoma em relação aos indivíduos saudáveis; (2) níveis aumentados de citocinas pró-inflamatórias e reduzidos da citocina anti-inflamatória nos casos quando comparado aos controles, corroborando o potencial do *cfDNA* como marcador inflamatório neste câncer; e (3) demonstramos o impacto do tratamento cirúrgico na redução dos níveis de *cfDNA*. Consideramos nossos achados cientificamente relevantes por este ser o único estudo na literatura a quantificar este marcador com amostras pré e pós-cirúrgicas em pacientes com melanoma.

Seguindo a linha de marcadores relacionados à inflamação no câncer, avaliamos a associação do melanoma com o polimorfismo da enzima

antioxidante superóxido dismutase dependente de manganês (MnSOD). O que encontramos, também de forma inédita na literatura, foi que o genótipo AA (Ala/Ala) do polimorfismo Ala16Val-SOD2, está associado com maior risco de melanoma. Entretanto, esta associação é influenciada pelo sexo e idade. Sendo assim, observamos que mulheres e idosos portadores do genótipo AA estão mais susceptíveis ao câncer. A partir destes resultados, foi desenvolvido um artigo científico encaminhado para a revista *Pathology and Oncology Research*.

5. ANEXOS

5.1 Parecer do Comitê de Ética em Pesquisa da UFCSPA

UNIVERSIDADE FEDERAL DE
CIÊNCIAS DA SAÚDE DE
PORTO ALEGRE



PARECER CONSUBSTANCIADO DO CEP

Elaborado pela Instituição Coparticipante

DADOS DO PROJETO DE PESQUISA

Título da Pesquisa: Avaliação de Marcadores Prognósticos no Melanoma

Pesquisador: Claudia Giuliano Bica

Área Temática:

Versão: 3

CAAE: 14404513.4.0000.5335

Instituição Proponente: Irmandade da Santa Casa de Misericórdia de Porto Alegre - ISCMPA

Patrocinador Principal: Universidade Federal de Ciências da Saúde de Porto Alegre

DADOS DO PARECER

Número do Parecer: 465.145

Data da Relatoria: 21/11/2013

Apresentação do Projeto:

Trata-se de um projeto de doutorado da aluna Munique Pereira Mendonça, orientado pela Profa. Cláudia G. Bica. Neste estudo, pacientes com diagnóstico de melanoma, provenientes do Núcleo de Melanoma do Hospital Santa Rita, serão seguidos semestralmente durante dois anos e avaliados quanto a identificação de potenciais biomarcadores que possam influenciar no prognóstico do mesmo. A presença de biomarcadores será comparada com população controle sem melanoma.

O referido projeto será realizado no Complexo da Santa Casa onde já recebeu aprovação do CEP. A Universidade Federal de Ciências da Saúde de Porto Alegre (UFCSPA) será centro coparticipante uma vez que algumas análises moleculares e bioquímicas serão realizadas no Laboratório de Patologia da Pós-Graduação da UFCSPA.

Objetivo da Pesquisa:

O objetivo geral do projeto é avaliar o potencial de marcadores moleculares no prognóstico do melanoma maligno cutâneo.

Os objetivos secundários do estudo são: I. Comparar os parâmetros inflamatórios/oxidativos e a quantidade de cell-free DNA entre indivíduos hígidos e portadores de melanoma. II. Quantificar os níveis de cell-free DNA (cfDNA) de pacientes com melanoma; III. Estabelecer o perfil inflamatório/oxidativo dos pacientes portadores da doença; IV. Identificar a expressão de moléculas de adesão nas peças cirúrgicas.

UNIVERSIDADE FEDERAL DE
CIÊNCIAS DA SAÚDE DE
PORTO ALEGRE



Continuação do Parecer: 465.145

Avaliação dos Riscos e Benefícios:

o projeto não oferece maiores riscos aos pacientes, uma vez que serão aproveitados a sobra de sangue dos exames de rotina realizados no Hospital Santa Rita da ISCMPA para o estudo. Exames esses feitos assistencialmente, rotineiramente.

Embora sem benefícios diretos poderá auxiliar na identificação de fatores que possam auxiliar prognóstico da doença.

Comentários e Considerações sobre a Pesquisa:

O projeto é relevante e exequível. O mesmo já foi aprovado pelo CEP da Santa Casa. A Universidade Federal de Ciências da Saúde de Porto Alegre (UFCSPA) será centro coparticipante uma vez que algumas análises moleculares e bioquímicas serão realizadas no Laboratório de Patologia da Pós-Graduação da UFCSPA.

Considerações sobre os Termos de apresentação obrigatória:

Documentos regulatórios estão adequados. Possui carta de concordância das instituições.

Recomendações:

Conclusões ou Pendências e Lista de Inadequações:

O projeto está dentro dos padrões e julgo que o mesmo possa ser aprovado.

Situação do Parecer:

Aprovado

Necessita Apreciação da CONEP:

Não

Considerações Finais a critério do CEP:

Data de início 23/10/13

Data de término 06/03/17

Endereço: Rua Sarmento Leite ,245

Bairro:

CEP: 90.050-170

UF: RS

Município: PORTO ALEGRE

Telefone: (513)303 -8804

E-mail: cep@ufcspa.edu.br

UNIVERSIDADE FEDERAL DE
CIÊNCIAS DA SAÚDE DE
PORTO ALEGRE



Continuação do Parecer: 465.145

PORTO ALEGRE, 22 de Novembro de 2013

Assinador por:
José Geraldo Vernet Taborda
(Coordenador)

Endereço: Rua Sarmento Leite ,245

Bairro:

CEP: 90.050-170

UF: RS

Município: PORTO ALEGRE

Telefone: (513)303 -8804

E-mail: cep@ufcspa.edu.br

5.2 Parecer do Comitê de Ética em Pesquisa da ISCMPA versão 3

IRMANDADE DA SANTA CASA
DE MISERICORDIA DE PORTO
ALEGRE - ISCMPA



PARECER CONSUBSTANCIADO DO CEP

DADOS DO PROJETO DE PESQUISA

Título da Pesquisa: Avaliação de Marcadores Prognósticos no Melanoma

Pesquisador: Claudia Giuliano Bica

Área Temática:

Versão: 3

CAAE: 14404513.4.0000.5335

Instituição Proponente: Irmandade da Santa Casa de Misericórdia de Porto Alegre - ISCMPA

Patrocinador Principal: Universidade Federal de Ciências da Saúde de Porto Alegre

DADOS DO PARECER

Número do Parecer: 436.339

Data da Relatoria: 25/09/2013

Apresentação do Projeto:

Projeto de pós-graduação em nível de doutorado de MUNIQUE PEREIRA MENDONÇA. ORIENTADORA: DRA. CLÁUDIA G. BICA. Estudo Prospectivo de Coorte com pacientes portadores de melanoma maligno cutâneo sem metástases do Núcleo de Melanoma do Hospital Santa Rita. Os pacientes serão acompanhados semestralmente, totalizando 4 amostragens de cada paciente.

Objetivo da Pesquisa:

Objetivo Primário: Avaliar o potencial de marcadores moleculares no prognóstico do melanoma maligno cutâneo.

Objetivo Secundário: I. Comparar os parâmetros inflamatórios/oxidativos e a quantidade de cell-free DNA entre indivíduos hígidos e portadores de melanoma. II. Quantificar os níveis de cell-free DNA (cfDNA) de pacientes com melanoma; III. Estabelecer o perfil inflamatório/oxidativo dos pacientes portadores da doença; IV. Identificar a expressão de moléculas de adesão nas peças cirúrgicas.

Avaliação dos Riscos e Benefícios:

Riscos: Segundo os autores, o projeto não oferece maiores riscos aos pacientes, uma vez que serão aproveitados a sobra de sangue dos exames de rotina realizados no Hospital Santa Rita da ISCMPA para o estudo. Do mesmo modo, serão utilizadas peças cirúrgicas dos pacientes, as quais

IRMANDADE DA SANTA CASA DE MISERICORDIA DE PORTO ALEGRE - ISCMPA



Continuação do Parecer: 436.339

serão devolvidas ao Laboratório de Patologia da Santa Casa assim que o estudo for concluído. NO TCLE foi descrito que as informações contidas no prontuário serão de acesso da equipe do estudo, e que a equipe garante a realização de todas as medidas cabíveis para que os dados sejam mantidos em sigilo absoluto. Benefícios: Os pacientes não terão nenhum benefício direto com esta pesquisa, entretanto os dados científicos gerados poderão contribuir para o maior conhecimento desta importante neoplasia.

Comentários e Considerações sobre a Pesquisa:

Estudo prospectivo de coorte.

Tamanho da Amostra: 170 (85 casos e 85 controles), orçamento R\$ 10.595,00, apoio financeiro Universidade Federal de Ciências da Saúde de Porto Alegre. Local de realização do projeto: Hospital Santa Rita (Núcleo de melanoma e Laboratório de Patologia) e Laboratório Central e Análises Clínicas - Irmandade da Santa Casa de Misericórdia de Porto Alegre; Laboratório de Patologia da Universidade Federal de Ciências da Saúde de Porto Alegre (UFCSPA) e Laboratório de Biogenômica da Universidade Federal de Santa Maria (UFSM).

Quem pagará as despesas: Verbas dos pesquisadores, agência de fomento ao qual o projeto será submetido e apoio da Universidade.

Critérios de Inclusão: Casos: serão incluídos à pesquisa pacientes portadores de melanoma maligno cutâneo sem metástases e maiores de idade, após a assinatura de Termo de Consentimento Livre e Esclarecido (TCLE).

Controles: serão incluídos indivíduos hígidos, pareados aos casos por sexo e idade e cor da pele, oriundos da comunidade universitária da UFCSPA ou da UFSM (Universidade Federal de Santa Maria), onde a colaboradora do projeto atua em cooperação, após a assinatura Termo de Consentimento Livre e Esclarecido (TCLE).

Critério de Exclusão: Ficarão excluídos deste estudo pacientes com melanoma metastático, menores de idade e que não concordarem em assinar o TCLE.

Considerações sobre os Termos de apresentação obrigatória:

Foi apresentada declaração de isenção de ônus à instituição. Declaração de confidencialidade do sujeito no estudo e declaração de uso de dados e materiais.

TCLE redigido de forma clara, com 3 páginas.

Recomendações:

Solicita-se e aguarda-se modificação no TCLE a respeito de comparação com estudos futuros,

IRMANDADE DA SANTA CASA
DE MISERICORDIA DE PORTO
ALEGRE - ISCMPA



Continuação do Parecer: 436.339

onde deve ser colocado a seguinte consideração, na solicitação do paciente de autorizar ou não: "desde que os futuros projetos, sejam aprovados pelo CEP."

Conclusões ou Pendências e Lista de Inadequações:

Pendências anteriormente solicitadas foram atendidas.

Aguarda-se inclusão de frase no TCLE, conforme descrito nas "recomendações" que são necessárias novas aprovações do CEP em projetos futuros.

Situação do Parecer:

Aprovado

Necessita Apreciação da CONEP:

Não

Considerações Finais a critério do CEP:

Após reavaliação do protocolo acima descrito, o presente comitê não encontrou óbices quanto ao desenvolvimento do estudo em nossa Instituição e poderá ser iniciado a partir da data deste parecer.

Obs.: 1 - O pesquisador responsável deve encaminhar à este CEP, Relatórios de Andamento dos Projetos desenvolvidos na ISCMPA. Relatórios Parciais (pesquisas com duração superior à 6 meses), Relatórios Finais (ao término da pesquisa) e os Resultados Obtidos (cópia da publicação).

2 - Para o início do projeto de pesquisa, o investigador deverá apresentar a chefia do serviço (onde será realizada a pesquisa), o Parecer Consubstanciado de aprovação do protocolo pelo Comitê de Ética.

Endereço: R. Profº Annes Dias, 285 Hosp. Dom Vicente Scherer
Bairro: 6º andar - Centro **CEP:** 90.020-090
UF: RS **Município:** PORTO ALEGRE
Telefone: (51)3214-8571 **Fax:** (51)3214-8571 **E-mail:** cep@santacasa.tche.br

IRMANDADE DA SANTA CASA
DE MISERICORDIA DE PORTO
ALEGRE - ISCMPA



Continuação do Parecer: 436.339

PORTO ALEGRE, 25 de Outubro de 2013

Assinador por:
Claudio Teloken
(Coordenador)

Endereço: R. Profº Annes Dias, 285 Hosp. Dom Vicente Scherer
Bairro: 6º andar - Centro **CEP:** 90.020-090
UF: RS **Município:** PORTO ALEGRE
Telefone: (51)3214-8571 **Fax:** (51)3214-8571 **E-mail:** cep@santacasa.tche.br

5.3 Parecer do Comitê de Ética em Pesquisa da ISCMPA versão 4

IRMANDADE DA SANTA CASA
DE MISERICORDIA DE PORTO
ALEGRE - ISCMPA



PARECER CONSUBSTANCIADO DO CEP

DADOS DO PROJETO DE PESQUISA

Título da Pesquisa: Avaliação de Marcadores Prognósticos no Melanoma

Pesquisador: Claudia Giuliano Bica

Área Temática:

Versão: 4

CAAE: 14404513.4.0000.5335

Instituição Proponente: Irmandade da Santa Casa de Misericórdia de Porto Alegre - ISCMPA

Patrocinador Principal: Universidade Federal de Ciências da Saúde de Porto Alegre

DADOS DO PARECER

Número do Parecer: 843.196

Data da Relatoria: 25/09/2014

Apresentação do Projeto:

Solicitação de Emenda, referente:

-Projeto modificado 20/08/2014: critérios de inclusão para pacientes.

-Termo de Consentimento Livre e Esclarecido, modificado - Versão 3.0 datada de 23/09/2013.

_Inclusão de Pesquisador: Luciana Schuch.

Objetivo da Pesquisa:

Já descrito no parecer anterior.

Avaliação dos Riscos e Benefícios:

Adequados e já apresentados.

Comentários e Considerações sobre a Pesquisa:

Solicitação de Emenda referente ao critérios de Inclusão, TCLE modificado e inclusão de pesquisadora, todos apresentados em anexo e adequados.

Considerações sobre os Termos de apresentação obrigatória:

Adequados.

Recomendações:

Não se aplica.

IRMANDADE DA SANTA CASA
DE MISERICORDIA DE PORTO
ALEGRE - ISCMPA



Continuação do Parecer: 843.196

Conclusões ou Pendências e Lista de Inadequações:

Solicitamos no TCLE, modificado - Versão 3.0 datada de 23/09/13, retirar onde cita a resolução 196/96, trocar para Resolução nº.: 466/12 vigente!.

Situação do Parecer:

Aprovado

Necessita Apreciação da CONEP:

Não

Considerações Finais a critério do CEP:

Após reavaliação do protocolo acima descrito, o presente comitê não encontrou óbices quanto ao desenvolvimento do estudo em nossa Instituição e poderá ser iniciado a partir da data deste parecer.

-Projeto modificado 20/08/2014: critérios de inclusão para pacientes.

-Termo de Consentimento Livre e Esclarecido, modificado - Versão 3.0 datada de 23/09/2013.

_Inclusão de Pesquisadora: Luciana Schuch.

Obs.: 1 - O pesquisador responsável deve encaminhar à este CEP, Relatórios de Andamento dos Projetos desenvolvidos na ISCMPA. Relatórios Parciais (pesquisas com duração superior à 6 meses), Relatórios Finais (ao término da pesquisa) e os Resultados Obtidos (cópia da publicação).

2 – Para o início do projeto de pesquisa, o investigador deverá apresentar a chefia do serviço (onde será realizada a pesquisa), o Parecer Consubstanciado de aprovação do protocolo pelo Comitê de Ética.

Endereço: R. Profº Annes Dias, 285 Hosp. Dom Vicente Scherer

Bairro: 6º andar - Centro

CEP: 90.020-090

UF: RS

Município: PORTO ALEGRE

Telefone: (51)3214-8571

Fax: (51)3214-8571

E-mail: cep@santacasa.tche.br

IRMANDADE DA SANTA CASA
DE MISERICORDIA DE PORTO
ALEGRE - ISCMPA



Continuação do Parecer: 843.196

PORTO ALEGRE, 23 de Outubro de 2014

Assinado por:
Claudio Teloken
(Coordenador)

Endereço: R. Profª Annes Dias, 285 Hosp. Dom Vicente Scherer
Bairro: 6º andar - Centro **CEP:** 90.020-090
UF: RS **Município:** PORTO ALEGRE
Telefone: (51) 3214-8571 **Fax:** (51) 3214-8571 **E-mail:** cep@santacasa.tche.br

5.4 Comprovante de submissão de artigo à revista Pathology & Oncology Research



Munique Mendonça <muniquepm@gmail.com>

[Fwd: PORE-D-17-00535 - Submission Confirmation]

claudia@ufcspa.edu.br <claudia@ufcspa.edu.br>

6 de novembro de 2017 07:45

Para: muniquepm@gmail.com

----- Mensagem Original -----

Assunto: PORE-D-17-00535 - Submission Confirmation

De: "PORE" <em@editorialmanager.com>

Data: Seg, Novembro 6, 2017 7:45 pm

Para: "Claudia Giuliano Bica" <claudia@ufcspa.edu.br>

Dear Mrs Giuliano Bica,

Thank you for submitting your manuscript,
"Genetic and epigenetic biomarkers in melanoma prognosis: a last five
years update", to Pathology & Oncology Research

The submission id is: PORE-D-17-00535
Please refer to this number in any future correspondence.

During the review process, you can keep track of the status of your
manuscript by accessing the following:

Your username is: Claudia Bica

If you forgot your password, you can click the 'Send Login Details' link
on the EM Login page at <http://pore.edmgr.com/>.

Springer offers authors the option of making their articles available with
open access via our Open Choice programme. We advise you to familiarise
yourself with the details of Springer Open Choice in advance, to be able
to decide quickly should your paper be accepted for publication. Further
information can be found at www.springer.com/openchoice.

With kind regards,

Journals Editorial Office PORE
Springer

5.5 Normas da revista Pathology & Oncology Research



Pathology & Oncology Research

Official Journal of the Arányi Lajos Foundation

Editors-in-Chief: L. **Kopper**; J. **Tímár**

ISSN: 1219-4956 (print version)

ISSN: 1532-2807 (electronic version)

Journal no. 12253

Instructions for authors

LANGUAGE

We appreciate any efforts that you make to ensure that the language is corrected before submission. This will greatly improve the legibility of your paper if English is not your first language.

TYPES OF PAPERS

The journal publishes Original Articles, Reviews, Short Communications and Letters to the Editor.

Case Reports are not considered for publication.

Short Communications

The length and the number of figures are limited to 2000 words (text, including 10 references), plus 2 figures (which can be multi-panel figures) and a maximum of two tables. The format remains the same as for regular articles, except that we ask the authors to be very concise in writing their introduction and discussion.

Reviews and Commentaries

Solicited and non-solicited papers on topical subjects will be published.

Letters to the Editor

The length is limited to a maximum of 1000 words and 5 references.

MANUSCRIPT SUBMISSION

Manuscript Submission

Submission of a manuscript implies: that the work described has not been published before; that it is not under consideration for publication anywhere else; that its publication has been approved by all co-authors, if any, as well as by the responsible authorities – tacitly or explicitly

– at the institute where the work has been carried out. The publisher will not be held legally responsible should there be any claims for compensation.

Permissions

Authors wishing to include figures, tables, or text passages that have already been published elsewhere are required to obtain permission from the copyright owner(s) for both the print and online format and to include evidence that such permission has been granted when submitting their papers. Any material received without such evidence will be assumed to originate from the authors.

Online Submission

Please follow the hyperlink “Submit online” on the right and upload all of your manuscript files following the instructions given on the screen.

TITLE PAGE

Title Page

The title page should include:

- The name(s) of the author(s)
- A concise and informative title
- The affiliation(s) and address(es) of the author(s)
- The e-mail address, and telephone number(s) of the corresponding author
- If available, the 16-digit ORCID of the author(s)

Abstract

Please provide an abstract of 150 to 250 words. The abstract should not contain any undefined abbreviations or unspecified references.

Keywords

Please provide 4 to 6 keywords which can be used for indexing purposes.

TEXT

Text Formatting

Manuscripts should be submitted in Word.

- Use a normal, plain font (e.g., 10-point Times Roman) for text.
- Use italics for emphasis.
- Use the automatic page numbering function to number the pages.
- Do not use field functions.
- Use tab stops or other commands for indents, not the space bar.
- Use the table function, not spreadsheets, to make tables.
- Use the equation editor or MathType for equations.
- Save your file in docx format (Word 2007 or higher) or doc format (older Word versions).

Manuscripts with mathematical content can also be submitted in LaTeX.

- [LaTeX macro package \(zip, 182 kB\)](#)

Headings

Please use no more than three levels of displayed headings.

Abbreviations

Abbreviations should be defined at first mention and used consistently thereafter.

Footnotes

Footnotes can be used to give additional information, which may include the citation of a reference included in the reference list. They should not consist solely of a reference citation, and they should never include the bibliographic details of a reference. They should also not contain any figures or tables.

Footnotes to the text are numbered consecutively; those to tables should be indicated by superscript lower-case letters (or asterisks for significance values and other statistical data). Footnotes to the title or the authors of the article are not given reference symbols.

Always use footnotes instead of endnotes.

Acknowledgments

Acknowledgments of people, grants, funds, etc. should be placed in a separate section on the title page. The names of funding organizations should be written in full.

SCIENTIFIC STYLE

Please always use internationally accepted signs and symbols for units (SI units).

REFERENCES

Citation

Reference citations in the text should be identified by numbers in square brackets. Some examples:

1. Negotiation research spans many disciplines [3].
2. This result was later contradicted by Becker and Seligman [5].
3. This effect has been widely studied [1-3, 7].

Reference list

The list of references should only include works that are cited in the text and that have been published or accepted for publication. Personal communications and unpublished works should only be mentioned in the text. Do not use footnotes or endnotes as a substitute for a reference list.

The entries in the list should be numbered consecutively.

- Journal article
Gamelin FX, Baquet G, Berthoin S, Thevenet D, Nourry C, Nottin S, Bosquet L (2009) Effect of high intensity intermittent training on heart rate variability in prepubescent children. *Eur J Appl Physiol* 105:731-738. doi: 10.1007/s00421-008-0955-8
Ideally, the names of all authors should be provided, but the usage of “et al” in long author lists will also be accepted:
Smith J, Jones M Jr, Houghton L et al (1999) Future of health insurance. *N Engl J Med* 341:325–329
- Article by DOI

Slifka MK, Whitton JL (2000) Clinical implications of dysregulated cytokine production. *J Mol Med*. doi:10.1007/s001090000086

- Book
South J, Blass B (2001) *The future of modern genomics*. Blackwell, London
- Book chapter
Brown B, Aaron M (2001) The politics of nature. In: Smith J (ed) *The rise of modern genomics*, 3rd edn. Wiley, New York, pp 230-257
- Online document
Cartwright J (2007) Big stars have weather too. IOP Publishing PhysicsWeb. <http://physicsweb.org/articles/news/11/6/16/1>. Accessed 26 June 2007
- Dissertation
Trent JW (1975) *Experimental acute renal failure*. Dissertation, University of California

Always use the standard abbreviation of a journal's name according to the ISSN List of Title Word Abbreviations, see

- [ISSN.org LTWA](http://www.issn.org/LTWA)

If you are unsure, please use the full journal title.

For authors using EndNote, Springer provides an output style that supports the formatting of in-text citations and reference list.

- [EndNote style \(zip, 2 kB\)](#)
Authors preparing their manuscript in LaTeX can use the bibtex file `spbasic.bst` which is included in Springer's LaTeX macro package.

TABLES

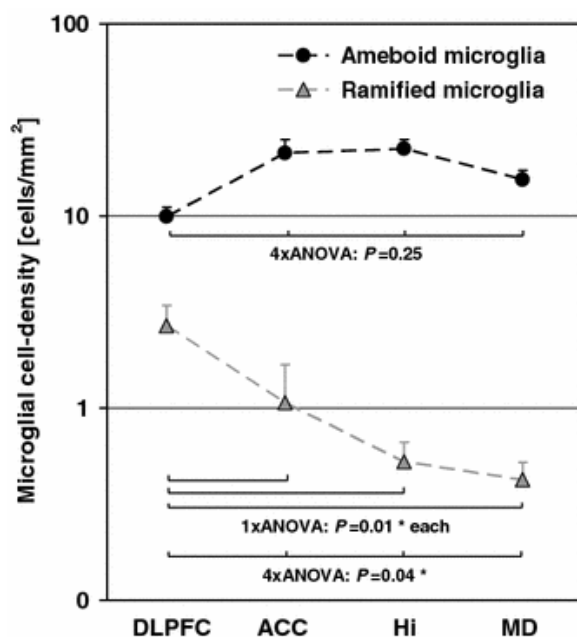
- All tables are to be numbered using Arabic numerals.
- Tables should always be cited in text in consecutive numerical order.
- For each table, please supply a table caption (title) explaining the components of the table.
- Identify any previously published material by giving the original source in the form of a reference at the end of the table caption.
- Footnotes to tables should be indicated by superscript lower-case letters (or asterisks for significance values and other statistical data) and included beneath the table body.

ARTWORK AND ILLUSTRATIONS GUIDELINES

Electronic Figure Submission

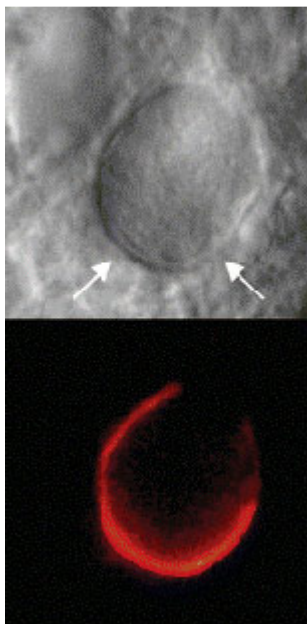
- Supply all figures electronically.
- Indicate what graphics program was used to create the artwork.
- For vector graphics, the preferred format is EPS; for halftones, please use TIFF format. MSOffice files are also acceptable.
- Vector graphics containing fonts must have the fonts embedded in the files.
- Name your figure files with "Fig" and the figure number, e.g., Fig1.eps.

Line Art



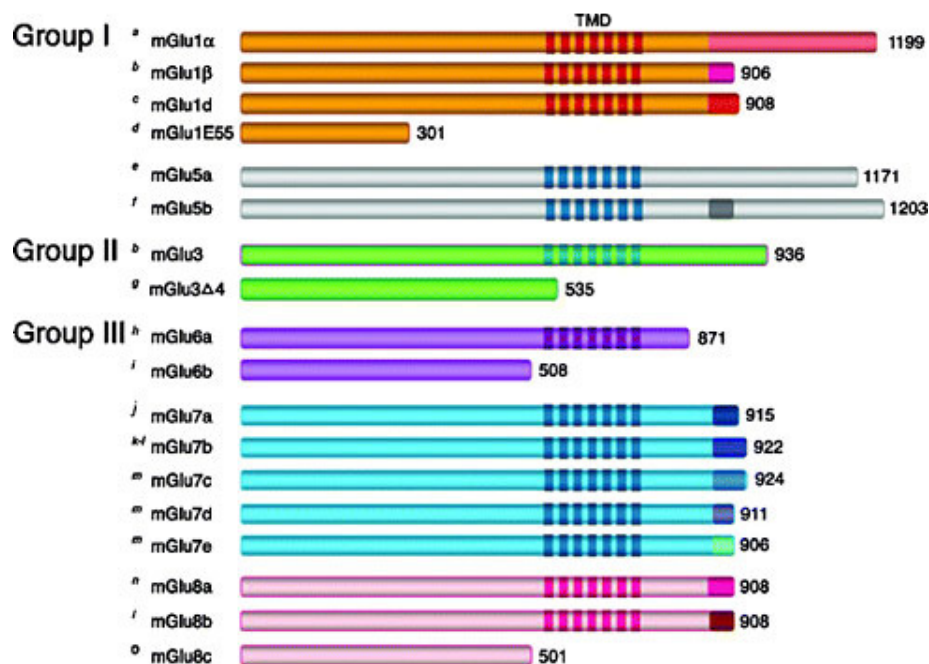
- Definition: Black and white graphic with no shading.
- Do not use faint lines and/or lettering and check that all lines and lettering within the figures are legible at final size.
- All lines should be at least 0.1 mm (0.3 pt) wide.
- Scanned line drawings and line drawings in bitmap format should have a minimum resolution of 1200 dpi.
- Vector graphics containing fonts must have the fonts embedded in the files.

Halftone Art



- Definition: Photographs, drawings, or paintings with fine shading, etc.
- If any magnification is used in the photographs, indicate this by using scale bars within the figures themselves.
- Halftones should have a minimum resolution of 300 dpi.

Combination Art



- Definition: a combination of halftone and line art, e.g., halftones containing line drawing, extensive lettering, color diagrams, etc.
- Combination artwork should have a minimum resolution of 600 dpi.

Color Art

- Color art is free of charge for online publication.
- If black and white will be shown in the print version, make sure that the main information will still be visible. Many colors are not distinguishable from one another when converted to black and white. A simple way to check this is to make a xerographic copy to see if the necessary distinctions between the different colors are still apparent.
- If the figures will be printed in black and white, do not refer to color in the captions.
- Color illustrations should be submitted as RGB (8 bits per channel).

Figure Lettering

- To add lettering, it is best to use Helvetica or Arial (sans serif fonts).
- Keep lettering consistently sized throughout your final-sized artwork, usually about 2–3 mm (8–12 pt).
- Variance of type size within an illustration should be minimal, e.g., do not use 8-pt type on an axis and 20-pt type for the axis label.
- Avoid effects such as shading, outline letters, etc.
- Do not include titles or captions within your illustrations.

Figure Numbering

- All figures are to be numbered using Arabic numerals.
- Figures should always be cited in text in consecutive numerical order.
- Figure parts should be denoted by lowercase letters (a, b, c, etc.).
- If an appendix appears in your article and it contains one or more figures, continue the consecutive numbering of the main text. Do not number the appendix figures, "A1, A2, A3, etc." Figures in online appendices (Electronic Supplementary Material) should, however, be numbered separately.

Figure Captions

- Each figure should have a concise caption describing accurately what the figure depicts. Include the captions in the text file of the manuscript, not in the figure file.
- Figure captions begin with the term **Fig.** in bold type, followed by the figure number, also in bold type.
- No punctuation is to be included after the number, nor is any punctuation to be placed at the end of the caption.
- Identify all elements found in the figure in the figure caption; and use boxes, circles, etc., as coordinate points in graphs.
- Identify previously published material by giving the original source in the form of a reference citation at the end of the figure caption.

Figure Placement and Size

- Figures should be submitted separately from the text, if possible.
- When preparing your figures, size figures to fit in the column width.
- For most journals the figures should be 39 mm, 84 mm, 129 mm, or 174 mm wide and not higher than 234 mm.
- For books and book-sized journals, the figures should be 80 mm or 122 mm wide and not higher than 198 mm.

Permissions

If you include figures that have already been published elsewhere, you must obtain permission from the copyright owner(s) for both the print and online format. Please be aware that some publishers do not grant electronic rights for free and that Springer will not be able to refund any costs that may have occurred to receive these permissions. In such cases, material from other sources should be used.

Accessibility

In order to give people of all abilities and disabilities access to the content of your figures, please make sure that

- All figures have descriptive captions (blind users could then use a text-to-speech software or a text-to-Braille hardware)
- Patterns are used instead of or in addition to colors for conveying information (colorblind users would then be able to distinguish the visual elements)
- Any figure lettering has a contrast ratio of at least 4.5:1

ELECTRONIC SUPPLEMENTARY MATERIAL

Springer accepts electronic multimedia files (animations, movies, audio, etc.) and other supplementary files to be published online along with an article or a book chapter. This feature can add dimension to the author's article, as certain information cannot be printed or is more convenient in electronic form.

Before submitting research datasets as electronic supplementary material, authors should read the journal's Research data policy. We encourage research data to be archived in data repositories wherever possible.

Submission

- Supply all supplementary material in standard file formats.
- Please include in each file the following information: article title, journal name, author names; affiliation and e-mail address of the corresponding author.

- To accommodate user downloads, please keep in mind that larger-sized files may require very long download times and that some users may experience other problems during downloading.

Audio, Video, and Animations

- Aspect ratio: 16:9 or 4:3
- Maximum file size: 25 GB
- Minimum video duration: 1 sec
- Supported file formats: avi, wmv, mp4, mov, m2p, mp2, mpg, mpeg, flv, mxf, mts, m4v, 3gp

Text and Presentations

- Submit your material in PDF format; .doc or .ppt files are not suitable for long-term viability.
- A collection of figures may also be combined in a PDF file.

Spreadsheets

- Spreadsheets should be converted to PDF if no interaction with the data is intended.
- If the readers should be encouraged to make their own calculations, spreadsheets should be submitted as .xls files (MS Excel).

Specialized Formats

- Specialized format such as .pdb (chemical), .wrl (VRML), .nb (Mathematica notebook), and .tex can also be supplied.

Collecting Multiple Files

- It is possible to collect multiple files in a .zip or .gz file.

Numbering

- If supplying any supplementary material, the text must make specific mention of the material as a citation, similar to that of figures and tables.
- Refer to the supplementary files as “Online Resource”, e.g., “... as shown in the animation (Online Resource 3)”, “... additional data are given in Online Resource 4”.
- Name the files consecutively, e.g. “ESM_3.mpg”, “ESM_4.pdf”.

Captions

- For each supplementary material, please supply a concise caption describing the content of the file.

Processing of supplementary files

- Electronic supplementary material will be published as received from the author without any conversion, editing, or reformatting.

Accessibility

In order to give people of all abilities and disabilities access to the content of your supplementary files, please make sure that

- The manuscript contains a descriptive caption for each supplementary material
- Video files do not contain anything that flashes more than three times per second (so that users prone to seizures caused by such effects are not put at risk)

ENGLISH LANGUAGE EDITING

For editors and reviewers to accurately assess the work presented in your manuscript you need to ensure the English language is of sufficient quality to be understood. If you need help with writing in English you should consider:

- Asking a colleague who is a native English speaker to review your manuscript for clarity.
- Visiting the English language tutorial which covers the common mistakes when writing in English.
- Using a professional language editing service where editors will improve the English to ensure that your meaning is clear and identify problems that require your review. Two such services are provided by our affiliates Nature Research Editing Service and American Journal Experts.
- [English language tutorial](#)
- [Nature Research Editing Service](#)
- [American Journal Experts](#)

Please note that the use of a language editing service is not a requirement for publication in this journal and does not imply or guarantee that the article will be selected for peer review or accepted.

If your manuscript is accepted it will be checked by our copyeditors for spelling and formal style before publication.

ETHICAL RESPONSIBILITIES OF AUTHORS

This journal is committed to upholding the integrity of the scientific record. As a member of the Committee on Publication Ethics (COPE) the journal will follow the COPE guidelines on how to deal with potential acts of misconduct.

Authors should refrain from misrepresenting research results which could damage the trust in the journal, the professionalism of scientific authorship, and ultimately the entire scientific endeavour. Maintaining integrity of the research and its presentation can be achieved by following the rules of good scientific practice, which include:

- The manuscript has not been submitted to more than one journal for simultaneous consideration.
- The manuscript has not been published previously (partly or in full), unless the new work concerns an expansion of previous work (please provide transparency on the re-use of material to avoid the hint of text-recycling (“self-plagiarism”).
- A single study is not split up into several parts to increase the quantity of submissions and submitted to various journals or to one journal over time (e.g. “salami-publishing”).
- No data have been fabricated or manipulated (including images) to support your conclusions
- No data, text, or theories by others are presented as if they were the author’s own (“plagiarism”). Proper acknowledgements to other works must be given (this includes material that is closely copied (near verbatim), summarized and/or paraphrased), quotation marks are used for verbatim copying of material, and permissions are secured for material that is copyrighted.

Important note: the journal may use software to screen for plagiarism.

- Consent to submit has been received explicitly from all co-authors, as well as from the responsible authorities - tacitly or explicitly - at the institute/organization where the work has been carried out, **before** the work is submitted.
- Authors whose names appear on the submission have contributed sufficiently to the scientific work and therefore share collective responsibility and accountability for the results.
- Authors are strongly advised to ensure the correct author group, corresponding author, and order of authors at submission. Changes of authorship or in the order of authors are **not** accepted **after** acceptance of a manuscript.
- Adding and/or deleting authors **at revision stage** may be justifiably warranted. A letter must accompany the revised manuscript to explain the role of the added and/or deleted author(s). Further documentation may be required to support your request.
- Requests for addition or removal of authors as a result of authorship disputes after acceptance are honored after formal notification by the institute or independent body and/or when there is agreement between all authors.
- Upon request authors should be prepared to send relevant documentation or data in order to verify the validity of the results. This could be in the form of raw data, samples, records, etc. Sensitive information in the form of confidential proprietary data is excluded.

If there is a suspicion of misconduct, the journal will carry out an investigation following the COPE guidelines. If, after investigation, the allegation seems to raise valid concerns, the accused author will be contacted and given an opportunity to address the issue. If misconduct has been established beyond reasonable doubt, this may result in the Editor-in-Chief's implementation of the following measures, including, but not limited to:

- If the article is still under consideration, it may be rejected and returned to the author.
- If the article has already been published online, depending on the nature and severity of the infraction, either an erratum will be placed with the article or in severe cases complete retraction of the article will occur. The reason must be given in the published erratum or retraction note. Please note that retraction means that the paper is **maintained on the platform**, watermarked "retracted" and explanation for the retraction is provided in a note linked to the watermarked article.
- The author's institution may be informed.

COMPLIANCE WITH ETHICAL STANDARDS

To ensure objectivity and transparency in research and to ensure that accepted principles of ethical and professional conduct have been followed, authors should include information regarding sources of funding, potential conflicts of interest (financial or non-financial), informed consent if the research involved human participants, and a statement on welfare of animals if the research involved animals.

Authors should include the following statements (if applicable) in a separate section entitled "Compliance with Ethical Standards" when submitting a paper:

- Disclosure of potential conflicts of interest
- Research involving Human Participants and/or Animals
- Informed consent

Please note that standards could vary slightly per journal dependent on their peer review policies (i.e. single or double blind peer review) as well as per journal subject discipline. Before submitting your article check the instructions following this section carefully.

The corresponding author should be prepared to collect documentation of compliance with ethical standards and send if requested during peer review or after publication.

The Editors reserve the right to reject manuscripts that do not comply with the above-mentioned guidelines. The author will be held responsible for false statements or failure to fulfill the above-mentioned guidelines.

DISCLOSURE OF POTENTIAL CONFLICTS OF INTEREST

Authors must disclose all relationships or interests that could have direct or potential influence or impart bias on the work. Although an author may not feel there is any conflict, disclosure of relationships and interests provides a more complete and transparent process, leading to an accurate and objective assessment of the work. Awareness of a real or perceived conflicts of interest is a perspective to which the readers are entitled. This is not meant to imply that a financial relationship with an organization that sponsored the research or compensation received for consultancy work is inappropriate. Examples of potential conflicts of interests **that are directly or indirectly related to the research** may include but are not limited to the following:

- Research grants from funding agencies (please give the research funder and the grant number)
- Honoraria for speaking at symposia
- Financial support for attending symposia
- Financial support for educational programs
- Employment or consultation
- Support from a project sponsor
- Position on advisory board or board of directors or other type of management relationships
- Multiple affiliations
- Financial relationships, for example equity ownership or investment interest
- Intellectual property rights (e.g. patents, copyrights and royalties from such rights)
- Holdings of spouse and/or children that may have financial interest in the work

In addition, interests that go beyond financial interests and compensation (non-financial interests) that may be important to readers should be disclosed. These may include but are not limited to personal relationships or competing interests directly or indirectly tied to this research, or professional interests or personal beliefs that may influence your research.

The corresponding author collects the conflict of interest disclosure forms from all authors. In author collaborations where formal agreements for representation allow it, it is sufficient for the corresponding author to sign the disclosure form on behalf of all authors. Examples of forms can be found

- [here:](#)

The corresponding author will include a summary statement in the text of the manuscript in a separate section before the reference list, that reflects what is recorded in the potential conflict of interest disclosure form(s).

See below examples of disclosures:

Funding: This study was funded by X (grant number X).

Conflict of Interest: Author A has received research grants from Company A. Author B has received a speaker honorarium from Company X and owns stock in Company Y. Author C is a member of committee Z.

If no conflict exists, the authors should state:

Conflict of Interest: The authors declare that they have no conflict of interest.

RESEARCH INVOLVING HUMAN PARTICIPANTS AND/OR ANIMALS

1) Statement of human rights

When reporting studies that involve human participants, authors should include a statement that the studies have been approved by the appropriate institutional and/or national research ethics committee and have been performed in accordance with the ethical standards as laid down in the 1964 Declaration of Helsinki and its later amendments or comparable ethical standards.

If doubt exists whether the research was conducted in accordance with the 1964 Helsinki Declaration or comparable standards, the authors must explain the reasons for their approach, and demonstrate that the independent ethics committee or institutional review board explicitly approved the doubtful aspects of the study.

The following statements should be included in the text before the References section:

Ethical approval: “All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.”

For retrospective studies, please add the following sentence:

“For this type of study formal consent is not required.”

2) Statement on the welfare of animals

The welfare of animals used for research must be respected. When reporting experiments on animals, authors should indicate whether the international, national, and/or institutional guidelines for the care and use of animals have been followed, and that the studies have been approved by a research ethics committee at the institution or practice at which the studies were conducted (where such a committee exists).

For studies with animals, the following statement should be included in the text before the References section:

Ethical approval: “All applicable international, national, and/or institutional guidelines for the care and use of animals were followed.”

If applicable (where such a committee exists): “All procedures performed in studies involving animals were in accordance with the ethical standards of the institution or practice at which the studies were conducted.”

If articles do not contain studies with human participants or animals by any of the authors, please select one of the following statements:

“This article does not contain any studies with human participants performed by any of the authors.”

“This article does not contain any studies with animals performed by any of the authors.”

“This article does not contain any studies with human participants or animals performed by any of the authors.”

INFORMED CONSENT

All individuals have individual rights that are not to be infringed. Individual participants in studies have, for example, the right to decide what happens to the (identifiable) personal data gathered, to what they have said during a study or an interview, as well as to any photograph that was taken. Hence it is important that all participants gave their informed consent in writing prior to inclusion in the study. Identifying details (names, dates of birth, identity numbers and other information) of the participants that were studied should not be published in written descriptions, photographs, and genetic profiles unless the information is essential for scientific purposes and the participant (or parent or guardian if the participant is incapable) gave written informed consent for publication. Complete anonymity is difficult to achieve in some cases, and informed consent should be obtained if there is any doubt. For example, masking the eye region in photographs of participants is inadequate protection of anonymity. If identifying characteristics are altered to protect anonymity, such as in genetic profiles, authors should provide assurance that alterations do not distort scientific meaning.

The following statement should be included:

Informed consent: “Informed consent was obtained from all individual participants included in the study.”

If identifying information about participants is available in the article, the following statement should be included:

“Additional informed consent was obtained from all individual participants for whom identifying information is included in this article.”

AFTER ACCEPTANCE

Upon acceptance of your article you will receive a link to the special Author Query Application at Springer’s web page where you can sign the Copyright Transfer Statement online and indicate whether you wish to order OpenChoice and offprints.

Once the Author Query Application has been completed, your article will be processed and you will receive the proofs.

Copyright transfer

Authors will be asked to transfer copyright of the article to the Publisher (or grant the Publisher exclusive publication and dissemination rights). This will ensure the widest possible protection and dissemination of information under copyright laws.

- [Creative Commons Attribution-NonCommercial 4.0 International License](#)

Offprints

Offprints can be ordered by the corresponding author.

Color illustrations

Publication of color illustrations is free of charge.

Proof reading

The purpose of the proof is to check for typesetting or conversion errors and the completeness and accuracy of the text, tables and figures. Substantial changes in content, e.g., new results, corrected values, title and authorship, are not allowed without the approval of the Editor.

After online publication, further changes can only be made in the form of an Erratum, which will be hyperlinked to the article.

Online First

The article will be published online after receipt of the corrected proofs. This is the official first publication citable with the DOI. After release of the printed version, the paper can also be cited by issue and page numbers.

OPEN CHOICE

In addition to the normal publication process (whereby an article is submitted to the journal and access to that article is granted to customers who have purchased a subscription), Springer provides an alternative publishing option: Springer Open Choice. A Springer Open Choice article receives all the benefits of a regular subscription-based article, but in addition is made available publicly through Springer's online platform SpringerLink.

5.6 Comprovante de submissão de artigo à revista Biomarkers



Munique Mendonça Azevedo <azevedo.munique@gmail.com>

[Fwd: BIOMARKERS – Manuscript ID TBMK-2017-0033]

Claudia@ufcspa.edu.br <claudia@ufcspa.edu.br>

24 de novembro de 2017 10:13

Para: azevedo.munique@gmail.com

----- Mensagem Original -----

Assunto: BiOMARKERS - Manuscript ID TBMK-2017-0033

De: "Biomarkers"

<onbehalfof+office-biomarkers+charite.de@manuscriptcentral.com>

Data: Sex, Novembro 24, 2017 10:08 am

Para: claudia@ufcspa.edu.br

Dear :

Your manuscript entitled "Cell-free DNA as an inflammatory marker in post-surgical monitoring of melanoma patients" has been successfully submitted online and is presently being given full consideration for publication in the Biomarkers.

Your manuscript ID is TBMK-2017-0033.

Please mention the above manuscript ID in all future correspondence. If there are any changes in your street address or e-mail address, please log in to Manuscript Central at <https://mc.manuscriptcentral.com/tbmk> and edit your user information as appropriate.

Biomarkers offers an optional FastTrack publication service, with a submission to online publication timeline of 3-5 weeks (subject to a 1 week author revision following initial peer review and prompt turnaround of proofs). The fee for this service is \$850/€625/£550 per published page. If you would like to opt for the FastTrack publication service or would like to know more about it, please contact Temis Vasili (Temis.Vasili@informa.com), as soon as possible.

If you have already chosen the FastTrack service, you will receive further details regarding this shortly.

Please also note that all manuscripts (whether or not they are prioritised) are subject to strict and rigorous peer review. Acceptance for publication is solely dependent on the peer-review outcome.

You can also view the status of your manuscript at any time by checking your Author Centre after logging in to <https://mc.manuscriptcentral.com/tbmk>.

Thank you for submitting your manuscript to Biomarkers.

Sincerely, Biomarkers Editorial Office

5.7 Normas da revista Clinical Biochemistry



Instructions for authors

Thank you for choosing to submit your paper to us. These instructions will ensure we have everything required so your paper can move through peer review, production and publication smoothly. Please take the time to read and follow them as closely as possible, as doing so will ensure your paper matches the journal's requirements. For general guidance on the publication process at Taylor & Francis please visit our [Author Services website](#).

AUTHORSERVICES
Supporting Taylor & Francis authors
SCHOLARONE MANUSCRIPTS™

This journal uses ScholarOne Manuscripts (previously Manuscript Central) to peer review manuscript submissions. Please read the [guide for ScholarOne authors](#) before making a submission. Complete guidelines for preparing and submitting your manuscript to this journal are provided below.

About the journal

Biomarkers is an international, peer reviewed journal, publishing high-quality, original research. Please see the journal's [Aims & Scope](#) for information about its focus and peer-review policy.

Please note that this journal only publishes manuscripts in English.

This journal accepts the following article types: original papers, reviews, technical briefs and letters to the Editor.

Peer review

Taylor & Francis is committed to peer-review integrity and upholding the highest standards of review. Once your paper has been assessed for suitability by the editor, it will then be single blind peer-reviewed by independent, anonymous expert referees. Find out more about [what to expect during peer review](#) and read our guidance on [publishing ethics](#).

Preparing your paper

All authors submitting to medicine, biomedicine, health sciences, allied and public health journals should conform to the Uniform Requirements for Manuscripts Submitted to Biomedical Journals, prepared by the International Committee of Medical Journal Editors (ICMJE).

Structure

Your paper should be compiled in the following order: title page; abstract; keywords; main text: introduction, clinical significance, methods, results, discussion, conclusion; acknowledgments; declaration of interest statement; references; appendices (as appropriate); table(s) with caption(s) (on individual pages); figures; figure captions (as a list).

Word count

Please include a word count for your paper. A typical paper for this journal should not exceed 12000 words; inclusive of tables, references, figure captions.

Style guidelines

Please refer to these style guidelines when preparing your paper, rather than any published articles or a sample copy.

Please use British spelling consistently throughout your manuscript.

Please use single quotation marks, except where 'a quotation is "within" a quotation'. Please note that long quotations should be indented without quotation marks.

Formatting and templates

Papers may be submitted in any standard format, including Word and LaTeX. Figures should be saved separately from the text. To assist you in preparing your paper, we provide formatting template(s).

Word templates are available for this journal. Please save the template to your hard drive, ready for use.

If you are not able to use the templates via the links (or if you have any other template queries) please contact authortemplate@tandf.co.uk

References

Please use this reference guide when preparing your paper. An EndNote output style is also available to assist you.

Checklist: what to include

1. **Author details.** Please ensure everyone meeting the International Committee of Medical Journal Editors (ICJME) [requirements for authorship](#) is included as an author of your paper. Please include all authors' full names, affiliations, postal addresses, telephone numbers and email addresses on the cover page. Where available, please also include [ORCiDs](#) and social media handles (Facebook, Twitter or LinkedIn). One author will need to be identified as the corresponding author, with their email address normally displayed in the article PDF (depending on the journal) and the online article. Authors' affiliations are the affiliations where the research was conducted. If any of the named co-authors moves affiliation during the peer-review process, the new affiliation can be given as a footnote. Please note that no changes to affiliation can be made after your paper is accepted. [Read more on authorship](#).
2. A structured **abstract** of no more than 200 words. A structured abstract should cover (in the following order) the *purpose* of the article, its *materials and methods* (the experimental system and procedures used), the *results* and *conclusions*. Read tips on [writing your abstract](#).
3. You can opt to include a **video abstract** with your article. [Find out how these can help your work reach a wider audience, and what to think about when filming](#).
4. 5-10 **keywords**. Read [making your article more discoverable](#), including information on choosing a title and search engine optimization **keywords**. Read [making your article more discoverable](#), including information on choosing a title and search engine optimization.
5. **Clinical significance.** This is a mandatory section and should include a bulleted list of no more than 100 words detailing the clinical significance of the manuscript.
6. **Funding details.** Please supply all details required by your funding and grant-awarding bodies as follows: *For single agency grants:* This work was supported by the under Grant . *For multiple agency grants:* This work was supported by the under Grant ; under Grant ; and under Grant .
7. **Disclosure statement.** This is to acknowledge any financial interest or benefit that has arisen from the direct applications of your research. [Further guidance on what is a conflict of interest and how to disclose it](#).
8. **Geolocation information.** Submitting a geolocation information section, as a separate paragraph before your acknowledgements, means we can index your paper's study area accurately in JournalMap's geographic literature database and make your article more discoverable to others. [More information](#).
9. **Supplemental online material.** Supplemental material can be a video, dataset, fileset, sound file or anything which supports (and is pertinent to) your paper. We publish supplemental material online via Figshare. Find out more about [supplemental material and how to submit it with your article](#).
10. **Figures.** Figures should be high quality (1200 dpi for line art, 600 dpi for grayscale and 300 dpi for colour). Figures should be saved as TIFF, PostScript or EPS files.
11. **Tables.** Tables should present new information rather than duplicating what is in the text. Readers should be able to interpret the table without reference to the text. Please supply editable files.

12. **Equations.** If you are submitting your manuscript as a Word document, please ensure that equations are editable. More information about [mathematical symbols and equations](#).
13. **Units.** Please use [SI units](#) (non-italicized).

Using third-party material in your paper

You must obtain the necessary permission to reuse third-party material in your article. The use of short extracts of text and some other types of material is usually permitted, on a limited basis, for the purposes of criticism and review without securing formal permission. If you wish to include any material in your paper for which you do not hold copyright, and which is not covered by this informal agreement, you will need to obtain written permission from the copyright owner prior to submission. More information on [requesting permission to reproduce work\(s\) under copyright](#).

Disclosure statement

Please include a disclosure of interest statement, using the subheading "Disclosure of interest." If you have no interests to declare, please state this (suggested wording: *The authors report no conflicts of interest*). For all NIH/Wellcome-funded papers, the grant number(s) must be included in the disclosure of interest statement. [Read more on declaring conflicts of interest](#).

Clinical Trials Registry

In order to be published in a Taylor & Francis journal, all clinical trials must have been registered in a public repository at the beginning of the research process (prior to patient enrolment). Trial registration numbers should be included in the abstract, with full details in the methods section. The registry should be publicly accessible (at no charge), open to all prospective registrants, and managed by a not-for-profit organization. For a list of registries that meet these requirements, please visit the [WHO International Clinical Trials Registry Platform \(ICTRP\)](#). The registration of all clinical trials facilitates the sharing of information among clinicians, researchers, and patients, enhances public confidence in research, and is in accordance with the [ICMJE guidelines](#).

Complying with ethics of experimentation

Please ensure that all research reported in submitted papers has been conducted in an ethical and responsible manner, and is in full compliance with all relevant codes of experimentation and legislation. All papers which report *in vivo* experiments or clinical trials on humans or animals must include a written statement in the Methods section. This should explain that all work was conducted with the formal approval of the local human subject or animal care committees (institutional and national), and that clinical trials have been registered as legislation requires. Authors who do not have formal ethics review

committees should include a statement that their study follows the principles of the Declaration of Helsinki.

Consent

All authors are required to follow the ICMJE requirements on privacy and informed consent from patients and study participants. Please confirm that any patient, service user, or participant (or that person's parent or legal guardian) in any research, experiment, or clinical trial described in your paper has given written consent to the inclusion of material pertaining to themselves, that they acknowledge that they cannot be identified via the paper; and that you have fully anonymized them. Where someone is deceased, please ensure you have written consent from the family or estate.

Health and safety

Please confirm that all mandatory laboratory health and safety procedures have been complied with in the course of conducting any experimental work reported in your paper. Please ensure your paper contains all appropriate warnings on any hazards that may be involved in carrying out the experiments or procedures you have described, or that may be involved in instructions, materials, or formulae.

Please include all relevant safety precautions; and cite any accepted standard or code of practice. Authors working in animal science may find it useful to consult the International Association of Veterinary Editors' Consensus Author Guidelines on Animal Ethics and Welfare and Guidelines for the Treatment of Animals in Behavioural Research and Teaching. When a product has not yet been approved by an appropriate regulatory body for the use described in your paper, please specify this, or that the product is still investigational.

Submitting your paper

This journal uses ScholarOne to manage the peer-review process. If you haven't submitted a paper to this journal before, you will need to create an account in the submission centre. Please read the guidelines above and then submit your paper in the relevant Author Centre, where you will find user guides and a helpdesk.

If you are submitting in LaTeX, please convert the files to PDF beforehand (you will also need to upload your LaTeX source files with the PDF).

Please note that *Biomarkers* uses Crossref™ to screen papers for unoriginal material. By submitting your paper to *Biomarkers* you are agreeing to originality checks during the peer-review and production processes.

On acceptance, we recommend that you keep a copy of your Accepted Manuscript. Find out more about sharing your work.

Publication charges

There are no submission fees or page charges for this journal.

Colour figures will be reproduced in colour in your online article free of charge. If it is necessary for the figures to be reproduced in colour in the print version, a charge will apply.

Charges for colour figures in print are £250 per figure (\$395 US Dollars; \$385 Australian Dollars; €315). For more than 4 colour figures, figures 5 and above will be charged at £50 per figure (\$80 US Dollars; \$75 Australian Dollars; €63). Depending on your location, these charges may be subject to local taxes.

Copyright options

Copyright allows you to protect your original material, and stop others from using your work without your permission. Taylor & Francis offers a number of different license and reuse options, including Creative Commons licenses when publishing open access. [Read more on publishing agreements.](#)

Complying with funding agencies

We will deposit all National Institutes of Health or Wellcome Trust-funded papers into PubMedCentral on behalf of authors, meeting the requirements of their respective open access (OA) policies. If this applies to you, please tell our production team when you receive your article proofs, so we can do this for you. Check funders' OA policy mandates [here](#). Find out more about [sharing your work](#).

Open access

This journal gives authors the option to publish open access via our [Open Select publishing program](#), making it free to access online immediately on publication. Many funders mandate publishing your research open access; you can check [open access funder policies and mandates here](#).

Taylor & Francis Open Select gives you, your institution or funder the option of paying an article publishing charge (APC) to make an article open access. Please contact openaccess@tandf.co.uk if you would like to find out more, or go to our [Author Services website](#).

For more information on license options, embargo periods and APCs for this journal please go [here](#).

Accepted Manuscripts Online (AMO)

This journal publishes manuscripts online as rapidly as possible, as a PDF of the final, accepted (but unedited and uncorrected) paper. This is clearly

identified as an unedited manuscript and is referred to as the Accepted Manuscript Online (AMO). No changes will be made to the content of the original paper for the AMO version but, after copy-editing, typesetting, and review of the resulting proof, the final corrected version (the Version of Record [VoR]), will be published, replacing the AMO version.

The VoR is the article version that will appear in an issue of the journal. Both the AMO version and VoR can be cited using the same DOI (digital object identifier). To ensure rapid publication, we ask you to return your signed publishing agreement as quickly as possible, and return corrections within 48 hours of receiving your proofs.

My Authored Works

On publication, you will be able to view, download and check your article's metrics (downloads, citations and Altmetric data) via My Authored Works on Taylor & Francis Online. This is where you can access every article you have published with us, as well as your free eprints link, so you can quickly and easily share your work with friends and colleagues.

We are committed to promoting and increasing the visibility of your article. Here are some tips and ideas on how you can work with us to promote your research.

Article reprints

For enquiries about reprints, please contact the Taylor & Francis Author Services team at reprints@tandf.co.uk. To order a copy of the issue containing your article, please contact our Customer Services team at Adhoc@tandf.co.uk.

Sponsored supplements

This journal occasionally publishes sponsored supplements alongside its schedule of regular issues. Like our articles, all our supplements are expected to meet the same editorial standards as the journal and are subject to approval of the Editor prior to acceptance. They are indexed by the journal and offer targeted, cost-effective and high-impact reach. For more information and to discuss your requirements, please contact charles.whalley@tandf.co.uk.

Queries

Should you have any queries, please visit our Author Services website or contact us at authorqueries@tandf.co.uk.

Updated April 2016

5.8 Comprovante de submissão de artigo à revista The American Journal of Dermatopathology



Munique Mendonça <mAuniquepm@gmail.com>

[Fwd: AJD Submission Confirmation for Gender and age-specific association between manganese superoxide dismutase polymorphism (Ala16Val-SOD2) and melanoma risk]

claudia@ufcspa.edu.br <claudia@ufcspa.edu.br>

24 de novembro de 2017
11:42

Para: muniquepm@gmail.com

Your submission titled "Gender and age-specific association between manganese superoxide dismutase polymorphism (Ala16Val-SOD2) and melanoma risk" has been received by the journal editorial office.

You will be able to check on the progress of your paper by logging on to Editorial Manager as an author.

<http://ajd.edmgr.com/>

Your username is: MAzevedo

<http://ajd.edmgr.com/l.asp?i=72128&l=RTU5X0L7>

Additionally, you may view the Additional Information questions to obtain the copyright information by clicking here: Additional Information

Your manuscript will be given a reference number once an Editor has been assigned.

Thank you for your interest in the American Journal of Dermatopathology.

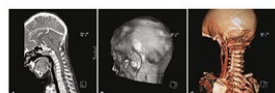
Kind Regards,

The American Journal of Dermatopathology

5.9 Normas da revista The American Journal of Dermatopathology



The American Journal of Dermatopathology



The Journal of the International Society of Dermatopathology

Editor-in-Chief

Omar P. Sangüeza, MD



Preparation of Manuscript

Manuscripts that do not adhere to the following instructions will be returned to the corresponding author for technical revision before undergoing peer review.

Title page: Include on the title page (a) complete manuscript title; (b) authors' full names, highest academic degrees, and affiliations; (c) name and address for correspondence, including fax number, telephone number, and e-mail address; (d) address for reprints if different from that of corresponding author; and (e) sources of support that require acknowledgment.

The title page must also include disclosure of funding received for this work from any of the following organizations: National Institutes of Health (NIH); Wellcome Trust; Howard Hughes Medical Institute (HHMI); and other(s).

Unstructured abstract and key words: Limit the abstract to 250 words. It must be factual and comprehensive. Limit the use of abbreviations and acronyms, and avoid general statements (eg, "the significance of the results is discussed"). List three to five key words or phrases.

Text: Organize the manuscript into four main headings: Introduction, Materials and Methods, Results, and Discussion. Define abbreviations at first mention in text and in each table and figure. If a brand name is cited, supply the manufacturer's name and address (city and state/country). Acknowledge all forms of support, including pharmaceutical and industry support, in an Acknowledgments paragraph.

Abbreviations: For a list of standard abbreviations, consult the *Council of Biology Editors Style Guide* (available from the Council of Science Editors, 9650 Rockville Pike, Bethesda, MD 20814) or other standard sources. Write out the full term for each abbreviation at its first use unless it is a standard unit of measure.

References: The authors are responsible for the accuracy of the references. Key the references (double-spaced) at the end of the manuscript. Cite the references in text in the order of appearance. Cite unpublished data—such as papers submitted but not yet accepted for publication and personal communications, including e-mail communications—in parentheses in the text. If there are more than three authors, name only the first three authors and then use et al. Refer to the *List of Journals Indexed in Index Medicus* for abbreviations of journal names, or access the list at <http://www.nlm.nih.gov/tsd/serials/lji.html>. Sample references are given below:

Journal article

1. Farkas LG, Tompson B, Phillips JH, et al. Comparison of anthropometric and cephalometric measurements of the adult face. *J Craniofacial Surg.* 1999;10:18–25.

Book chapter

2. Todd VR. Visual information analysis: frame of reference for visual perception. In: Kramer P, Hinojosa J, eds. *Frames of Reference for Pediatric Occupational Therapy*. Philadelphia, PA: Lippincott Williams & Wilkins; 1999:205–256.

Entire book

3. Kellman RM, Marentette LJ. *Atlas of Craniomaxillofacial Fixation*. Philadelphia, PA: Lippincott Williams & Wilkins; 1999.

Software

4. *Epi Info* [computer program]. Version 6. Atlanta, GA: Centers for Disease Control and Prevention; 1994.

Online journals

5. Friedman SA. Preeclampsia: a review of the role of prostaglandins. *Obstet Gynecol* [serial online]. January 1988;71:22-37. Available from: BRS Information Technologies, McLean, VA. Accessed December 15, 1990.

Database

6. CANCERNET-PDQ [database online]. Bethesda, MD: National Cancer Institute; 1996. Updated March 29, 1996.

World Wide Web

7. Gostin LO. Drug use and HIV/AIDS [JAMA HIV/AIDS Web site]. June 1, 1996. Available at: <http://www.ama-assn.org/special/hiv/ethics>. Accessed June 26, 1997.

Figure & Artwork:

A) Creating Digital Artwork

1. Learn about the publication requirements for Digital Artwork: <http://links.lww.com/ES/A42>

2. Create, Scan and Save your artwork and compare your final figure to the Digital Artwork Guideline Checklist (below).
3. Upload each figure to Editorial Manager in conjunction with your manuscript text and tables.

B) Digital Artwork Guideline Checklist

Here are the basics to have in place before submitting your digital artwork:

- Artwork should be saved as TIFF, EPS, or MS Office (DOC, PPT, XLS) files. High resolution PDF files are also acceptable.
- Crop out any white or black space surrounding the image.
- Diagrams, drawings, graphs, and other line art must be vector or saved at a resolution of at least 1200 dpi. If created in an MS Office program, send the native (DOC, PPT, XLS) file.
- Photographs, radiographs and other halftone images must be saved at a resolution of at least 300 dpi.
- Photographs and radiographs with text must be saved as postscript or at a resolution of at least 600 dpi.
- Each figure must be saved and submitted as a separate file. Figures should not be embedded in the manuscript text file.

Remember:

- Cite figures consecutively in your manuscript.
- Number figures in the figure legend in the order in which they are discussed.
- Upload figures consecutively to the Editorial Manager web site and enter figure numbers consecutively in the Description field when uploading the files.
- When submitting single-column width figures, the editor strongly recommends that these be oriented vertically to allow larger image size. The orientation of the figures in the electronic submission will be interpreted by the production staff as the desired orientation in the final publication. Therefore, authors should submit single-citation figures oriented vertically if at all possible.
- The publisher will charge \$50 for each image file that must be repurposed from hard copy.

Figure legends: Legends must be submitted for all figures. They should be brief and specific, and they should appear on a separate manuscript page after the references. Use scale markers in the image for electron micrographs, and indicate the type of stain used.

Color figures: The journal accepts for publication color figures that will enhance an article. Beginning with the 2009 volume, there is no additional charge for color figures.

Tables: Create tables using the table creating and editing feature of your word processing software (eg, Word, WordPerfect). Do not use Excel or comparable spreadsheet programs. Group all tables at the end of the manuscript, or supply them together in a separate file. Cite tables consecutively in the text, and number them in that order. Key each on a separate sheet, and include the table title, appropriate column heads, and explanatory legends (including definitions of any abbreviations used). **Do not embed tables within the body of the manuscript.** They should be self-explanatory and should supplement, rather than duplicate, the material in the text.